

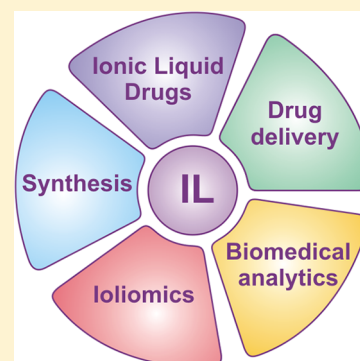
Biological Activity of Ionic Liquids and Their Application in Pharmaceuticals and Medicine

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ABSTRACT: Ionic liquids are remarkable chemical compounds, which find applications in many areas of modern science. Because of their highly tunable nature and exceptional properties, ionic liquids have become essential players in the fields of synthesis and catalysis, extraction, electrochemistry, analytics, biotechnology, etc. Apart from physical and chemical features of ionic liquids, their high biological activity has been attracting significant attention from biochemists, ecologists, and medical scientists. This Review is dedicated to biological activities of ionic liquids, with a special emphasis on their potential employment in pharmaceuticals and medicine. The accumulated data on the biological activity of ionic liquids, including their antimicrobial and cytotoxic properties, are discussed in view of possible applications in drug synthesis and drug delivery systems. Dedicated attention is given to a novel active pharmaceutical ingredient-ionic liquid (API-IL) concept, which suggests using traditional drugs in the form of ionic liquid species. The main aim of this Review is to attract a broad audience of chemical, biological, and medical scientists to study advantages of ionic liquid pharmaceuticals. Overall, the discussed data highlight the importance of the research direction defined as "Ioliomics", studies of ions in liquids in modern chemistry, biology, and medicine.



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1. INTRODUCTION

The first report of Wilkes and Zaworotko on air- and moisture-stable room-temperature liquid salts in 1992¹ has initiated paramount research activity in various areas of chemistry. Further studies have shown that ionic liquids (ILs) are very efficient in chemical synthesis and catalysis,^{2–24} electrochemistry,^{25–32} biomass conversion,^{33–41} fuel production and processing,^{42,43} liquid crystal development,⁴⁴ biotransformation,^{16,45} biotechnol-

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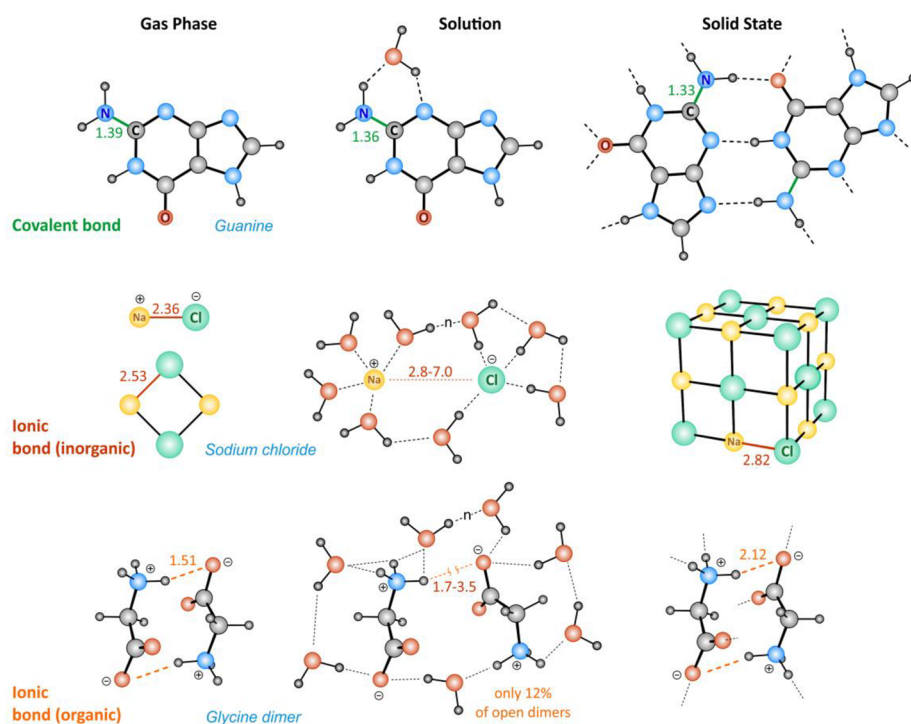


Figure 1. Differences between structures of covalent and ionic compounds in the gaseous phase, solution, and solid phase (the interatomic distances are in Å).

ogy,^{20,21,24,46–50} and many other fields.^{35,51–61} The scope of in-depth studies on ILs covers a wide range of topics and includes their structures and properties,^{40,57,59,62–66} nano-organization and self-assembly,^{53,67,68} and practical applications in advanced chemical transformations.^{7–9,13,14,16,40,51,57,58,64,69–73} Improvements of separation,^{35,60,74–77} electrochemical behavior,^{26,29,35,40,57,64,78,79} analytics,^{64,80–83} extraction,^{42,84–87} purification and recovery,^{88,89} energetic characteristics,^{43,61,90} and biocatalytic properties^{10,39,57,70,91–97} are well-established areas. Recently, the ecological impact of ILs has also been recognized as an important problem.^{57,98–103} Studies on the properties and applications of ILs are discussed in a number of excellent reviews, with only some representative examples cited above.

Besides their unique properties and usage as solvents, the tunable nature of ILs has facilitated the development of novel valuable topics. ILs are already described as the basis of a quiet revolution in material science.²⁹ The number of articles involving ILs is impressive and is growing significantly more rapidly than that for several other actively developing research areas. It is interesting to note that the focus of many IL studies now evolves in the direction of life sciences and medicine. A thorough literature analysis of publications on ILs suggested that in the next decade bio-medical applications of ILs would become one of the major research trends.^{104,105} Moreover, in a near-term perspective, efficient drugs based on ILs may be considered for approval by FDA.¹⁰⁶

Studies on the biological activity of ILs and their application in pharmaceuticals and medicine are newly emerging, highly important areas, which are the subject of this Review. As a necessary introduction to the topic, we briefly emphasize the fundamental origins of tunable properties of ionic interactions in liquids (section 1.1). The structure and organization of ILs in solution is the crucial point for understanding their interactions with living organisms, and it is discussed in section 1.2, followed by the description of the subject coverage and scope of this

Review (section 1.3). The key areas considered in this Review include biological activity of ILs (section 2), usage of ILs as components of drugs (section 3), and applications of ILs in drug synthesis (section 4).

1.1. Tunable Nature of Ionic Compounds: Ionic versus Covalent Bonds

The tunable nature of ionic compounds, which reflects a fundamental difference between covalent and ionic binding, deserves a special note. Indeed, one of the central topics in chemistry is the nature of chemical interactions, their classification, and physically relevant, quantitative description. At the beginning of the 20th century, the concepts of covalent and ionic bonds were established; later, they were validated from the point of electronic structure of molecules.¹⁰⁷ The covalent bond is a binding interatomic interaction, which is realized by sharing a pair of valence electrons between the interacting atoms. Consequently, the electron pair localizes in the interatomic region and “belongs” to both covalent bond participants.^{108,109} The ionic bond involves an electrostatic interaction, which is formed between atoms with significantly different electronegativity; this difference results in an almost complete (or significant) shift of the shared electron pair from the interatomic region to the more electronegative atom and therefore leads to formation of ions. Currently, the physically relevant division of interatomic interactions into covalent and ionic ones may be carried out according to the analysis of parameters of electron density in the interatomic regions of the interacting atoms.^{110–112}

Significant differences in the electronic structure of covalent and ionic compounds result in substantially different behavior of the corresponding molecules, both in the solid phase and in solution. In the solid phase, ionic compounds, such as salts of alkali metals, are prone to form crystal lattices, as a rule, with close packing; in polar solvents, these compounds dissociate

easily into ions surrounded by apparent solvate shells providing high ionic conductivity of the solution. Molecules with covalent bonds form molecular crystals and preserve molecular structure upon dissolution in liquid phase (without breakage of covalent bonds).¹¹³

Because of the strong interactions of ionic compounds with the solvent medium, the impact of liquid phase on chemical processes involving ions becomes essential, which, in turn, makes ionic chemistry both dramatically manifold (same ion demonstrates different chemistry depending on the media) and very tricky to study by experimental and theoretical methods (especially in the dynamic liquid phase). In particular, ions are always surrounded by solvate shells, which may have a significant influence on mechanisms and kinetics of chemical reactions.¹¹⁴ Association of ions with molecules of the medium results in reorganization, arrangement, and structuring of the latter, which, in turn, influence the mobility of the ions¹¹⁵ and rheological properties of the medium, the latter ones being especially important in biological systems.^{102,116} The effects of structuring and self-organization are most pronounced in ionic liquids, which are effective solvents and components of modern catalytic systems.⁶⁷

A representative example of the differences in the properties of ionic and covalent compounds is shown in Figure 1. For guanine (one of the nucleobases), insignificant changes in the covalent interatomic bond occur upon the transition between phases. The length of the covalent C–N bond in the gas phase, denoted in Figure 1 and determined by the gas-phase electron diffraction (GED) method, is 1.39 Å.¹¹⁷ Microsolvation of the guanine molecule by a water molecule results in decreasing this distance only by 0.03 Å.¹¹⁸ In the anhydrous crystal, guanine molecules are connected by the net of hydrogen bonds forming flat parallel sheets; as a result, the C–N distance decreases only by 0.03 Å, in comparison with the microsolvation state.¹¹⁹ Therefore, for the exemplified guanine molecule, the maximum change of the covalent bond length upon the transition between the phases is less than 0.1 Å.

The ionic bond is significantly more sensitive to the influence of external factors. Thus, according to GED experiments in the gaseous phase, sodium chloride molecules exist both in the monomeric form, characterized by the Na–Cl interatomic distance of 2.36 Å, and in the dimeric form.¹²⁰ At the same time, even upon the dimerization in the gaseous phase, the Na–Cl interatomic distance increases to 2.53 Å, that is, by 0.17 Å. In the solution, sodium chloride dissociates and forms ions. The distance between the ions in solutions varies widely and reaches 7.00 Å, as established by molecular dynamic modeling.¹²¹ In the solid state, sodium chloride forms an ionic crystal with the interionic distance of 2.82 Å.¹²²

The lengths of ionic bonds in organic compounds, for example, in the dimer of the zwitterionic form of glycine, also vary widely depending on the molecular environment. In the gaseous phase, the NH–O distance is about 1.50 Å (Figure 1).¹²³ In the aqueous media, the distance between the zwitterions depends on the solvent concentration. In diluted solutions, the amino acid molecules are entirely separated by the solvent molecules, and even in concentrated solutions only 12% of the glycine molecules form open dimers (i.e., dimers containing monomers bonded by one hydrogen bond), while in the gaseous phase, cyclic dimers with two hydrogen bonds between the monomers are formed.^{124–126} In the crystal form, the length of the hydrogen bond between the zwitterions of α -glycine is 2.12

Å, which is considerably longer than that in the gaseous phase.^{127–129}

The principal role of tunable nature of ionic bonds is not limited to the simple examples described above. The crucial role in the structuring of complex natural biological molecules and self-organizing materials is established.^{130–134} Controlled and predictable production of such compounds, including artificial enzymes,^{135,136} nanoreactors,^{137,138} and nanotubes,¹³⁹ is possible, when quantitative data on the strength of ionic interactions in the solution are available. Recently, an atomic force microscopy-based method for assessing single ionic bonds between organic substrates at the molecular level has been developed.¹⁴⁰ One of the most attractive ionic topics is ion-mediated conformational switches, or molecules, which change their conformation and function upon interacting with anions or cations.¹⁴¹

Ions are key participants supporting vital activity of living organisms on all levels of their organization. Thus, at the level of a single cell, catalytic metal ions, being efficient stabilizers of anionic transition states, are associated with ribozymes and protein metalloenzymes,^{142,143} and are involved in the stabilization of secondary and tertiary protein structures.¹⁴⁴ At the level of a whole organism (mammal), extracellular ions are shown to control neuronal activity during sleep–wakefulness cycles.¹⁴⁵ Unsurprisingly, some severe disorders (e.g., Parkinson's and Alzheimer's diseases) arise due to abnormal functioning of ionic channels, which are positioned in the lipid bilayer of cell membranes and ensure ionic currents into and out of the cell.^{146,147} Members of the family of channel kinases, ones of the most interesting ionic channel proteins, which combine the functions of an ionic channel and a protein kinase within a single polypeptide chain, are implicated in the regulation of such important processes as cell proliferation, differentiation and migration, embryonic development, and cardiac automaticity.^{148–153} Studying molecular mechanisms of disturbances in the functioning of ionic channels is required for understanding causes of the associated diseases and for developing strategies of their treatment. Lately, such data have been appearing especially frequently, in particular, due to methods of molecular modeling.^{147,154,155} Investigations of the mechanisms of action of ionic channels have provided the possibilities of generating ionic channel-based detectors, which are able to capture single ions, molecules, and viral particles, and of developing the method of “single-molecule” mass spectrometry.¹⁵⁶ Such active penetration into the “atomic” details of ionic reactions leads to uprising of new cytological areas, for example, nanophysiology.¹⁵⁷ The importance of ionic interactions in solutions has been highlighted in a number of recent chemical and biological problems.^{44,145,158–162}

Therefore, ionic interactions possess outstanding intrinsic potential for flexible and dynamic behavior, which can be utilized to tune the properties of ionic compounds at different levels. The dynamic and tunable potential of ionic compounds can be most powerfully employed in the liquid phase.

