

It Takes Three to Tango: The Length of the Oligothiophene Chain Determines the Nature of the Long-Lived Excited State and the Resulting Photocytotoxicity of a Ruthenium (II) Photodrug

Avinash Chettri,^[a, b] John A. Roque, III,^[c, d] Kilian R. A. Schneider,^[a, b] Houston D. Cole,^[c] Colin G. Cameron,^[c] Sherri A. McFarland,^{*[c]} and Benjamin Dietzek^{*[a, b]}

TLD1433 is the first Ru(II) complex to be tested as a photodynamic therapy agent in a clinical trial. In this contribution we study TLD1433 in the context of structurally-related Ru(II)imidozo[4,5-f][1,10]phenanthroline (ip) complexes appended with thiophene rings to decipher the unique photophysical properties which are associated with increasing oligothiophene chain length. Substitution of the ip ligand with ter- or quaterthiophene changes the nature of the long-lived triplet state from metal-to-ligand charge-transfer to ${}^{3}\pi\pi^{*}$ character. The addition of the third thiophene thus presents a critical juncture which not only determines the photophysics of the complex but most importantly its capacity for ${}^{1}O_{2}$ generation and hence the potential of the complex to be used as a photocytotoxic agent.

Recent developments highlight Ru(II) polypyridyl complexes with π -expanded ligands as a promising class of new compounds for photodynamic therapy (PDT).^[1-11] By extending the pyridyl ligands with organic chromophores, low-lying intraligand (IL) excited states become accessible, and these appear to

-	
[a]	A. Chettri, K. R. A. Schneider, Prof. Dr. B. Dietzek
	Department Functional Interfaces Department
	Leibniz Institute of Photonic Technology Jena
	Albert-Einstein-Straße 9, 07745 Jena (Germany)
	E-mail: benjamin.dietzek@leibniz-ipht.de
[b]	A. Chettri, K. R. A. Schneider, Prof. Dr. B. Dietzek
	Institute of Physical Chemistry
	Friedrich-Schiller University Jena
	Helmholtzweg 4, 07743 Jena (Germany)
	J. A. Roque, III, H. D. Cole, Dr. C. G. Cameron, Prof. Dr. S. A. McFarland
	Department of Chemistry and Biochemistry
	The University of Texas Arlington
	Arlington, TX 76019 (USA)
	E-mail: sherri.macfarland@uta.edu
	J. A. Roque, III
	Department of Chemistry and Biochemistry
	The University of North Carolina at Greensboro
	Greensboro, North Carolina 27402 (USA)
~~~~	Supporting information for this article is available on the WWW under
	https://doi.org/10.1002/cptc.202000283
pecial dection	An invited contribution to the "GDCh and ChemPhotoChem: 5-Year
meetion	Anniversary" Special Collection.
$\mathbf{c}$	© 2021 The Authors. ChemPhotoChem published by Wiley-VCH GmbH.
01	

© 2021 The Authors. ChemPhotoChem published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution Non-Commercial NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. be crucial to the photophysical function of these systems.^[12] Our TLD1433 (**Ru-ip-3T** in this manuscript, Figure 1) is a compound of this type, having three appended thiophene rings, and has the distinction of being the first Ru(II)-based PDT agent ever to enter a human clinical trial;^[1,10,13] **Ru-ip-3T** is currently being tested in a Phase II PDT trial for non-invasive bladder cancer (ClinicalTrials.gov identifier: NCT03945162).^[10]

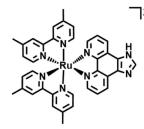
In vitro studies have been previously conducted on the Ruip-nT series of compounds, where n indicates the number of appended thiophene rings attached to an imidazo[4,5-f][1,10] phenanthroline (ip) ligand (Figure 1). Visible light illumination of SK-MEL-28 cancer cells treated with the compounds in the series led to increased photocytotoxicity with increasing n. The light EC₅₀ values (effective concentration to reduce cell viability by 50 %) were 0.72  $\mu M,$  0.26  $\mu M,$   $1.9 \times 10^{-4}$   $\mu M$  and  $2.8 \times 10^{-9}$   $\mu M$ for Ru-ip-1T to Ru-ip-4T, respectively (Figure 2d)^[1] and the corresponding PI values (ratio of dark to light EC₅₀) were 225, 434,  $7.2 \times 10^5$  and  $45 \times 10^9$  respectively. While these previous findings demonstrate a clear correlation between the length of the thiophene chain and the in vitro phototherapeutic effects, herein we present the key photophysical properties of the compounds in the Ru-ip-nT series that could be responsible for the observed photocytotoxicity.