1.2. Structure and Organization of Ionic Liquids

As discussed in the previous section, the combination of liquid phase and ionic character of bonding provides an outstanding opportunity to generate a diversity of physical and chemical properties. Indeed, this combination is most efficiently encoded in the structure of ionic liquids, and here we will briefly highlight the main aspects. Ionic liquids are molten salts, which are liquid at temperatures below 100 °C due to bulky, asymmetric cations

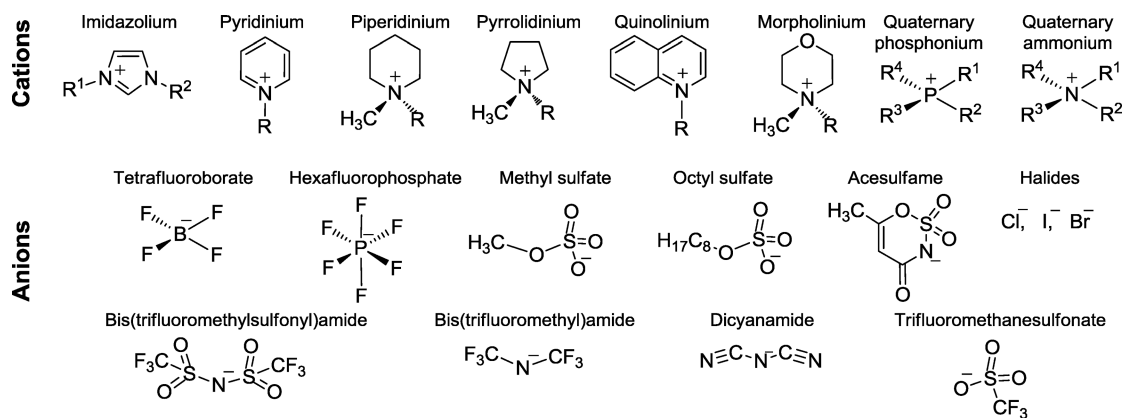


Figure 2. Examples of cations and anions commonly used in ionic liquids.

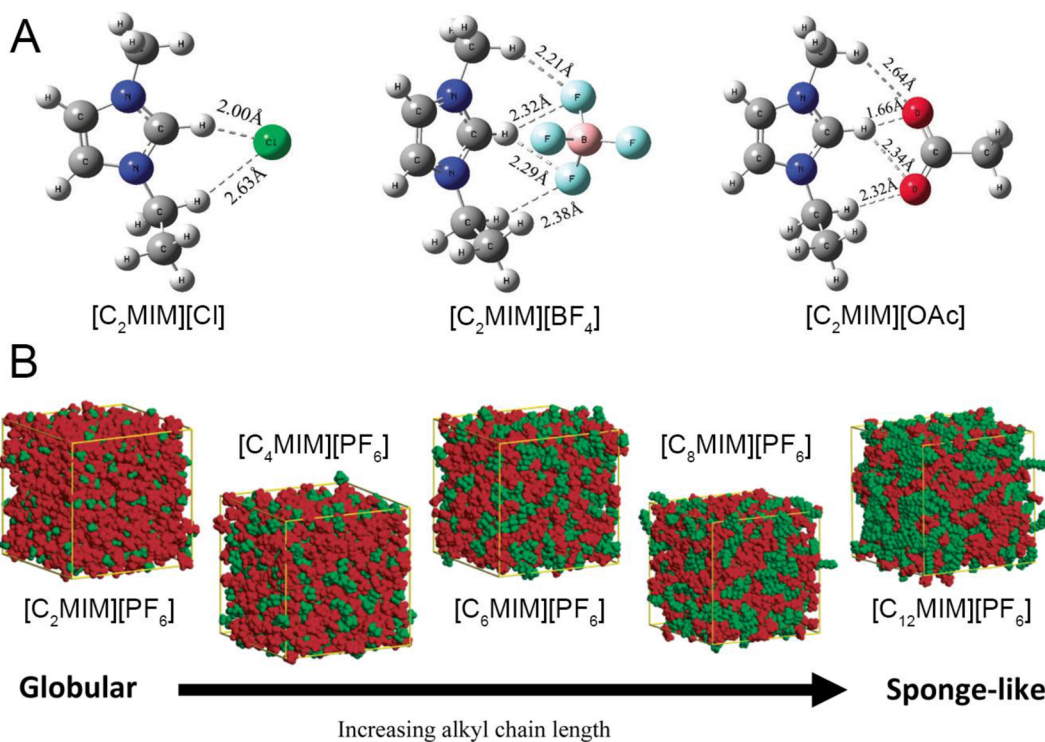


Figure 3. (A) Representative ion pairs and H-bonds in $[\text{C}_2\text{MIM}][\text{Cl}]$, $[\text{C}_2\text{MIM}][\text{BF}_4]$, and $[\text{C}_2\text{MIM}][\text{OAc}]$ (the interatomic distances are in Å). (B) Structure of $[\text{C}_n\text{MIM}][\text{PF}_6]$, where $n = 2-12$ (each box contains 700 IL ion pairs; polar domains are shown in red (anion, cation imidazolium ring), and nonpolar domains are shown in green (cation alkyl chains)). Reproduced with permission from refs 67 (Copyright 2015 American Chemical Society), 185 (Copyright 2016 Royal Society of Chemistry), and 186 (Copyright 2006 American Chemical Society).

and weakly coordinating anions that destabilize the crystal lattice.^{20,53,67} The impact and wide areas of application of ILs may be explained by their extraordinary flexibility. The number of possible IL combinations is overwhelming and approaches 10^{18} , suggesting that almost any desirable properties may be combined within one IL molecule.¹⁶³ Some representative structures of cations and anions are shown in Figure 2.

Recent progress in analytical studies of ILs using NMR,^{55,164-169} mass spectrometry,¹⁷⁰⁻¹⁷³ thermal techniques,¹⁷⁴⁻¹⁷⁶ theoretical modeling,^{41,52,53,177-180} and other analytical approaches¹⁸¹⁻¹⁸⁴ has revealed an important mechanistic picture. As expected, ILs fully realize the potential of tunable interactions at the molecular level, as discussed above (Figure 1). Moreover, the molecular properties of ILs (Figure 3A) modulate higher-order nano- and microscale levels of organization (Figure 3B).

Accumulated evidence points out that, in contrast to conventional molecular liquids, ILs can be considered as nanoheterogeneous media. The ions in the liquid phase tend to self-assemble and form long-living amphiphilic nanostructures. Such organization is uncommon, because most of molecular solvents do not demonstrate structural ordering aside from interactions between neighboring molecules. Nanoscale and microscale ordering possibly provides ILs with their unique properties, which, in conjunction with the vast variability of available ions, make ILs a substance of choice for numerous applications.⁶⁷

Experimental studies on the microstructure and spatial characteristics of ILs are rather complicated and difficult to perform. Most data on the IL structuring at the molecular level have been obtained by theoretical methods, in particular, by

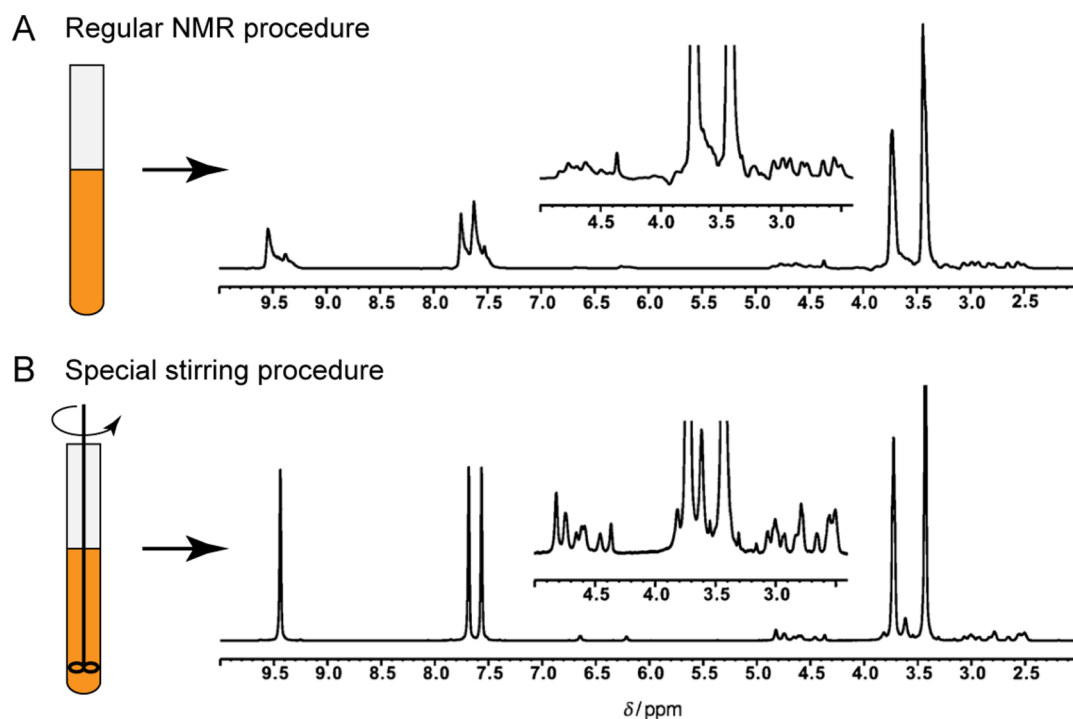


Figure 4. Spectral evidence of nanoheterogeneous nature of IL system: (A) ^1H NMR spectra of glucose in $[\text{C}_4\text{MIM}][\text{Cl}]$ with conventional sample preparation shows characteristic distortions, and (B) sample preparation with additional external stirring eliminates these distortions (magnified regions containing the glucose signals are shown in the insets). Spectra reproduced with permission from ref 165. Copyright 2012 Wiley-VCH Verlag GmbH & Co. KGaA.

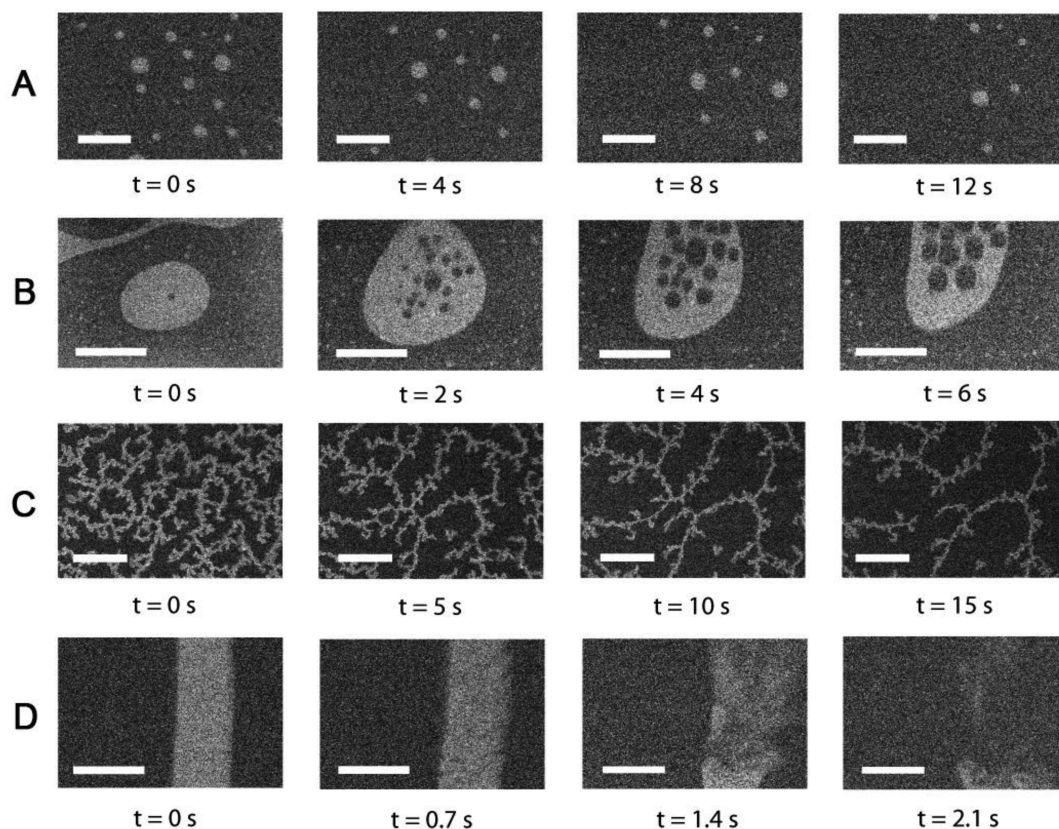


Figure 5. Self-organization and dynamics of water-containing structures in $[\text{C}_4\text{MIM}][\text{BF}_4]$ under electron-beam irradiation: (A) droplet movement; (B) formation of cavities within the droplet; (C) meshwork movement and cleavage; and (D) channel rupture (scale bars correspond to $10\ \mu\text{m}$ (A,C), $20\ \mu\text{m}$ for $t = 0\text{ s}$ and $15\ \mu\text{m}$ for $t = 2\text{--}6\text{ s}$ (B), and $5\ \mu\text{m}$ (D)). Reproduced with permission from ref 200. Copyright 2016 Wiley-VCH Verlag GmbH & Co. KGaA.

classical and DFT molecular dynamics modeling (Figure 3B).^{67,179}

The theoretically revealed nanoheterogeneous nature of ILs was confirmed by spectral studies. The heterogeneity turned out to be the main reason for difficulties in the registration of NMR spectra in some IL systems. In regular NMR studies, the sample is homogenized by self-diffusion of solvent and solute molecules in the liquid phase. However, in the case of ILs, it was not sufficient for recording high-quality NMR spectra (Figure 4A). Higher stability of the microphase structures demands a special procedure with intensive stirring directly in the NMR tube to get a required degree of homogenization (Figure 4B).¹⁶⁵

Both ionic interactions and hydrogen bonds have been shown to play significant roles in the IL organization.^{67,68,187,188} Some reports suggest that networks of hydrogen bonds, which stabilize ionic clusters, are, at least partly, responsible for differences between ILs and simple systems consisting of separate ions.⁶⁸ One of the main differences between “classic” organic salts and ILs is the size of the ions. Ions of inorganic salts can be considered as spherical species, whereas bulky ions of ILs possess more complicated shape, which influences the local IL structure,¹⁸⁹ especially the cation volume.¹⁹⁰ In the bulky organic ions of ILs, the charge is distributed over the large volume and is more delocalized in the space. Therefore, the charge density of IL ions is lower than that of small ions of inorganic salts. This effect reduces the strength of electrostatic repulsion between similarly charged organic ions and allows them to converge and to get additional structuring from noncovalent interactions. For example, in 1-(2-hydroxyethyl)-3-methylimidazolium tetrafluoroborate ($[\text{HOCE}_2\text{MIM}][\text{BF}_4]$), the cations formed so-called anti-electrostatic hydrogen bonds, which were found by spectroscopic and theoretical methods.^{191,192} In general, IL ions are capable of forming more diverse and complicated interactions, as compared to inorganic high-temperature molten salts.¹⁸⁷ This capability also may explain considerable differences between the IL structures in the crystalline and liquid states.^{193,194}

Despite significant progress, there are still many questions concerning the IL nanostructure, and a recent review warns us against sticking to a single theory, especially considering some disagreements between different theoretical approaches.⁶⁷ For example, CPMD (Car–Parinello molecular dynamics) modeling of the pure 1-butyl-3-methylimidazolium hexafluorophosphate ($[\text{C}_4\text{MIM}][\text{PF}_6]$) showed numerous cation–anion hydrogen bonds, which were absent in the results of empirical potential molecular dynamics modeling.¹⁹⁵ A crucial structuring role of the hydrogen bonds between the imidazolium and chloride ions was established by CPMD modeling for 1,3-dimethylimidazolium chloride ($[\text{C}_1\text{MIM}][\text{Cl}]$); it was also shown that classical MD underestimated the influence of hydrogen bonding.^{196–198}

The important feature of the IL microstructure is the existence of stable cage structures established by interionic interactions in the liquid phase. Such structures cause the cage effect confirmed by theoretical modeling¹⁸⁹ and experiments.¹⁹⁹ The cage effect manifests itself by trapping and holding substrate molecules in the cage cavity for a relatively long time. Sometimes this time is enough for facilitating a chemical reaction in the molecular cage, as in a dedicated nanoreactor.¹³⁷ The cage effect can exhibit as a catalytic effect due to an increase of molecular collision probability and due to “improvement” of the entropic factor of a chemical reaction.

Recently, the first direct experimental observation of IL microstructuring effect has been done.²⁰⁰ Self-organized water compartments of various morphologies were detected in water–

IL mixtures by using electron microscopy (Figure 5). The dynamic nature of the self-organized structures has been visualized by microscopy observations (Figure 5). It was also found that nanostructured domains in the solution promoted higher yields and selectivity in the biomass conversion reaction.²⁰⁰ Structuring effect of water–IL interactions is known to play an important role (see, for example, refs 201–207). Small-angle neutron scattering (SANS) studies also confirmed the existence of water nanoclusters in $[\text{C}_4\text{MIM}][\text{BF}_4]$, when the water–IL molecular ratio was higher than $\sim 2:1$, and the cluster size was shown to grow upon the increase of the water content.²⁰⁸ Very recently, the effect of water on the nanostructure of alkylammonium alkanooates ($[\text{N}_{00n}][\text{C}_m\text{CO}_2]$) has been studied in details using small- (SAXS) and wide-angle (WAXS) X-ray diffraction.²⁰⁹ The phenomenon concerning water structuring and the influence on molecular interactions have also been recognized in biochemical studies.^{210,211}

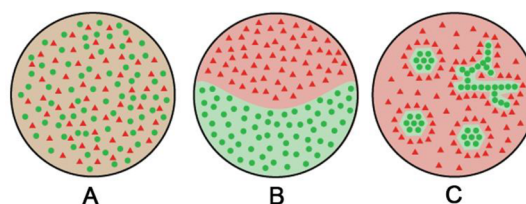


Figure 6. Different liquid-phase systems: (A) uniform mixing of components; (B) separation of phases; and (C) formation of structured system. Reproduced with permission from ref 200. Copyright 2016 Wiley-VCH Verlag GmbH & Co. KGaA.

To summarize the section, the unique nature of IL systems should be emphasized (Figure 6). Usually, liquid-phase chemistry deals with homogeneous solvent–solute systems (A) or with heterogeneous systems involving phase separation (B). In the case of ILs, both types of mixtures can exist depending on the system. However, in many cases, self-organization in ILs leads to the formation of dynamic nano- or micro-sized structures (C), which substantially change the properties of the system. This difference is crucial for the analysis of biological activity of ILs and for discussion of the material described in this Review.

1.3. Subject Coverage and Scope of This Review

A brief introduction into different generations of ILs is required to classify the studies on the biological activity. There are three generations of ILs depending on their chemical structure and properties (Figure 7). The first generation was sensitive to water and air and combined basically dialkylimidazolium and alkylpyridinium cations with metal halide anions. These ILs attracted attention mostly due to their physical properties. The second generation is air- and water-stable; the most common cations include dialkylimidazolium, alkylpyridinium, ammonium, and phosphonium, whereas halides, tetrafluoroborate, and hexafluorophosphate are among the most common anions. These ILs have attracted interest of physics and chemists alike and have found application in various chemical and physical fields. The third generation of ILs employs biodegradable and natural ions, such as choline and amino acids, or ions with known biological activities. These ILs are of interest not only for chemical topics, but also for studies in biology and ecology.^{20,45,167,181,212,213}

As advantageous properties of ILs have been revealed, task-specific ionic liquids (TSILs) have begun to emerge. TSILs carry

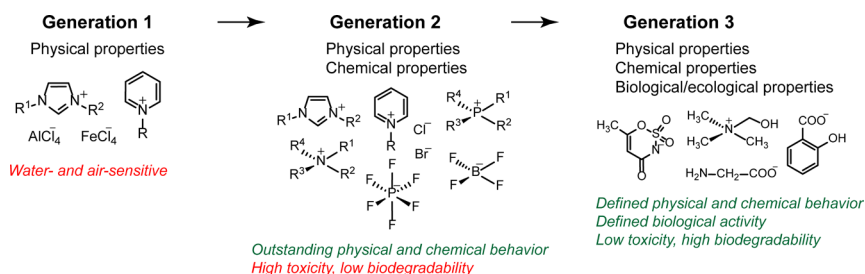


Figure 7. Evolution of ionic liquids. Figure concept reproduced with permission from ref 212. Copyright 2007 Royal Society of Chemistry.

covalently linked functionalities and are tuned for particular applications, where they perform better than traditional solvents/reagents.^{45,55,214–216} Among the examples of TSILs are metal-containing ILs used in catalysis, chiral ILs employed for enantiomer sensing, magnetic ILs, etc.^{55,216}

ILs of generation 2 had been thought to become a benign alternative to toxic organic solvents. Because of their nonvolatile and nonflammable nature, ILs had been supposed to be environmentally safe and “green” chemicals. However, as the data on biological activity of ILs have begun to accumulate, this belief has changed. Now it is obvious that many ILs demonstrate significant biological activity, which is of similar level or even higher, as compared to regular organic solvents and small molecules. Moreover, good water solubility assists the penetration of ILs into various ecological systems. Therefore, in the last decade, the evaluation of environmental impact of ILs has gained ground.²¹⁷ High biological activity of ILs presents disadvantages for using them as solvents or catalysts in chemical sciences. On the other hand, the biological activity of ILs has attracted special attention in pharmaceuticals.^{212,218} Such opportunities demand for a broad discussion and investigation of bio-related properties of ILs.

There are several books and book chapters dedicated to biological properties of ILs.^{20,24,46,100–102,213,219} Dedicated reviews address biocatalytic usage,^{10,39,57,70,91–97,220} toxicity and environmental effects,^{56,57,217,221,222} biodegradation,^{57,103} and pharmaceutical aspects^{35,61,218,223–232} of ILs.

The main aim of this Review is to draw the attention of a broad audience of chemical, biological, and medical scientists to new advantages in the field of ionic liquid pharmaceuticals and to discuss a novel active pharmaceutical ingredient-ionic liquid (API-IL) concept. Here, we address the advantages of using ILs in modern pharmaceuticals and describe state-of-the-art in the research on their biological activity and biomedical applications. Main fields of pharmaceutical employment include usage of ILs as components of drug or drug delivery systems and as complementary participants in drug synthesis.

This Review is divided into three sections. In the first, we give a brief overview of known biological activities of ILs, environmental toxicity, antimicrobial properties, and cytotoxicity against cancer cells, and touch upon possibilities of IL biodegradation. The second section discusses direct employment of ILs as components of drug delivery systems and highlights the concept of API-ILs, together with available data on medical properties and clinical trials of ionic liquid pharmaceuticals. The third section is dedicated to IL usage in drug synthesis and also provides concise coverage on biomedical analytical applications of ILs. The overall summary of the topics considered upon preparation of this Review is shown in Figure 8.

Throughout this Review, we pay attention to mechanistic studies of ILs using experimental and theoretic approaches.

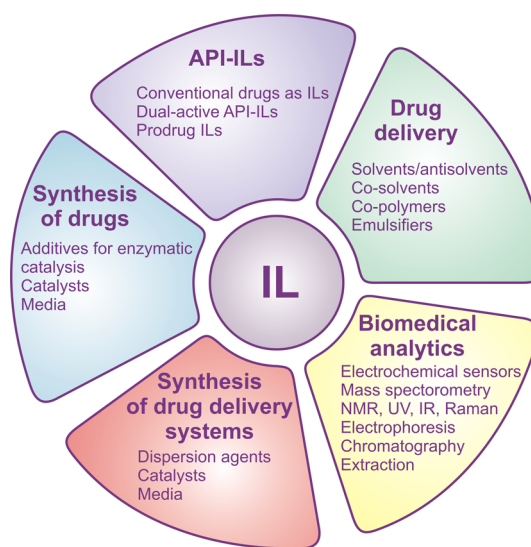


Figure 8. Graphical summary of possible pharmaceutical and medicinal applications of ionic liquids.

Understanding the nature of IL systems is important for explaining and predicting their behavior in biological and pharmaceutical systems. A graphical overview of the methods currently applied for studying biology- and medicine-related characteristics of ILs, which are discussed in this Review, is provided in Figure 9.

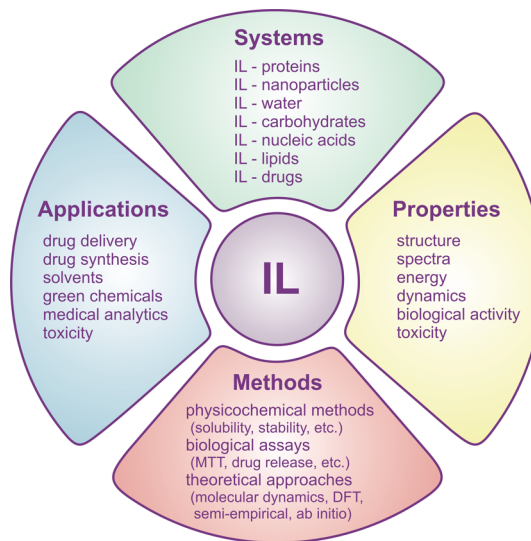


Figure 9. Systems, approaches, and methods currently applied for studying biology- and medicine-related properties of ionic liquids.