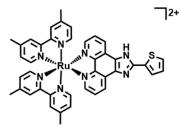
The electronic absorption spectra of the compounds within the **Ru-ip-nT** series (Figure 2a) are characterized by IL transitions localized to the dmb and phen portion of the ip ligand below 300 nm as well as a broad metal-to-ligand charge transfer (MLCT) band in the 420–550 nm range, consistent with the spectrum of the parent [Ru(dmb)₃]²⁺ complex.^[14,15] An additional band centered near 370 nm is visible in **Ru-ip-2T** that shifts to longer wavelengths in **Ru-ip-3T** and **Ru-ip-4T**. This band corresponds to the  $\pi\pi^*$  ¹IL transition associated with the oligothiophene.^[16] These oligothiophene-localized transitions have been computed to have substantial charge transfer character and are best described as intraligand charge transfer (ILCT) states, but are referred to herein more generally as IL.^[17,18]

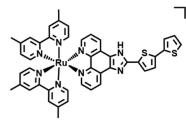
The steady-state emission spectra of the complexes show a single structureless emission band centered at 625 nm (Figure 2b), suggesting a similar emissive ³MLCT state for all complexes in the **Ru-ip-nT** family.^[19] The emission quantum yields ( $\Phi_{em.}$ ) in deaerated water (five freeze pump thaw cycles under a nitrogen atmosphere) drop with increasing *n*, from 6% for **Ru-ip-1T** to below 0.1% for **Ru-ip-3T** (Table 1). Emission from **Ru-ip-4T** is barely detectable (Figure 2d). The emission

```
ChemPhotoChem 2021, 5, 421-425
```





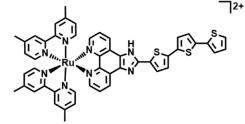




Ru-ip-0T



Ru-ip-2T



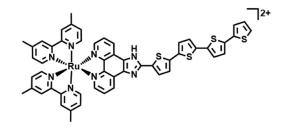




Figure 1. Chemical structures of Ru-ip-nT complexes.

Ru-ip-3T

Complex	τ _{em.} ^[d] Aerated/Deaerated [μs]/[μs]	τ _{τΑ.} ^[d] Aerated/Deaerated [μs]/[μs]	$\Phi_{em.^{[d]}}$ Aerated/Deaerated	$\Phi_{\!\Delta}{}^{[e]}$
Ru-ip-0T	0.4/0.7	0.4/0.7	0.038/0.05	0.68
Ru-ip-1T	0.4/0.8	0.4/0.7	0.044/0.06	0.61
Ru-ip-2T	0.3; 1[b]/0.6; 11[b]	1.4/14	0.006/0.04	0.71
Ru-ip-3T	0.3/0.6	1.4/48	$-[c]/4.8 \times 10^{-4}$	0.77
Ru-ip-4T	0.3/0.6	1.4/29	$-[c]/8.1 \times 10^{-4}$	0.81

[a]  $\tau_{em}$ : emission lifetime;  $\tau_{TA}$ : recovery lifetime;  $\Phi_{em}$ : quantum yield of emission;  $\Phi_A$ : singlet oxygen quantum yield. Singlet oxygen was determined spectroscopically by monitoring its emission centered at 1275 nm in aerated MeCN. [**Ru(bpy)**₃][**PF**₆]₂ is used as the reference obtaining  $\Phi_{em}^{[27]}$  and  $\Phi_A^{[28]}$  [b] A biexponential decay of the respective signal is observed. [c] Emission is too low to obtain a reasonable estimate of the emission quantum yield. [d] Solvent used is H₂O. [e] Solvent used is MeCN.