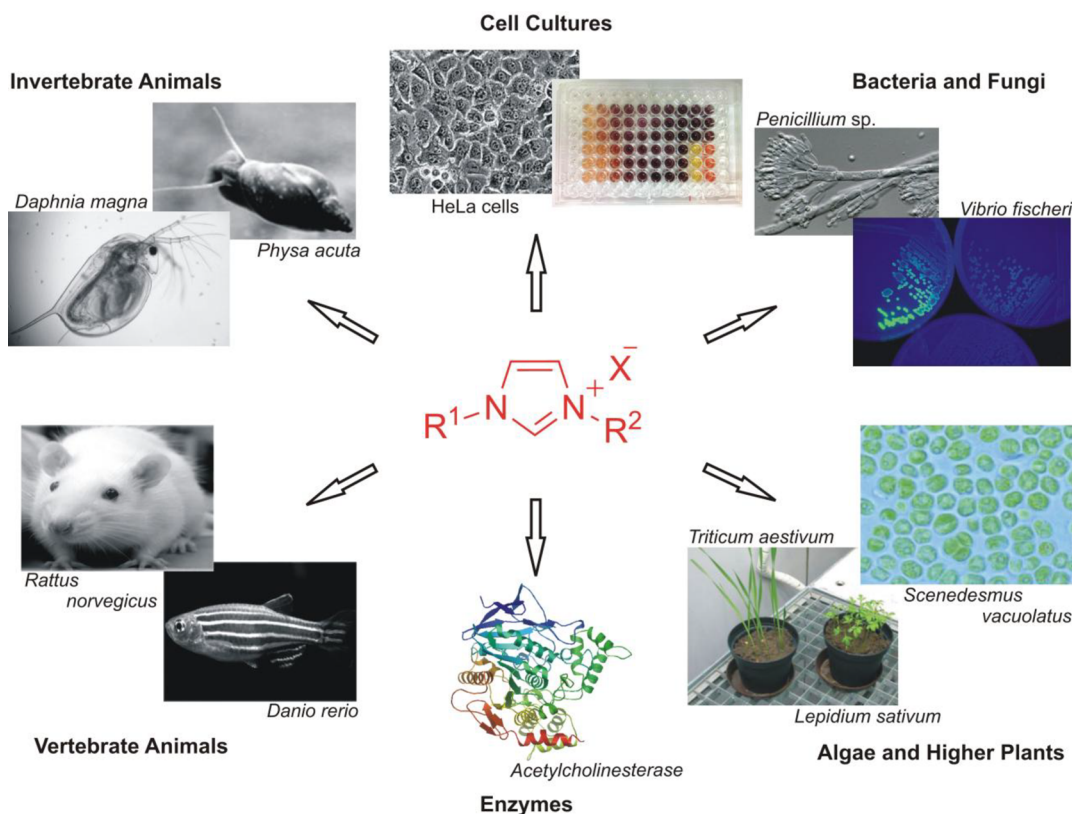


Figure 10. Ionic liquids are active toward all levels of life. Reproduced with permission from ref 217. Copyright 2014 Wiley-VCH Verlag GmbH & Co. KGaA.

2. REVEALING BIOLOGICAL ACTIVITY OF IONIC LIQUIDS

2.1. Organism-Dependent Biological Activity of Ionic Liquids

Nowadays, it is clearly established that ILs have an impact on different levels of life, from single proteins to higher multicellular organisms (Figure 10).^{233–235} In this section, we point out the major key points of biological activity of ILs.

First, an apparently high bioavailability of ILs should be taken into account. Although the particular mechanism of biological activity of ILs may vary from one organism to another, water is crucial for all living systems. Therefore, solubility and interactions with water are one of the factors determining environmental/biological activity of ILs. In the study on ILs with various cations containing butyl or isobutyl side chains and the bis(trifluoromethylsulfonyl)amide anion ($[\text{NTf}_2]^-$), the water solubility of ILs decreased as follows, imidazolium ILs > pyrrolidinium ILs > pyridinium ILs > piperidinium ILs, and was suggested to depend on the water cavitation potential, which was influenced by the size of the IL cation and, to some degree, by the IL aromaticity.²³⁶ A theoretical study on imidazolium ILs with small inorganic anions suggested that both the cation and the chloride ($[\text{Cl}]^-$), bromide ($[\text{Br}]^-$), and tetrafluoroborate ($[\text{BF}_4]^-$) anions interacted strongly with water molecules, whereas the hydrophobic hexafluorophosphate ($[\text{PF}_6]^-$) anion formed no stable interactions with water. According to this study, the ion pairs also interacted with water, but these interactions were significantly weaker.²³⁷ In the case of aqueous 1-ethyl-3-methylimidazolium acetate ($[\text{C}_2\text{MIM}][\text{OAc}]$), the main interactions were hydrogen bonds between water molecules and the anion.²³⁸ $[\text{C}_4\text{MIM}][\text{OAc}]$ demonstrated weakening of cation–

anion interactions due to interactions between the anion and water in water solutions.²³⁹

Biological activity of ILs was shown to depend on their hydration state, and a threshold hydration number was established. About seven water molecules per ion pair were suggested to be the boundary at which biological effects of IL–water mixtures changed dramatically, independent of the ion nature. The first six molecules of water supposedly formed strong interactions with the ion pair.²⁴⁰ Interestingly, when the ratio of water molecules to IL molecules did not exceed one to three, the solution retained the polar ionic network.²³⁶ Upon subsequent dilution, water molecules surrounded the IL network causing its stretching and, finally, disruption.²⁴¹

Corroborating the above-mentioned dependence of biological activity of ILs on the presence of water (for possible structural factors, see also section 1.2), simulations of interactions between water solutions of ILs and phospholipid bilayers suggest that the behavior of the IL anion is governed by its interactions with water molecules. Thus, small hydrophilic anions, such as $[\text{Cl}]^-$, stayed in the solution, whereas more hydrophobic, bulkier $[\text{PF}_6]^-$ formed a film at the lipid/water boundary. In the case of the hydrophobic $[\text{NTf}_2]^-$, the anion followed the imidazolium cation into the lipid bilayer and was located in the interjacent region between the hydrocarbon tails and polar heads of the phospholipid (Figure 11).²⁴² The chloride anion reduced the cation insertion, as compared to lactate.²⁴³

The analysis of the general profile of biological activity of ILs shows that it is strongly dependent on the organism.^{217,221,222} Thus, in the case of the common IL $[\text{C}_4\text{MIM}][\text{NTf}_2]$, the experimental values of EC_{50} (the effective concentration resulting in 50% reduction of processes) vary as follows: 500 and 1170 μM for the cell cultures HeLa and IPC-81, respectively;

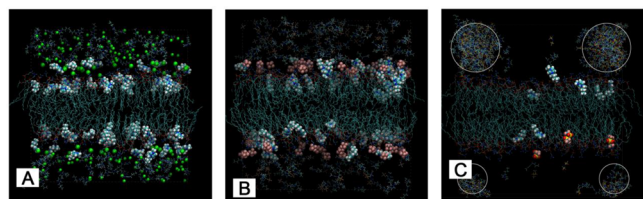


Figure 11. $[\text{C}_4\text{MIM}][\text{Cl}]$ (A), $[\text{C}_4\text{MIM}][\text{PF}_6]$ (B), and $[\text{C}_4\text{MIM}][\text{NTf}_2]$ (C) in the phospholipid bilayer. Figures reproduced with permission from ref 242. Copyright 2012 American Chemical Society.

$30 \mu\text{mol kg}^{-1}$ dry weight soil for reproduction inhibition of the springtail *Folsomia candida*; $50 \mu\text{M}$ for reproduction inhibition of the alga *Scenedesmus vacuolatus*; and $380 \mu\text{M}$ for growth inhibition of the freshwater plant *Lemna minor*. The 24-hour minimum inhibitory concentrations (MIC) of the IL for various fungi and Gram-positive and Gram-negative bacteria exceed $500 \mu\text{M}$.²¹⁷ Similar trends are observed for other ILs, such as $[\text{C}_4\text{MIM}][\text{Br}]$ (48-h EC_{50} 2692 and $2750 \mu\text{M}$ for the IPC-81 and HeLa cells, respectively; 48-h LC_{50} $70 \mu\text{M}$ for the crustacean *Daphnia magna*; 96-h LC_{50} 5887 and $1045 \mu\text{M}$ for the zebra mussel *Dreissena polymorpha* and freshwater snail *Physa acuta*, respectively; 96-h EC_{50} (growth inhibition) 102 and $110 \mu\text{M}$ for the green algae *Scenedesmus obliquus* and *Chlorella vulgaris*, respectively; 4-d LC_{50} $9800 \mu\text{M}$ for the marine macroalga *Ulva lactuca*; and 15 min EC_{50} (luminescence inhibition) $3359 \mu\text{M}$ for the Gram-negative marine bacterium *Vibrio fischeri*), $[(\text{C}_4)_4\text{N}][\text{Br}]$ (48-h EC_{50} $178 \mu\text{M}$ for the IPC-81 cells; 96-h LC_{50} $1800 \mu\text{M}$ for *Physa acuta*; and 15 min EC_{50} (luminescence inhibition) $1862 \mu\text{M}$ for *Vibrio fischeri*), and $[\text{C}_1\text{C}_4\text{Pyr}][\text{NTf}_2]$ (48-h EC_{50} $1000 \mu\text{M}$ for the IPC-81 cells; 48-h LC_{50} $88 \mu\text{M}$ for *Daphnia magna*; 72-h EC_{50} (growth inhibition) $>237 \mu\text{M}$ for *Scenedesmus capricornutum*; EC_{50} (growth inhibition) $955 \mu\text{M}$ for *Lemna minor*; and 30 min EC_{50} (luminescence inhibition) $350 \mu\text{M}$ for *Vibrio fischeri*).²¹⁷

In the subsequent sections 2.2–2.5, we discuss particular aspects of biological activity of ILs important for the topic of this Review. Note that these sections provide data on common ILs, which are most often used in chemical research and industry. ILs with tailored biological activity are discussed later in section 3.

2.2. Ionic Liquids as Antibacterial and Antifungal Agents

Antimicrobial activity of ionic liquids has quickly attracted the attention of researchers. When it became obvious that numerous types of ILs inhibited growth of various bacterial and fungal species, the medical and industrial potentials of ILs have manifested.

Imidazolium, pyridinium, pyrrolidinium, piperidinium, ammonium, and other ILs were shown to inhibit growth of pathogenic and nonpathogenic bacteria and fungi.^{217,244–257} These results have presented both advantages and disadvantages. Thus, high activity toward microorganisms may seriously hinder the application of ILs in biotechnology; however, it may be used as a valuable property in medicine (Figure 12).

From the biotechnological point of view, toxicity of new IL solvents may become a serious problem, and evaluation and selection of appropriate ILs for various biotechnological processes have been actively investigated lately.^{258–261} Imidazolium ILs were shown to be toxic toward the probiotic bacterium *Propionibacterium freudenreichii* subsp. *freudenreichii* applied in the dairy industry,²⁶² whereas pyridinium ILs inhibited *Clostridium* sp. involved in biosorption of uranium.²⁶³ Residual 1-ethyl-3-methylimidazolium acetate ($[\text{C}_2\text{MIM}][\text{OAc}]$) in

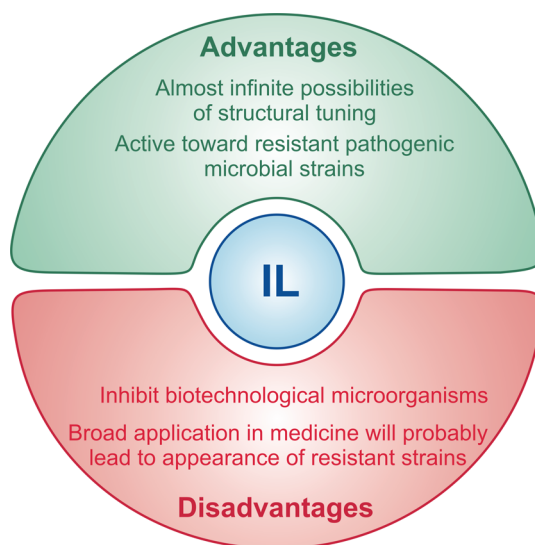


Figure 12. Graphic summary of current advantages and disadvantages of using ionic liquids as antimicrobial agents.

hydrolysates of pretreated lignocellulose biomass inhibited growth and ethanol production of the model yeast *Saccharomyces cerevisiae*.²⁶⁴ Addition of either 1-ethyl-3-methylimidazolium acetate or 1-ethyl-3-methylimidazolium methylphosphonate in small quantities ($\leq 5\%$) led to a metabolic switch from respiration to fermentation (ethanol production);²⁶⁵ in higher concentrations, both ILs inhibited the yeast growth.^{265,266}

The lignocellulolytic bacterium *Enterobacter lignolyticus* was tolerant to high concentrations of $[\text{C}_2\text{MIM}][\text{Cl}]$, possibly due to changes in the cell membrane composition, down-regulation of membrane porins, and up-regulation of drug efflux pumps.^{267,268} Correspondingly, a method for tolerance enhancement via heterologous expression of efflux pumps in bacteria was proposed,²⁶⁹ and a technique for enrichment of IL-tolerant microorganisms in microbial communities was suggested.²⁷⁰ Interestingly, a single mutation in a transcriptional regulator made *Escherichia coli* tolerant to $[\text{C}_2\text{MIM}][\text{OAc}]$.²⁷¹ $[\text{C}_4\text{MIM}][\text{PF}_6]$ and $[\text{C}_4\text{MIM}][\text{NTf}_2]$ were considered compatible with whole-cell biotechnological applications.²⁷² *Penicillium* sp., the specimen of which could produce valuable bioactive substances, demonstrated very high tolerance to IL treatment; in some cases, even 50 mM of IL did not inhibit the growth of the fungi.²⁷³ *Aspergillus* isolates were tolerant to $[\text{C}_2\text{MIM}][\text{OAc}]$.²⁷⁴ Morpholinium ILs demonstrated low toxicity toward bacteria and fungi.²⁷⁵ Thus, a search for IL-tolerant valuable microbial communities is ongoing.²⁷⁶

From the medical point of view, ever-evolving resistance of microorganisms to the existing drugs is an urgent issue, and a whole new class of possible antimicrobial agents is a very timely finding. Targeted attempts to develop antimicrobial ILs for medical purposes have been made. Thus, comparison of activity of ampicillin-carrying ILs with that of sodium ampicillin clearly showed possible advantages of using ILs as antimicrobial agents.²⁷⁷ More examples of ILs with targeted antimicrobial activity are discussed in section 3.4.

ILs have demonstrated impressive inhibitory results even in the case of resistant and biofilm-forming microorganisms. Low concentrations of $[\text{C}_{16}\text{MIM}][\text{Cl}]$ strongly inhibited growth of multidrug-resistant *Candida tropicalis*,²⁷⁸ whereas 1-alkyl-3-methylimidazolium chlorides, bromides, and iodide were active toward both planktonic and biofilm bacteria and fungi.^{279–281} 1-

Alkyl-3-methylimidazolium fumarates possessed high antimicrobial potential and were suggested to be used for medical purposes.²⁸² Ammonium ILs with azolate anions were found to be powerful antibacterial and antifungal substances;²⁸³ hydroxylammonium ILs were highly active against such human pathogens as *Staphylococcus aureus*, *Salmonella typhi*, and *Vibrio cholera*,²⁸⁴ whereas diphosphonium ILs displayed a broad spectrum of antimicrobial activity against ocular pathogens.²⁸⁵ Of note, triphenylamine phosphonium ILs, which self-assembled into nanostructures, showed potent activity against Gram-positive bacteria, including *S. aureus*.²⁸⁶

However, possible negative effects associated with broad application of ILs in medicine should not be overlooked. While increased IL tolerance is a benefit in biotechnologically relevant microorganisms, in pathogenic bacteria and fungi it may become a significant difficulty. It seems that, similarly to conventional antimicrobial agents, ILs also may provoke the appearance of resistant strains. Thus, [C₄MIM][PF₆] was shown to induce the expression of antibiotic resistance genes and to activate horizontal gene transfer in freshwater bacteria.²⁸⁷ Of note, cholinium chloride ([Cho][Cl]) and [C₂MIM][Cl] significantly influenced the expression of genes involved in primary and secondary metabolism in *Aspergillus nidulans*; the authors suggested the treatment with ILs to be employed for boosting the diversity of natural active compounds produced by the fungus.²⁸⁸

As we discuss in section 2.5, high antimicrobial activity of many ILs may be possibly explained by interactions between IL and the cell membrane.^{49,259,289,290} Thus, alkyltributylphosphonium chlorides with long alkyl chains provoked substantial damage in conidia of *A. nidulans*.²⁹¹ For imidazolium ILs with short alkyl chains, the toxicity was found to depend on the chaotropicity of the anion.²⁹²

Antimicrobial properties of ILs were suggested to be employed in the development of new IL fungicides,²⁹³ and in the protection of natural fabric,²⁹⁴ paper,²⁹⁵ and metal surfaces^{296,297} against microbial growth. Similarly, graft polymers of poly(ionic liquids) were proposed to be used in antibacterial coatings.^{298,299} Alkoxyethyl(2-decanoyloxyethyl)dimethylammonium bis-(trifluoromethylsulfonyl)amides and 1-methyl-3-octyloxymethylimidazolium tetrafluoroborate performed good in preservation and fixation of tissues.^{245,300,301} Attempts to create imidazolium-based poly-IL membranes with improved antibacterial activity have been made.³⁰²

2.3. Ionic Liquids as Anticancer Agents

Cytotoxicity of a substance is a form of its biological activity, and numerous studies on cytotoxicity of ionic liquids toward various cells have been reported. Both normal and cancerous cells of invertebrate and vertebrate species were investigated, including insects (*S2, Drosophila melanogaster* cell culture),³⁰³ fish (CCO, channel catfish ovary cell line),^{304–306} mouse (3T3, fibroblasts;^{307,308} EMT6, mammary cell line;³⁰⁹ J774, macrophages;³¹⁰ MC3T3-E1, osteoblasts³¹¹), rat (C₆, glial cells;^{257,312} IPC-81, promyelotic leukemia;^{234,275,312–317} PC12, pheochromocytoma^{318–320}), Chinese hamster (V79, lung fibroblasts),³²¹ and human (3215 LS, fibroblasts;³²² A549, lung carcinoma;^{323–325} A431, squamous carcinoma;³²⁶ CaCo-2, colon carcinoma;^{308,322,327–332} HaCaT, immortalized keratinocytes;³³³ HCT-116, colon carcinoma;^{323,324,326,334} HEK, embryonic kidney;^{257,335} HeLa, cervical cancer;^{304,336–339} HepG2, hepatocyte carcinoma;^{332,334,340} HT-29, colon carcinoma;^{323,324,326,328} MCF-7, breast cancer;^{323,324,326,334,341–344}

T98G, brain cancer;³³⁵ U937, histiocytic lymphoma;³⁴⁵ and other^{323,324,326}).

There are several methods for assessing cytotoxicity of a chemical, and the MTT assay is among the most popular. It is based on detecting the activity of mitochondrial enzymes, which reduce the tetrazolium dye MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) or the related MTS, XTT, and WST dyes.³⁴⁶ Preliminary studies on the mechanism of IL cytotoxicity have also been carried out. By using fluorescence microscopy and dedicated apoptosis assays, ILs were shown to induce oxidative stress, DNA damage, and apoptosis in the cells.^{307,308,319,320,326,338,340}

The cytotoxicity of ILs depends on their structure and varies widely, from micromolar to millimolar ranges. Thus, cholinium ILs in general demonstrated lower cytotoxicity than ILs with other common cations, such as imidazolium and pyridinium. Several attempts to create a prediction model for assessing IL cytotoxicity in silico have been made;^{329,347–351} however, at the moment, there is no possibility to distinguish the cytotoxicity of ILs only by their structure, because it also strongly depends on the external factors, such as the cell type.²¹⁷

When the cytotoxic activity of ILs has become evident, studies on possible application of ILs as anticancer agents have begun. Once again, the high tunability of ILs has become the major driving force of research: an idea of creating therapeutic agents with tailored anticancer activity and reduced toxicity toward the human organism seems very attractive.^{323,324} Nevertheless, despite active investigations in the area, our knowledge on the possibility of using ILs as anticancer agents remains incomplete (Figure 13).

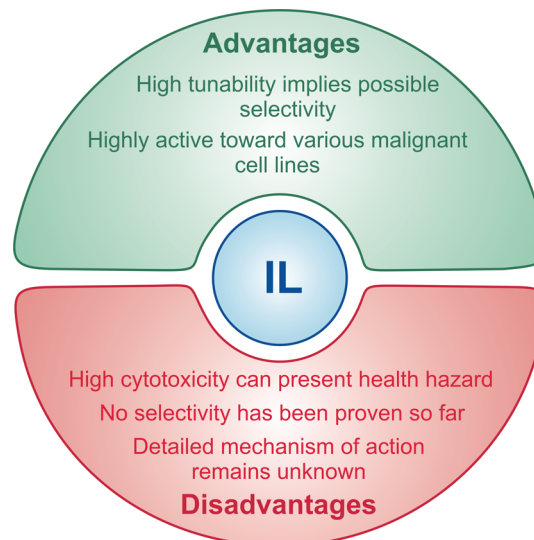


Figure 13. Graphic summary of current advantages and disadvantages of using ionic liquids as anticancer agents.

A screening of imidazolium, phosphonium, and ammonium ILs in 60 human tumor cell lines was conducted in search of potential anticancer substances with high cytostatic and low cytotoxic activity.^{323,324,326} Guanidinium ILs with long alkyl chains showed high cytotoxicity toward several tumor cell lines.³⁵² While searching for environmentally safe ILs, it was demonstrated that introduction of natural amino acids into ILs with the tetrafluoroborate anion led to an unexpected increase of their cytotoxicity, possibly due to the enhancement of transport of the toxic anion into the cells.³⁰⁸ Still, almost no attempts to

compare the cytotoxic effect of ILs on tumor and normal cells have been made so far.

Triethylammonium sulfate, triethylammonium phosphate, 1-methylimidazolium chloride, and 1-butyl-3-methylimidazolium chloride were less toxic toward nonmalignant HEK (human embryonic kidney) cells than toward T98G brain cancer cells.³³⁵ However, no significant difference was found for cytotoxicity exhibited by imidazolium-based ILs toward normal human fibroblasts and CaCo-2 (colorectal adenocarcinoma) cells,³²² which indicated that the IL effect was nonselective.

Accumulated data on mechanisms of IL cytotoxicity suggest that ILs disturb lipid bilayers of the cell membrane, and hydrophobicity and lipophilicity of the cation correlate with the cytotoxic action.^{307,314,315,318,345,353,354} Treatment with 1-alkylquinolinium ILs caused cell membrane disruption in the 3T3 cells.³⁰⁷ Imidazolium ILs induced mitochondrial failure, oxidative stress, and apoptosis both in malignant and in normal cells.^{308,319,320,326,338,340} In the mouse mammary carcinoma cells EMT6, treatment with 1-octyl-3-methylimidazolium chloride ([C₈MIM][Cl]) induced expression of cytochrome P450 genes, products of which were involved in metabolism of drugs and other exogenous substances.³⁰⁹ In HeLa cells, imidazolium ILs induced the multixenobiotic/multidrug resistance (MXR/MDR) system.³³⁹

Apparently, the mechanism of action is the current stumbling block of studies on anticancer properties of ILs. It will be impossible to design novel ILs with targeted antitumor activity until we know the details of transformations and actions displayed by ILs inside both the malignant and the normal cells.

2.4. Biodegradation of Ionic Liquids

Wide application of ionic liquids in chemistry, biochemistry, and industry inevitably results in their contact with the environment. From the point of view of ecological safety, chemical compounds should be easily degradable and should decompose into harmless substances that do not accumulate in the environment.^{355,356} Biodegradability of ILs is a hot research topic,^{103,357,358} and in this section we sum the current knowledge on natural possibilities to decompose ILs.

Biodegradation is realized by bacterial or fungal enzymes and leads to destruction of a chemical structure with corresponding loss of its properties. There are several categories of biodegradability of a substance: primary (the ability to lose a particular structural feature); inherent (the ability to undergo biodegradation); ready (the ability to biodegrade a given % during a given time period, suggesting the possibility of ultimate complete biodegradation); ultimate (the ability to breakdown completely); and mineralization (the ability to decompose into plant-accessible molecules).^{103,357} Substances are classified as "readily biodegradable", if they have demonstrated the ability to biodegrade ultimately and completely; those that have not passed ready biodegradation tests, but nevertheless have demonstrated clear evidence of partial biodegradation, are classified as "inherently biodegradable".³⁵⁸ The Organization of Economic Co-Operation and Development (OECD) proposes standard assays for assessment of the substance biodegradability, which include aerobic and anaerobic conversion in soil, aquatic sediment systems, and surface water.³⁵⁹ Ready biodegradation tests employ strict conditions and are not very realistic regarding a typical environment, whereas inherent biodegradation tests use less constrained conditions and therefore correspond better to the real environment.³⁵⁸

There are several general factors that facilitate or hamper biodegradability of a compound. Thus, the presence of long unsubstituted alkyl chains, oxygen atoms (hydroxyl, aldehyde, or carboxylic groups), sites for enzymatic hydrolysis (ester or amide bonds), and aromatic rings promote biodegradability, whereas branched alkyl chains, quaternary carbon or tertiary nitrogen atoms, halides, and heterocycles hinder biodegradation.^{103,360}

Existing data suggest that ILs with long alkyl side chains are more readily biodegradable but possess higher antimicrobial activity.^{103,361,362} For example, the 1-octyl-3-methylimidazolium cation was readily biodegradable by microbial community of a sewage treatment plant (Jeonju, Korea). The biodegradation process started with oxidation at one of three points in the alkyl chain, and the resulting oxidized compounds could initiate three different degradation pathways.³⁶³ 1-Ethyl-3-methylimidazolium and 1-butyl-3-methylimidazolium ILs showed negligible biodegradability^{272,364–368} and were suggested to be subjected to electrochemical treatment prior to biodegradation.³⁶⁹ Peralkylated and dicationic imidazolium ILs also were not readily biodegradable.^{317,368,370}

The introduction of target groups for enzymatic hydrolysis, such as esters, especially with long alkyl chains, substantially enhanced the biodegradability of imidazolium ILs,^{364,366,371} whereas the introduction of oxygen groups reduced their antibacterial activity.³⁷¹ The presence of long alkyl chains in the anion, for example, in alkylsulfates, also correlated with better biodegradability, whereas halogen-containing anions, such as chlorides, bromides, tetrafluoroborates, and hexafluorophosphates, as well as fluoroorganic and cyano-based anions, could not act as carbon source and therefore did not undergo biodegradation.^{103,365,366,369,372–374}

3-Methyl-1-propoxycarbonylimidazolium, 2,3-dimethyl-1-propoxycarbonylimidazolium, 3-methyl-1-pentoxycarbonylimidazolium, and 2,3-dimethyl-1-pentoxycarbonylimidazolium octyl sulfates were readily biodegradable;³⁶⁶ the same was observed for imidazolium ILs containing alkyl side chains with butoxy or propoxy termini and the octyl sulfate anion, whereas ILs with methoxy or ethoxy termini showed lower biodegradability.³⁷⁵ 1-Butyl-3-methylpyridinium, 1-hexyl-3-methylpyridinium, and 1-octyl-3-methylpyridinium bromides could be fully mineralized by microbial community, but only 1-octyl-3-methylpyridinium bromide was readily biodegradable.^{367,376}

A study on biodegradability of 27 ILs with imidazolium, pyridinium, pyrrolidinium, piperidinium, and morpholinium cations demonstrated that each IL class included readily biodegradable members. Good biodegradability was usually associated with the presence of a long unbranched alkyl side chain or hydroxyl-bearing side chain.³⁷⁷ According to a recent review, several dozens of readily biodegradable ILs have been synthesized to date, and almost 50% of them contain cholinium as cation and an organic acid as anion.¹⁰³ Amino acid-based ILs with the lauryl sulfate anion also demonstrated high biodegradability.³⁷⁸ Conventional imidazolium ILs,^{272,364–368,377,379–382} as well as most tetraalkylammonium,^{313,382,383} morpholinium,^{251,275,377} piperidinium,³⁷⁷ and pyrrolidinium ILs,^{103,357,383} are mostly resistant to biodegradation. Examples of highly and poorly biodegradable ILs are shown in Figure 14.