quantum yields for **Ru-ip-3T** and **Ru-ip-4T** are extremely small and do not notably increase upon deaeration of the solvent. This suggests that collisional deactivation by  ${}^{3}O_{2}$  is not the prime quenching process of the  ${}^{3}MLCT$  state in the complexes.

The excitation spectra recorded for **Ru-ip-0T**, **Ru-ip-1T** and **Ru-ip-2T** using the emission signal at 640 nm resemble the respective absorption spectra (Figure 2a, b), suggesting that the emissive ³MLCT state is populated regardless of whether the initially populated states are ¹MLCT or ¹IL (i.e.,  $\pi\pi^*$ ). The absence of excitation signals that would correspond to broad ¹IL transitions in the UV-vis spectra at 420 nm and 440 nm for **Ru-ip-3T** and **Ru-ip-4T**, respectively, indicates that excitation of the ¹IL state does not populate the emissive ³MLCT state.

In addition to absorption and emission characteristics, the singlet oxygen quantum yields ( $\Phi_{\Delta}$ ) were also measured. Values for  $\Phi_{\Delta}$  increase from 0.61 for **Ru-ip-1T** to 0.81 for **Ru-ip-4T** (Table 1). This trend is consistent with the trend for photocytotoxicity as reflected in the PI values that increase with *n* (see Figure 2d).⁽¹⁾ This positive correlation between singlet

oxygen quantum yields and photocytotoxicity suggests that singlet oxygen is involved in their mode of action.^[20-22] Enhanced sensitivity toward oxygen is normally reflected as a drastic decrease in triplet excited state lifetime in the presence of oxygen.^[8] The relative changes of excited state lifetimes for **Ru-ip-nT** upon exposure to oxygen will be discussed in the next section.

The photophysical properties of the longer-lived excited states, from which singlet oxygen sensitization occurs, were probed by transient absorption (TA) spectroscopy yielding excited-state absorption spectra and characteristic time-constants for ground state recovery ( $\tau_{TA}$ ) and emission decay ( $\tau_{em}$ ). Table 1 summarizes these key characteristics of the long-lived excited states in aerated and deaerated water. The emission and TA lifetimes of **Ru-ip-0T** and **Ru-ip-1T** are the same, 0.4  $\mu$ s, indicative of a single excited state being depopulated (Figure 3a). The fact that deaeration lengthens the lifetime by only two-fold suggests that this state is only weakly quenched by oxygen. **Ru-ip-2T** behaves somewhat differently than **Ru-ip-0T** 

Communications doi.org/10.1002/cptc.202000283



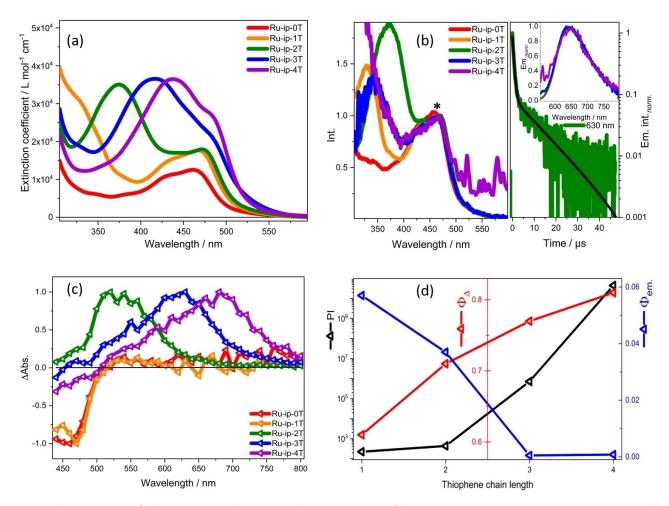


Figure 2. (a) Absorption spectra for the **Ru-ip-nT** complexes in water. (b) Excitation spectra of the **Ru-ip-nT** complexes in water at 640 nm emission normalized to the MLCT band at 480 nm (*, left); bi-exponential emission decay of **Ru-ip-2T** with lifetimes of 0.6  $\mu$ s and 11  $\mu$ s at 630 nm (right); Inset: Emission spectra of **Ru-ip-nT** complexes normalized to their respective maxima. (c) Nanosecond TA spectra of **Ru-ip-nT** complexes in aerated water normalized to their respective maxima at  $\lambda_{ex}$ : 410 nm at 300 ns. (d) Relationship between the PI values (black y-axis), singlet-oxygen quantum yield  $\Phi_{\Delta}$  (red y-axis) in aerated MeCN (excitation at 450 nm) and fluorescence quantum yield  $\Phi_{em}$ . (blue y-axis) in deaerated water (excitation at 450 nm) with increasing thiophene chain length (x-axis).^[1] The PI is defined as the ratio of EC_{50 dark} to EC_{50 visible light} values in SK-MEL-28 cells.

and Ru-ip-1T. In aerated aqueous solutions,  $\tau_{em}$  and  $\tau_{TA}$  for Ruip-2T are 1  $\mu s$  and 1.4  $\mu s,$  respectively. In deaerated solutions, however, the excited state(s) was much longer lived, and the emission decay was biexponential ( $\tau_{em} = 0.6$ ; 11 µs,  $\tau_{TA} = 14$  µs). An excited state lifetime that is ten-fold (or more) longer than the typical 1  $\mu s$  lifetime of  $^3\text{MLCT}$  states in Ru(II) polypyridyl complexes and is very sensitive to O2 is consistent with the involvement of an ³IL state.^[23,24] The second thiophene lowers the energy of this state sufficiently that the emissive ³MLCT and non-emissive ³IL states are in energetic proximity (Figure 3b). The biexponential emission decay, where the longer component matches the decay of the non-emissive state by TA, suggests that the shorter 0.6 µs component is due to prompt ³MLCT emission and the longer 11 µs lifetime corresponds to delayed emission from the ³MLCT state resulting from population of the ³MLCT from the nearly isoenergetic ³IL state (Figure 3b).^[24]

For **Ru-ip-3T** and **Ru-ip-4T**, the aerated and deaerated emission lifetimes were monoexponential and relatively short ( $\tau_{em} \approx 0.4 \ \mu s$ ) as observed for **Ru-ip-0T** and **Ru-ip-1T**, indicating

that the emissive ³MLCT state cannot be populated from the ³IL when n=3 or 4. Ru-ip-3T and Ru-ip-4T are barely emissive, with emission quantum yields too small to be calculated in the aerated solutions and vanishingly small in deaerated conditions. These observations point to the fact that a significant fraction of the Ru-ip-3T and Ru-ip-4T excited states must deactivate via non-emissive ³IL states. The aerated TA lifetimes for Ru-ip-3T and Ru-ip-4T were identical to that of Ru-ip-2T, consistent with efficient quenching of the ³IL states by O₂ and corroborated by all three being very good ¹O₂ sensitizers. The values for  $\tau_{TA}$  in deaerated water were considerably longer, 48 µs and 29 µs for Ru-ip-3T and Ru-ip-4T, respectively. This is attributed to the ³IL state of both Ru-ip-3T and Ru-ip-4T being sufficiently lower in energy than the emissive ³MLCT state (Figure 3c), which remains unchanged throughout the series (evidenced by a constant emission maximum). The triplet is thereby trapped in the longer-lived ³IL state, unable to populate the ³MLCT, which is significantly uphill in energy. The ³IL relaxes much slower due to the reduced intersystem crossing rate in organic chromophores compared to transition

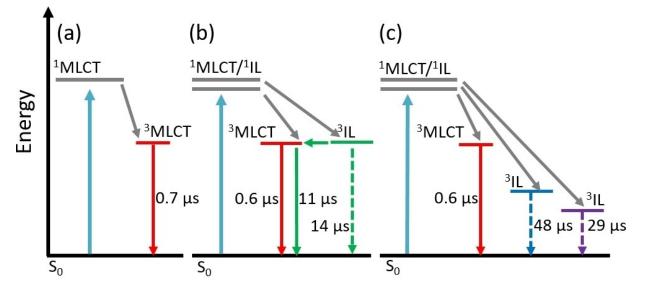


Figure 3. Jablonski diagrams depicting the photophysical models that describe the nanosecond-microsecond excited state dynamics of Ru-ip-nT complexes with 410 nm excitation in deaerated water: (a) Ru-ip-0T, Ru-ip-1T; (b) Ru-ip-2T; (c) Ru-ip-3T, Ru-ip-4T. [Note: grey arrows represent processes occurring faster than the investigated timescale.]