It also should be noted that the choice of a microorganism can significantly impact the biodegradation. Thus, by using the bacteria *Sphingomonas paucimobilis*, the ILs, which were previously shown to be resistant to degradation, were biodegraded successfully.³⁸⁴

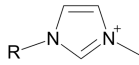




Cationic head group	Side chain (R)	Anion	Biodegradation potential
 Imidazolium	-C ₂ H ₅	Cl ⁻	Not readily
	-C ₃ H ₇	PF ₆ ⁻	Not readily
	-C ₄ H ₉	Cl ⁻	Not readily
	-C ₈ H ₁₇	Cl ⁻	Inherently
	-CH ₂ C(=O)OCH ₂ CH ₃	Br ⁻	Inherently
 Pyridinium	-C ₂ H ₅	Cl ⁻	Not readily
	-C ₃ H ₇	Br ⁻	Not readily
	-C ₄ H ₉	Br ⁻	Not readily
	-CH ₂ CH ₂ OH	Cl ⁻	Inherently
 Pyrrolidinium	-C ₄ H ₉	Br ⁻	Inherently and ultimately
	-C ₈ H ₁₇	Cl ⁻	Readily
	-CH ₂ CH ₂ OH	I ⁻	Not readily
 Morpholinium	-CH ₂ C(=O)OCH ₂ CH ₃	Br ⁻	Inherently and ultimately
	-C ₄ H ₉	Br ⁻	Not readily
	-CH ₂ CH ₂ OH	I ⁻	Not readily
 Morpholinium	-CH ₂ CH ₂ CH ₂ OH	Cl ⁻	Inherently
	-CH ₂ OCH ₂ CH ₃	Cl ⁻	Not readily

Figure 14. Biodegradation potential of exemplary ionic liquids using the original OECD classification of biodegradability (note that it can depend on the evaluation method). “Readily biodegradable” are substances that have passed a ready biodegradation test and have demonstrated the ability to biodegrade a given % during a given time period; “inherently biodegradable” are substances that have demonstrated the ability to biodegrade in an inherent biodegradation test; and those for which such biodegradation is complete are called “inherently and ultimately biodegradable”; “not readily biodegradable” are substances that have not passed a ready biodegradation test and have shown no evidence of inherent biodegradability. Figure concept and data reproduced with permission from ref 377. Copyright 2014 Royal Society of Chemistry.

Several methods of chemically induced degradation of nonbiodegradable ILs have been suggested. These approaches employ electrochemical degradation^{369,385,386} or advanced oxidation processes, such as UV/H₂O₂ oxidation,³⁸⁷ ultrasonic treatment,^{388–390} ozonation,³⁹¹ and the Fenton oxidation.^{390,392,393} In many cases, such pretreatment allows significantly decreasing ecotoxicity and increasing biodegradability of various structural classes of ILs.¹⁰³

As is evident from this section, a considerable part of the existing ionic liquids is nonbiodegradable. Therefore, the issue of their interaction with the environment is of high importance, and we discuss it in the next section.

2.5. Toxicity and Environmental Effects

The number of possible cationic–anionic combinations is virtually unlimited, and, unsurprisingly, a natural IL was discovered. This IL forms when solenopsins from the venom of the ant *Solenopsis invicta* mix with formic acid from the venom of its competitor, the ant *Nylanderia fulva*. Thus, *N. fulva* detoxifies the opponent’s venom by means of IL synthesis.³⁹⁴ Still, not every IL is safer than its precursors, and by now ILs have been shown to influence all levels of life (Figure 15).^{233–235} Several recent reviews discuss the subject of IL ecotoxicology in detail;^{217,222,395} here, we will mention only the major issues.

There is no uniformity in toxic activities exhibited by ILs. Some of them demonstrate relatively low toxicity, whereas others

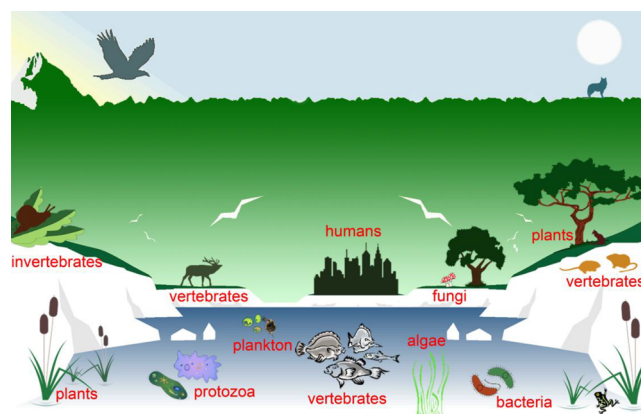


Figure 15. Ecosystems vulnerable to contamination by ionic liquids. Reproduced with permission from ref 217. Copyright 2014 Wiley-VCH Verlag GmbH & Co. KGaA.

impose significant inhibitory effects in various biological systems. Presumably, higher water solubility should correlate with higher environmental danger due to ease of penetration of a water-soluble chemical into various ecosystems. This suggestion is corroborated by the evident interrelation between the biological effect of IL and its interactions with water molecules (see sections 2.1 and 4.1). Many studies have been aimed at finding relationships that link the IL structure to its toxicity. Although no general correlations have been discovered so far, several trends have been revealed. Thus, according to the accumulated data, the IL toxicity depends on (1) the length of alkyl side chain in the cation; (2) the presence and nature of functional groups in the cation; (3) the anion and cation nature; and (4) interactions between anion and cation. It is also clear that the nature of the biological object should be taken into account.²¹⁷

From the environmental point of view, research on multicellular organisms and whole ecosystems provides the best estimation of the ecological impact of a substance. Thus, the influence of ILs has been investigated in plants,^{235,321,396–410} fish,^{411–420} rodents,^{421–427} mollusca,^{428–430} worms,^{431–438} and crustaceans.^{235,317,318,439–443} The small planktonic crustacean *Daphnia magna*, one of the favorite objects of ecologists, proved to be rather sensitive to imidazolium and phosphonium ILs.^{317,318,439–442} Imidazolium ILs provoked developmental damage,^{421,425} whereas ammonium ILs induced acute and subacute toxicity^{423,426,427} in mice and rats. Imidazolium ILs caused oxidative stress and growth inhibition in aquatic algae^{399,401–404,444} and terrestrial plants.^{396,397,400,405,407} Similar results in plants were obtained for ammonium ILs.^{406,408} Even cholinium ILs, which were supposed to be relatively safe due to the natural origin of the cation, exhibited toxicity toward *Daphnia magna* and the common duckweed *Lemna minor*.⁴⁴⁵

Because multicellular organisms are rather problematic and time-consuming objects, which usually do not allow conducting high-throughput screening, many toxicological studies have been carried out on bacteria, fungi, and cell cultures. High antimicrobial, antifungal, and cytotoxic activity of many ILs (see sections 2.2 and 2.3, respectively) may be explained by interactions between alkyl side chains of the IL cation and the membrane of the cell. Long alkyl chains may destabilize the membrane by penetrating into the lipid bilayer and inducing structural damage.^{255,283–285,307,314,315,318,345,353,446,447} According to molecular simulations, imidazolium cations formed hydrophobic contacts with the phosphatidylcholine lipid bilayer,

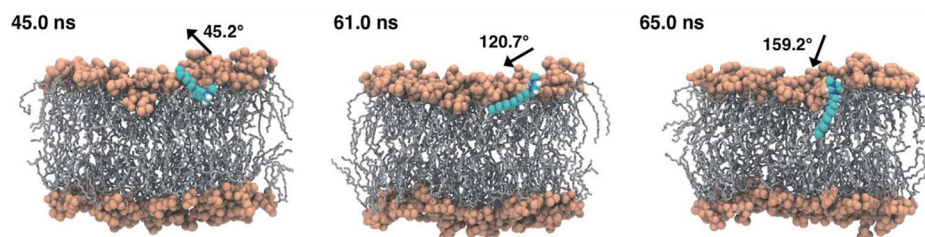


Figure 16. Penetration of the cation of $[\text{C}_{12}\text{MIM}][\text{Cl}]$ into the lipid bilayer. Reproduced with permission from ref 446. Copyright 2014 Royal Society of Chemistry.

entered into it, and subsequently caused roughening of its surface (Figure 16). After partial insertion of the imidazolium ring into the bilayer, the IL cation flipped to bring the whole alkyl chain into the hydrophobic area. Regardless of the alkyl side chain length ($n = 4, 8, 12$), the imidazolium ring and alkyl chain demonstrated strong interactions with the lipid head and tail group, correspondingly; however, the cations with longer alkyl chains penetrated deeper into the bilayer. In contrast to the cation, the hydrophobicity of the anion did not correlate with the structural transformations of the lipid bilayer.⁴⁴⁶ Upon reaching the saturation limit for the insertion of the IL cations, the membrane began to arch, therefore maximizing the area of contact between the lipid and IL. Such bending supposedly caused subsequent morphological defects.⁴⁴⁸ Asymmetric insertion of the IL cations into the lipid bilayer caused morphological transformations.⁴⁴⁹ The presence of oxygen in the side chain hindered the cation insertion,²⁴³ which agreed with lower toxicity of such ILs.^{217,318,345}

Empirical molecular dynamics simulations of a neutral cholesterol lipid bilayer in the water solution of $[\text{C}_4\text{MIM}][\text{Cl}]$ and $[\text{C}_4\text{MIM}][\text{NTf}_2]$ demonstrated that both ILs adsorbed selectively at the water–cholesterol interface, and the ions penetrated partially into the bilayer. $[\text{C}_4\text{MIM}][\text{Cl}]$ is highly soluble in water, and the $[\text{C}_4\text{MIM}]^+$ cations adsorbed at the cholesterol–water interface, whereas the butyl side chains entered the bilayer. $[\text{C}_4\text{MIM}][\text{Tf}_2\text{N}]$, which possessed lower water solubility, formed a thick IL film at the water–cholesterol interface. The hydrophobic $[\text{NTf}_2]^-$ anions easily penetrated into the cholesterol bilayer, which resulted in limited inclusion of the cation imidazolium rings and reduced penetration of the butyl side chains into the cholesterol. It should be noted that upon the contact with the cholesterol bilayer, NaCl and LiCl solutions demonstrated very restrained interactions between the ions and the organic part of the system; that is, the affinity of the bulky organic ions of ILs to the bilayer was governed by dispersion interactions and hydrophobic/hydrophilic effects.⁴⁵⁰

Incorporation of imidazolium ILs into lipid bilayers had an impact on activities of membrane proteins. By decreasing the membrane deformation energy, ILs increased the lifetime of transmembrane channels; positive charges, which accumulated on the membrane surface in the presence of ILs, hindered the cation flux.^{451,452} According to another study, $[\text{C}_4\text{MIM}][\text{Cl}]$ significantly decelerated the translocation of DNA and antibiotics through protein and graphene nanopores.^{453,454}

These theoretical results were supported by experimental data obtained on model lipid membranes,^{49,318,345,353,455–457} cell cultures,³⁰⁷ fungi,²⁹¹ worms,⁴³⁴ and the single-cell green alga *Chlamydomonas reinhardtii*, where ILs manifested their toxicity via cell membrane swelling with subsequent disruption; longer alkyl side chains penetrated into the membrane more easily and therefore displayed more pronounced cytotoxicity.⁴⁴⁸

The toxicity of ILs also may depend on interactions with more specific targets. For example, 1-alkyl-3-methylimidazolium bromide/hydroxide ILs with different alkyl side chains demonstrated the π – π stacking interaction with phenyl rings of the β -tubulin receptor. According to molecular docking studies, the alkyl side chain of the imidazolium core influenced the binding of ILs to the receptor and therefore the vermicial activity of these ILs. Thus, longer side chains correlated with higher vermicial activity.⁴³⁴ In cells expressing human organic cation transporter 2 (hOCT2), $[\text{C}_1\text{C}_4\text{Pyr}][\text{Cl}]$ was shown to inhibit hOCT2-mediated transport processes.⁴²⁴

Although this evidence betrays the hopes of ILs to be ecologically safe reagents, they also suggest both nonspecific and specific mechanisms of IL biological action, which can potentially be used in the development of IL-based biochemical and pharmaceutical agents (see section 3.4).

3. IONIC LIQUIDS AS COMPONENTS OF DRUG FORMULATIONS

3.1. Problems of Conventional Drugs: Low Solubility, Limited Bioavailability, and Polymorphism

The efficiency of a drug depends strongly on its bioavailability, which, in the case of the human organism, relates directly to drug permeability and solubility. Because physiological accessibility of a drug often implies its dissolution in the body liquids, lower solubility results in lower dissolution and absorption rates. Therefore, higher doses are required for reaching a therapeutic effect.^{458–460} Limited water solubility and low dissolution rate are among the major problems of modern drug development, especially for drugs that are delivered into the organism orally, which is one of the most common and easy administration routes. It should be noted that dissolution and solubility are different properties of a drug: the solubility reflects the ability of a compound to dissolve in a given solvent, whereas dissolution is a process per se and is characterized by rate.

Pharmaceutical substances are often obtained in a crystal form, which leads to another complication: polymorphism. Thus, one crystalline chemical can exist as several polymorphs or pseudopolymorphs with different properties, for example, mechanical characteristics, stability, melting point, solubility, and bioavailability. Moreover, pharmaceutical polymorphs can undergo unpredictable interchanges, and their formation is difficult to control.^{458,461,462}

There are numerous approaches for improving drug formulations, and the methodology is being updated regularly. The existing methods may be divided into those that imply chemical modification of a drug and those that do not. The first ones suggest the formation of drug salts and derivatives, for example, prodrugs, whereas the second ones include crystal engineering techniques for controlled crystallization, commin-

tion for decreasing the particle size of a drug, preparation of hydrates and solvates, cocrystallization, micellization or dispersion, and employment of alternative solvents and cosolvents (Figure 17).^{231,459,460,463–467}

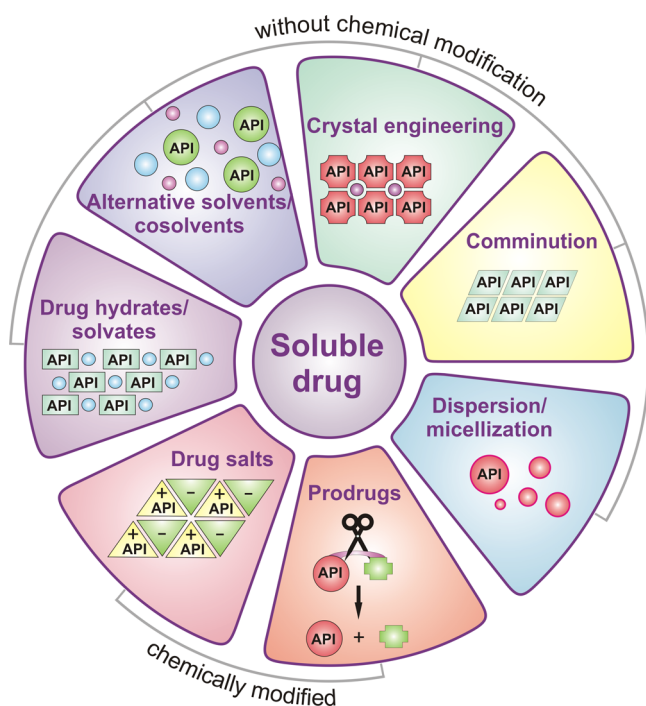


Figure 17. Main approaches to improve drug solubility: crystal engineering, comminution, preparation of hydrates and solvates, dispersion/micellization, and usage of alternative solvents/cosolvents do not imply chemical modifications of the drug structure, whereas formation of drug salts and prodrugs does.

Among the most popular strategies, decreasing the particle size of a drug is a very common approach for increasing its solubility and dissolution rate due to expansion of the surface area, which contacts with the solvent. The existing methods allow one to obtain nanoscale particles and nanocrystals.^{460,468}

Another common approach is disrupting the crystal lattice by using salts and cocrystals. If a drug is an ionizable substance, turning it into a salt is usually the easiest way to significantly increase its solubility and dissolution rate, although it should be remembered that soluble salts are often unstable and are subject to fast hydrolysis.⁴⁶⁹ The choice of a counterion may strongly influence the solubility and dissolution rate of a compound; thus, the solubility and dissolution rate of hydrochloride salts may drop in the stomach because of the presence of endogenous chloride ions (common-ion effect). Therefore, in some cases, basic drugs may demonstrate higher dissolution rates than salts at gastric pH.⁴⁶⁵

If a drug cannot be ionized, cocrystals may be used. There is no conventional definition of a cocrystal, but it can be described as crystalline matter, which includes at least two constituents. One of disadvantages of the cocrystal strategy is that upon dissolution the released free drug may form a supersaturated solution and may precipitate.⁴⁶⁴ Amorphous solid dispersions, which contain polymer carriers for stabilizing an amorphous drug, are also used for improving solubility and bioavailability of pharmaceutical agents; moreover, this strategy alleviates the problem of polymorphism.⁴⁶⁶ However, amorphous preparations are prone to spontaneous crystallization.⁴⁷⁰

Another way to increase the solubility of a drug is using solubilizing attachments, charged, or polar groups, which are added to the drug molecule so that its active part stays intact. The most popular solubilizing groups are amines, including cyclic ones, which can also be used for salt formation. If charged moieties are undesirable, amides or hydroxyl groups may be employed.⁴⁷¹

One of the latest and most promising strategies for improving solubility and bioavailability of medications is prodrugs. A prodrug is a chemically modified drug, from which the active substance is released in the organism. A good prodrug possesses negligible activity, but high water solubility and stability; it permeates through cell membranes efficiently, does not undergo hydrolysis during absorption, and releases the active part readily at the place of action. The prodrug strategy allows enhancing lipophilicity, solubility, stability and period of action of a parental pharmaceutical molecule, as well as relieving its toxicity and promoting its target delivery. There are two main categories of prodrugs: carrier-linked prodrugs and bioprecursors. The former consists of a drug part and a carrier, optionally linked via a spacer; the latter are inactive substances without carrier, which transform into an active substance in the organism.⁴⁷² In most cases, simple chemical or enzymatic changes are needed to turn a prodrug into an active drug. Depending on the functionality present in the parent drug molecule, various prodrug structures can be employed: esters, amides, oximes, and substances with disulfide bridges.^{472–475} Among the most interesting potentials of prodrugs is their employment in target delivery. A drug molecule may be conjugated with a specific antibody, peptide, or agonist.⁴⁷⁶ Prodrugs may selectively target cell membrane transporters in intestinal enterocytes and specific organs.^{474,477} The parent drug may be released by specific enzymes active only in specific organs/tissues.⁴⁷⁷ Several prodrug approaches have been suggested for “difficult cases”, such as surmounting the blood brain barrier and targeting tumors.^{476,478,479} Recently, a promising strategy on employing quaternary ammonium-based bioreversible linkages for connecting tertiary or heteroaryl amines, which are often present in therapeutic agents, to carrier proteins has been proposed as an instrument for target drug delivery. This strategy has also allowed significantly decreasing hydrophobicity of drugs upon conjugating with the linker, which can be proteolytically or reductively cleaved upon entering the target cell.⁴⁸⁰

Limitations of traditional drugs have led to the development of sophisticated encapsulation strategies for efficient delivery and prolonged release of active pharmaceutical ingredients at the destination. These strategies suggest using polymeric nanoparticles and micelles, lipid nanoparticles, liposomes, and nanotubes as colloid delivery carriers.^{481–483} The drug may be covalently linked to the nanoparticle or physically incorporated inside it, whereas amphiphilic drugs and prodrugs may self-assemble into nanoparticles without additional emulsifiers. The surface of a nanoparticle may bear specific targeting molecules, for example, antibodies or cell-penetrating peptides.^{478,484–486}

The cost of a drug is directly related to its ability to form polymorphs and to dissolve in the aqueous media. Nevertheless, despite obvious problems with solid drug formulations, the vast majority of medicinal substances are solids, whereas liquid formulations are very rare and are based on eutectic mixtures.²²³ Approximately one-half of modern drugs are salts.⁴⁸⁷ The development of pharmaceutically active “liquid salts” presents a good opportunity to alleviate polymorphism and solubility issues.

3.2. Benefits of Ionic Liquids in Drug Development

Considering numerous unique properties of ionic liquids, it is not surprising that they have drawn the attention of biomedical research not only as convenient catalytic media for drug synthesis, but also as potential components of drug formulations. To date, several notable reviews have emphasized the advantages of using ionic liquids in medicine chemistry.^{61,218,223,224,228–230}

Initial optimistic expectations for ILs to be “green”, environmentally friendly chemicals have been replaced by understanding that ILs possess high toxic potential, which may influence the environment significantly.²¹⁷ However, high biological activity of ILs may be seen not only as disadvantage, but also as opportunity. Several years ago, ionic liquid salts of active pharmaceutical ingredients were proposed to become an alternative to common crystalline salts.^{106,218,223,225}

What exactly makes ILs an interesting and beneficial object of study for medical reasons? As we have discussed in the previous sections, ILs are chemicals with an astounding fine-tuning potential: the number of possible IL combinations is about 10^{18} , and this is not the limit.¹⁶³ Thus, one may create an IL with practically any properties, including good water solubility, improved absorption, desired dissolution rate, and even targeting ability. Application of active pharmaceutical ingredients (API) in an ionic liquid form (API-ILs) also would solve the problem of polymorphism.^{223,228,230} Still, not every conventional drug can be turned into an ionic liquid salt, and new approaches for relieving the solubility and delivery issues are constantly demanded. Thus, apart from API-ILs, other possible applications of ILs in drug research and development include their employment as adjuvant agents for solubility enhancement and drug delivery.

In the following sections, we describe the recent progress in the application of ionic liquids in medicine formulations and delivery.

3.3. Ionic Liquids as Adjuvant Components in Drug Delivery

Striking solvent abilities of ionic liquids are widely known and are being exploited in extraction^{81,85,87,166,488–490} and dissolution of biomolecules.^{36,37,95,491–494} ILs can enhance water solubility of hydrophobic compounds, possibly due to formation of aggregates between ILs and biomolecules.⁴⁹⁵ Experimental data and molecular dynamics simulations suggest that upon dissolution in water, the continuous polar network, which is found in most pure ILs, disrupts into smaller domains; therefore, water solutions of ILs resemble a water matrix with incorporated ionic filaments (see also section 1.2). The anions in these filaments form hydrogen bonds with the water molecules, which stabilize the IL filamentous structure in the water solution. Addition of the model biomolecule vanillin to the system does not lead to the disappearance of the IL filaments, whereas vanillin molecules form cation–vanillin clusters via dispersion forces and other specific interactions, such as hydrogen bonds and π – π interactions, and mix with water more readily (Figure 18).⁴⁹⁵ Other studies support the correlation between excellent solubilizing properties of ILs and their ability to form numerous interactions with the solute.^{496–498} Thus, cholesterol underwent self-assembly and formed ordered mesoscopic structures in the nanostructured tetrabutylphosphonium carboxylate ILs; this strategy led to the enhancement of solubility of several model drug molecules.⁴⁹⁹

In the case of drugs, the solvent must not only dissolve the active pharmaceutical ingredient (API), but also prevent its precipitation and aggregation. Solubility of various API in various

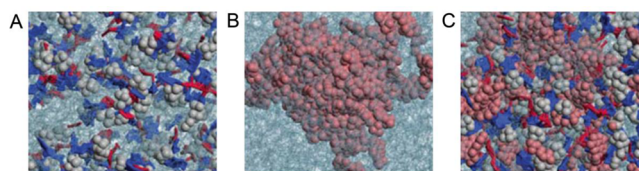


Figure 18. Simulation snapshots: (A) $[C_4MIM][N(CN)_2]$ + water; (B) vanillin + water; and (C) $[C_4MIM][N(CN)_2]$ + vanillin + water. Figures reproduced with permission from ref 495. Copyright 2015 Royal Society of Chemistry.

ionic liquids is a popular research subject, and employment of ILs for external⁵⁰⁰ and internal⁵⁰¹ drug delivery has been patented.

In general, ionic liquids may be used as (1) cosolvents, (2) emulsifiers, (3) copolymers, and (4) solvents or antisolvents for design of crystalline substances. Several dissolution, micellization, electrochemical, and spectroscopic studies on the behavior of drugs in the presence of IL solutions have been carried out.^{498,502,503} Attempts to decipher the dependence of the API solubility on the IL structure have been made, but have produced ambiguous results. It is clear that both IL anion and cation influence the solubility of a given API. In general, hydrophilic drugs show better solubility in hydrophilic ILs, whereas hydrophobic drugs prefer hydrophobic solvents. The length of alkyl chains in the imidazolium cation correlates with the solubility of drug molecules.^{504–507} However, the influence of the anion is complicated.⁵⁰⁴ Below we state the data on solubility of various API in various ILs, which have been accumulated so far. Examples of available studies on application of ILs as solubility enhancing and emulsifying agents are provided in Table 1. According to the table, ILs are applicable to broad classes of pharmaceutical substances, including antioxidants, anesthetics, anticancer drugs, antiviral and antimicrobial agents, anticoagulants, nonsteroidal anti-inflammatory drugs (NSAIDs), and others. Most studies are dedicated to imidazolium-based ILs; there are also several examples of using ammonium and phosphonium ILs.