metal complexes, in which the heavy metal ion increases spinorbit coupling. The lifetime of the ³IL state for **Ru-ip-4T** may be shorter than the corresponding lifetime for **Ru-ip-3T** due to the energy gap law.^[25] However, additional relaxation pathways cannot be excluded without further investigation.

The change in the nature of the long-lived photobiologically active excited states is also supported by the spectral shape of the ns TA spectra within the Ru-ip-nT series. The TA spectra of  $\ensuremath{\text{Ru-ip-0T}}$  and  $\ensuremath{\text{Ru-ip-1T}}$  show a ground-state bleach below 510 nm plus a very weak, unstructured excited-state absorption extending from 510 to 800 nm (Figure 2c). This signature is typical for the ³MLCT states of Ru(II) polypyridyl complexes that lack  $\pi$ -extended ligands and thus low-lying ³IL states.^[26] The TA spectra change substantially for Ru-ip-2T through Ru-ip-4T, which have very strong and much more structured excited state absorptions with maxima at 550, 630, and 680 nm, respectively. This systematic red-shift of the excited state absorption with increasing n, by over 100 nm, is characteristic of oligothiophene-based ³IL states.^[5,17] Thus, the ns TA spectra indicate that the character of the long-lived state changes from ³MLCT for Ru-ip-0T and Ru-ip-1T to predominantly ³IL with additional thiophene rings.

This study highlights the photophysical properties of the lowest lyingexcited states in a series of Ru(II) complexes, which underlie their previously reported photocytotoxicity. The results show that the spectroscopic signatures of the long-lived excited states and the biological activity in this series of complexes are determined by the energy of the non-emissive ³IL state relative to the energy of the emissive ³MLCT state. This energy depends on the length of the oligothiophene chain and determines the TA absorption and emission lifetimes, emission quantum yields and light  $EC_{50}$  values. In **Ru-ip-OT** and **Ru-ip-IT** the energy of the ³IL is too high and the state does not contribute to the ns-/µs-photophysics and photobiology of the

complexes. **Ru-ip-2T** represents the situation where the ³MLCT and ³IL states are close in energy. In this case, the ³IL state serves as an excited state reservoir for populating the ³MLCT state and results in delayed ³MLCT emission. In **Ru-ip-3T** and **Ru-ip-4T** the ³IL state is the lowest-lying triplet and plays a predominant role in the excited state relaxation. The oxygensensitive ³IL state appears to be responsible for the increased photocytotoxicity of these complexes. This is manifested in a sharp increase in the photocytotoxicity and efficiency of singlet oxygen sensitization upon increasing the length of the oligothiophene chain.

## **Experimental Section**

All samples were dissolved in the respective solvent and measured in a 1 cm quartz cell. All solvents were deaerated by freeze pump thaw cycles for five times using nitrogen as inert gas. For measuring  $\Phi_{em.}$  and  $\Phi_{\Delta}$  samples with an OD of about 0.05 were used. Quantum yields were calculated according to the equation  $\Phi_s = \Phi_r \cdot \frac{l_s}{l_r} \cdot \frac{OD_r}{OD_r} \cdot \frac{\eta_s^2}{\eta_r^2}$  where  $\Phi_s$  and  $\Phi_r$  are the quantum yields of the sample and a reference, respectively.  $I_s$  and  $I_r$  are the integrated emission intensity of the sample and the references, while  $OD_r$  and  $OD_s$  are the optical densities of the sample and the reference at the excitation wavelength.  $\eta_r$  and  $\eta_s$  refer to the refractive indices of the media in the sample and the reference. Since the same medium is used for measuring the sample and the reference  $\frac{\eta_r^2}{\eta_r^2}$ equals 1.

UV-vis absorption measurements utilized a Jasco V-670 spectrophotometer and emission measurements were carried out on a FLS980 spectrophotometer (Edinburgh Instruments). ¹O₂ emission was detected using a FLS980 spectrophotometer equipped with a NIR detector with parameters upon excitation at 450 nm.

Nanosecond transient absorption measurements used a 10 Hz Nd: YAG laser (Surelite) combined with an OPO for excitation. The Transient data was recorded by a commercial detection system



(Pascher Instruments AB) with a time resolution of 10 ns. The OD of the samples at the excitation wavelength was ~0.25 in a 1-cm cuvette. The integrity of the samples after nanosecond measurements were checked by measuring absorption spectra before and after the measurement. No degradation was observed. Time resolved emission measurements utilized time correlation single photon counting (TCSPC, Becker & Hickl GmbH) upon excitation at 390 nm.

## Acknowledgements

We are grateful to the German Science Foundation (grant No. 395358570), the Carl Zeiss Foundation. We also thank the National Cancer Institute (NCI) of the National Institutes of Health (NIH) (Award R01CA222227) for partial support of this work. The content in this article is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. Open access funding enabled and organized by Projekt DEAL.

## **Conflict of Interest**

S. A. M. has a potential research conflict of interest due to a financial interest with Theralase Technologies, Inc. and Photo-Dynamic, Inc. A management plan has been created to preserve objectivity in research in accordance with UTA policy.

**Keywords:** excited states • photodynamic therapy • ruthenium • singlet oxygen • TLD1433 • triplet states

- S. Monro, K. L. Colón, H. Yin, J. Roque, P. Konda, S. Gujar, R. P. Thummel, L. Lilge, C. G. Cameron, S. A. McFarland, *Chem. Rev.* 2019, *119*, 797–828.
- [2] M. Stephenson, C. Reichardt, M. Pinto, M. Wa, T. Sainuddin, G. Shi, H. Yin, S. Monro, E. Sampson, B. Dietzek, S. A. Mcfarland, J. Phys. Chem. A 2014, 118, 10507–10521.
- [3] S. Chakrabortty, B. K. Agrawalla, A. Stumper, N. M. Vegi, S. Fischer, C. Reichardt, M. Kögler, B. Dietzek, M. Feuring-Buske, C. Buske, S. Rau, T. Weil, J. Am. Chem. Soc. 2017, 139, 2512–2519.
- [4] B. S. Howerton, D. K. Heidary, E. C. Glazer, J. Am. Chem. Soc. 2012, 134, 8324–8327.
- [5] G. Ghosh, K. L. Colón, A. Fuller, T. Sainuddin, E. Bradner, J. McCain, S. M. A. Monro, H. Yin, M. W. Hetu, C. G. Cameron, S. A. McFarland, *Inorg. Chem.* **2018**, *57*, 7694–7712.
- [6] T. Sainuddin, J. McCain, M. Pinto, H. Yin, J. Gibson, M. Hetu, S. A. McFarland, *Inorg. Chem.* 2016, 55, 83–95.

- [7] T. Sainuddin, M. Pinto, H. Yin, M. Hetu, J. Colpitts, S. A. McFarland, J. Inorg. Biochem. 2016, 158, 45–54.
- [8] R. Lincoln, L. Kohler, S. Monro, H. Yin, M. Stephenson, R. Zong, A. Chouai, C. Dorsey, R. Hennigar, R. P. Thummel, S. A. McFarland, J. Am. Chem. Soc. 2013, 135, 17161–17175.
- [9] C. Reichardt, M. Pinto, M. Wächtler, M. Stephenson, S. Kupfer, T. Sainuddin, J. Guthmuller, S. A. McFarland, B. Dietzek, J. Phys. Chem. A 2015, 119, 3986–3994.
- [10] G. Shi, S. Monro, R. Hennigar, J. Colpitts, J. Fong, K. Kasimova, H. Yin, R. DeCoste, C. Spencer, L. Chamberlain, A. Mandel, L. Lilge, S. A. McFarland, *Coord. Chem. Rev.* 2015, 282–283, 127–138.
- [11] H. Yin, M. Stephenson, J. Gibson, E. Sampson, G. Shi, T. Sainuddin, S. Monro, S. A. McFarland, *Inorg. Chem.* 2014, *53*, 4548–4559.
- [12] B. A. Albani, B. Peña, N. A. Leed, N. A. B. G. De Paula, C. Pavani, M. S. Baptista, K. R. Dunbar, C. Turro, J. Am. Chem. Soc. 2014, 136, 17095–17101.
- [13] S. A. McFarland, A. Mandel, R. Dumoulin-White, G. Gasser, Curr. Opin. Chem. Biol. 2020, 56, 23–27.
- [14] Y. Tamaki, K. Tokuda, Y. Yamazaki, D. Saito, Y. Ueda, O. Ishitani, Front. Chem. 2019, 7, 1–9.
- [15] N. H. Damrauer, J. K. McCusker, J. Phys. Chem. A 1999, 103, 8440-8446.
- [16] D. Grebner, M. Helbig, S. Rentsch, J. Phys. Chem. 1995, 99, 16991– 16998.
- [17] J. A. Roque III, P. C. Barrett, H. D. Cole, L. M. Lifshits, E. Bradner, G. Shi, D. von Dohlen, S. Kim, N. Russo, G. Deep, C. G. Cameron, M. E. Alberto, S. A. McFarland, *Inorg. Chem.* 2020, *59*, 16341–16360.
- [18] J. A. Roque III, P. C. Barrett, H. D. Cole, L. M. Lifshits, G. Shi, S. Monro, D. Von Dohlen, S. Kim, N. Russo, G. Deep, C. G. Cameron, M. E. Alberto, S. A. McFarland, *Chem. Sci.* 2020, *11*, 9784–9806.
- [19] A. E. Curtright, J. K. McCusker, J. Phys. Chem. A 1999, 103, 7032-7041.
- [20] L. M. Lifshits, J. A. Roque III, P. Konda, S. Monro, H. D. Cole, D. von Dohlen, S. Kim, G. Deep, R. P. Thummel, C. G. Cameron, S. Gujar, S. A. McFarland, *Chem. Sci.* **2020**, *11*, 11740–11762.
- [21] C. Reichardt, S. Monro, F. H. Sobotta, K. L. Colón, T. Sainuddin, M. Stephenson, E. Sampson, J. Roque, H. Yin, J. C. Brendel, C. G. Cameron, S. McFarland, B. Dietzek, *Inorg. Chem.* 2019, *58*, 3156–3166.
- [22] I. J. MacDonald, T. J. Dougherty, J. Porphyrins Phthalocyanines 2001, 5, 105–129.
- [23] X. Y. Wang, A. Del Guerzo, R. H. Schmehl, J. Photochem. Photobiol. C 2004, 5, 55–77.
- [24] N. D. McClenaghan, Y. Leydet, B. Maubert, M. T. Indelli, S. Campagna, Coord. Chem. Rev. 2005, 249, 1336–1350.
- [25] R. Englman, J. Jortner, Mol. Phys. 1970, 18, 285-287.
- [26] K. Peuntinger, T. D. Pilz, R. Staehle, M. Schaub, S. Kaufhold, L. Petermann, M. Wunderlin, H. Görls, F. W. Heinemann, J. Li, T. Drewello, J. G. Vos, D. M. Guldi, S. Rau, *Dalton Trans.* 2014, 43, 13683–13695.
- [27] K. Suzuki, A. Kobayashi, S. Kaneko, K. Takehira, T. Yoshihara, H. Ishida, Y. Shiina, S. Oishi, S. Tobita, Phys. Chem. Chem. Phys. 2009, 11, 9850–9860.
- [28] A. A. Abdel-Shafi, P. D. Beer, R. J. Mortimer, F. Wilkinson, *Helv. Chim. Acta* 2001, 84, 2784–2795.

Manuscript received: November 27, 2020 Revised manuscript received: December 24, 2020 Version of record online: January 19, 2021