N-Acetyl-L-cysteine and 4-hydroxycoumarin were readily soluble in hydrophilic ILs with short alkyl side chains in the cation, such as $[C_2MIM][OTf]$, whereas coumarin, 4'-isobutylacetophenone, thymoquinone, and ibuprofen demonstrated high solubility in the hydrophobic ILs $[C_{10}MIM][NTf_2]$, $[(C_6)_3C_{14}P][Cl]$, and $[(C_6)_3C_{14}P][NTf_2]$.^{505,508,509} Water-soluble acetaminophen and caffeine showed high solubility in $[C_4MIM][BF_4]$, $[C_8MIM][BF_4]$, and $[C_4MIM][PF_6]$, but not in $[C_8MIM][PF_6]$;⁵⁰⁴ the presence of $[C_6MIM][Br]$ or $[C_4MIM][Br]$ led to even higher water solubility of acetaminophen.^{510,511} Poorly water-soluble albendazole was soluble in $[C_4MIM][PF_6]$, $[C_6MIM][PF_6]$, and $[C_8MIM][PF_6]$, whereas danazol dissolved in $[C_4MIM][PF_6]$, $[C_8MIM][PF_6]$, and $[C_8MIM][BF_4]$; addition of a water-miscible IL to a water-immiscible one improved the water miscibility of the system.^{504,512} The IL consisting of choline and tryptophan enhanced solubility of glibenclamide.⁴⁹⁷ Solubility of isoniazid and pyrazine-2-carboxamide was higher in imidazolium ILs with the trifluoromethanesulfonate anion than in those with the bis-(trifluoromethylsulfonyl)amide one, and $[C_{10}MIM][OTf]$ was the best solvent. An increase in the alkyl chain length in the cation led to a decrease in the solubility.^{506,507} $[(C_{10})_2(C_1)_2N][NO_3]$ also proved to be a good solvent for isoniazid and, to a lesser degree, for pyrazine-2-carboxamide.⁵¹³

Table 1. ILs as Solubility Enhancers and Emulsifiers in Drug Delivery

drug system	activity	IL	ref
<i>N</i> -acetyl-L-cysteine	antioxidant	[C ₂ MIM][OTf], [(C ₆) ₃ C ₁₄ P][Cl], [(C ₆) ₃ C ₁₄ P][NTf ₂]	505,508,509
acetaminophen	analgesic	[C ₄ MIM][BF ₄], [C ₈ MIM][BF ₄], [C ₄ MIM][PF ₆], [C ₄ MIM][Br], [C ₆ MIM][Br]	504,510,511
acyclovir	antiviral drug	[C ₁ MIM][DMP]	529–531
albendazole	antiparasitic agent	[C ₄ MIM][PF ₆], [C ₆ MIM][PF ₆], [C ₈ MIM][PF ₆]	504,512
amphotericin B	antifungal agent	[C ₂ MIM][OAc], [C _n NH ₃][OAc] (<i>n</i> = 4, 6, 8), [<i>m</i> -PEG ₃₅₀ -NH ₃][OAc]	514
coumarin	anticoagulant	[C ₁₀ MIM][NTf ₂], [(C ₆) ₃ C ₁₄ P][Cl], [(C ₆) ₃ C ₁₄ P][NTf ₂]	505,508,509
4-hydroxycoumarin	anticoagulant	[C ₂ MIM][OTf]	505,508,509
curcumin	antioxidant, anti-inflammatory, antitumor agent	[C ₄ MIM][BF ₄]	527
danazol	steroid drug	[C ₆ C ₆ OCOPY][NTf ₂], [C ₆ C ₆ OCOPY][N(CN) ₂], [C ₄ MIM][PF ₆], [C ₈ MIM][PF ₆], [C ₄ MIM][BF ₄]	504,515
dantrolene sodium	muscle relaxant	[C ₁ MIM][DMP]	530
dehydroepiandrosterone	steroid hormone	[C ₄ MIM][PF ₆], [C ₆ MIM][PF ₆], [C ₈ MIM][PF ₆]	330
dexametasone	steroid drug	[C ₄ MIM][PF ₆], [C ₆ MIM][PF ₆], [C ₈ MIM][PF ₆]	330,577
diclofenac	NSAID	[C ₆ MIM][Br], [C ₁₂ MIM][Br], [C ₄ MIM][Br]	500,578
diltiazem	calcium channel blocker	DABCO-based ILs, [C ₁₀ C ₁ Mor][Br]	523
doxorubicin	antitumor agent	poly(ionic liquid- <i>co</i> - <i>N</i> -isopropylacrylamide)	544
etodolac	NSAID	[C ₄ MIM][PF ₆]	533
5-fluorouracil	antitumor agent	[C ₄ MIM][Br], [C ₄ MIM][PF ₆]	532,533
glibenclamide	antidiabetic drug	[Cho][Trp]	497
gramicidin	antibiotic (pentadecapeptide)	1,4-bis(3-dodecylimidazolium-1-yl) butane bromide	572
ibuprofen	NSAID	[(C ₆) ₃ C ₁₄ P][Cl], [(C ₆) ₃ C ₁₄ P][NTf ₂], [C ₂ MIM][NTf ₂]	508,509,560
ibuprofen	NSAID	[C ₁₂ MIM][Cl], formation of [C ₁₂ MIM][Ibu] was observed	496
ibuprofen	NSAID	[C ₄ MIM][Ibu]-containing ionogels	545
4'-isobutylacetophenone	precursor in ibuprofen synthesis	[(C ₆) ₃ C ₁₄ P][Cl], [(C ₆) ₃ C ₁₄ P][NTf ₂]	508,509
isoniazid	antituberculosis agent	[C ₁₀ MIM][OTf], [(C ₁₀) ₂ (C ₁) ₂ N][NO ₃]	506,513
itraconazole	antifungal drug	[C ₆ C ₆ OCOPY][NTf ₂], [C ₆ C ₆ OCOPY][N(CN) ₂], [<i>m</i> -PEG ₃₅₀ -NH ₃][OAc], [<i>m</i> -PEG ₃₅₀ -NH ₃][C _n COO] (<i>n</i> = 3, 5, 7, 9)	514,515
lidocaine hydrochloride	local anesthetic	[C ₁₂ MIM][Cl], [C ₁₄ MIM][Cl]	520
methotrexate	anticancer, anti-autoimmune disease agent	[C ₁ MIM][DMP]	530
nimesulide	NSAID	[C ₂ MIM][BF ₄], [C ₂ MIM][OTf], [C ₂ MIM][Ms]	536
penicillin V	antibiotic	[C ₄ MIM][PF ₆], [C ₆ MIM][PF ₆], [C ₈ MIM][PF ₆]	330
progesterone	steroid hormone	[C ₄ MIM][PF ₆], [C ₆ MIM][PF ₆], [C ₈ MIM][PF ₆]	330
pyrazine-2-carboxamide	antituberculosis agent	[(C ₁₀) ₂ (C ₁) ₂ N][NO ₃], [C ₁₀ MIM][OTf]	507,513
rutaecarpine	plant alkaloid	[C ₁₂ MIM][Br]	537
thymoquinone	antibiotic, antitumor agent	[(C ₆) ₃ C ₁₄ P][Cl], [(C ₆) ₃ C ₁₄ P][NTf ₂]	508,509

One of the most promising applications of ILs as pharmaceutical solvents is the development of dedicated ILs for dissolving “hard-case” API with poor water solubility. This concept has been proved empirically by using targeted structural tuning. Thus, ILs with tunable lipophilicity in one ion (alkylamine cation or fatty acid-based anion) and tunable hydrophilicity in the other (polyethylene glycol-based cation) were suggested to be used as solubility-enhancing agents and drug delivery systems (Figure 19).⁵¹⁴

Fine-tuning of the IL structure was employed for optimization of solubility of danazol and itraconazole. These poorly water-soluble drugs effectively dissolved in [C₆C₆OCOPY][NTf₂] and [C₆C₆OCOPY][N(CN)₂]. The solubility of itraconazole was shown to be 100 times higher in [C₆C₆OCOPY][NTf₂] and 500 times higher in [C₆C₆OCOPY][N(CN)₂], as compared to soybean oil.⁵¹⁵

The IL-based vesicles, micelles, and microemulsions attract significant attention as potential carriers of poorly soluble drugs.^{516–519} This strategy has been studied thoroughly, and IL-containing formulations have shown promising potential in topical drug delivery. Thus, lidocaine hydrochloride was shown to adsorb on the surface of the aggregates formed by the surface-active ionic liquids [C₁₂MIM][Cl] and [C₁₄MIM][Cl]; the drug

also modulated the aggregation behavior of the ILs, which assisted efficient drug delivery.⁵²⁰ The model formulation containing the IL formed by octanoic or isostearic acid and diisopropanolamine or triisopropanolamine demonstrated improved skin permeability together with prolonged release of a model drug, and caused no skin damage.⁵²¹ Oil-in-water and water-in-oil emulsions with [C₆MIM][Cl] or [C₄MIM][PF₆] possessed antimicrobial activity and enhanced the skin penetration of a model substance in vitro.³³³ IL-containing solid-in-oil nanodispersions demonstrated enhanced skin penetration and were suggested to be used for transcutaneous vaccination with hydrophilic macromolecules.⁵²² An addition of 1% w/w of DABCO-based or morpholinium ILs enhanced the transdermal transport of the calcium channel blocker diltiazem.⁵²³ Cholinium geranate was shown to be an effective permeation enhancer with antimicrobial activity and low toxicity toward epithelial cells.⁵²⁴ Cholinium IL-containing microemulsions demonstrated long-term stability at room temperature.⁵²⁵ Another possible application of cholinium-based ILs is extraction and partitioning of drugs, which has been demonstrated by the examples of phenacetin, ibuprofen, lidocaine, and indomethacin.⁵²⁶

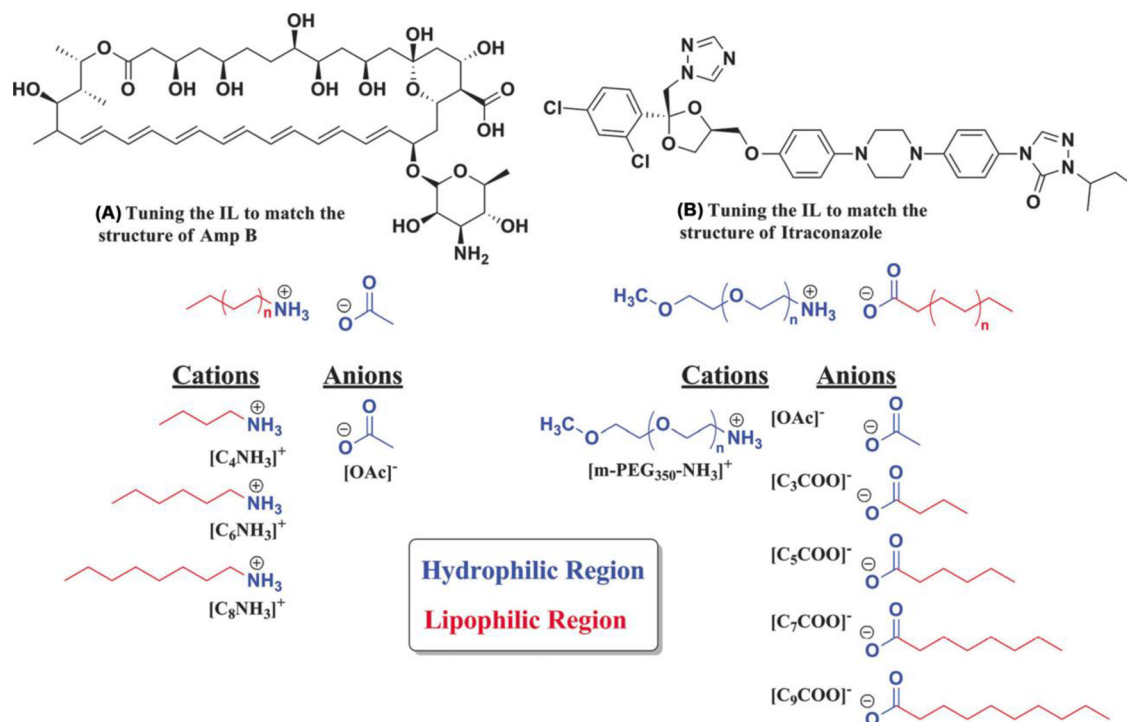


Figure 19. Ionic liquids as tunable solvents for (A) amphotericin B (cation hydrophobicity tuning) or (B) itraconazole (anion hydrophobicity tuning). Reproduced with permission from ref 514. Copyright 2013 Royal Society of Chemistry.

The hydrophilic IL $[C_4MIM][BF_4]$ affected micellization of various surfactant solutions of curcumin,⁵²⁷ whereas $[C_4MIM][C_8SO_4]$ stabilized curcumin, presumably via hydrogen bonds and strong hydrophobic interactions.⁵²⁸ Water-immiscible ILs demonstrated prolonged release of model drugs from IL microemulsions.³³⁰ A similar approach was employed for transdermal delivery of acyclovir and methotrexate.^{529–531} An ionic liquid-in-oil microemulsion containing $[C_4MIM][Br]$ or $[C_4MIM][PF_6]$ was developed for transdermal delivery of 5-fluorouracil or etodolac, respectively. The obtained formulations were used for in vivo treatment of skin cancer, arthritis, and inflammation in rodents, and produced promising results.^{532,533} The detailed role of IL ($[C_8MIM][Cl]$) and an IL-type imidazolium gemini surfactant in micellization was studied for the antidepressant amitriptyline hydrochloride.^{534,535}

$[C_2MIM][BF_4]$, $[C_2MIM][OTf]$, and $[C_2MIM][Ms]$ were shown to influence the interaction of nimesulide with human serum albumin (HSA); despite the increased lipophilicity of the system, nimesulide was able to bind to HSA efficiently.⁵³⁶ Addition of $[C_{12}MIM][Br]$ to a colloidal solution of rutaecarpine led to dissociation of the aggregates and changed their morphology.⁵³⁷ Similarly, $[C_4MIM][BF_4]$ modulated aggregation properties of the biological surfactant sodium deoxycholate.⁵³⁸ Interestingly, in a study on micellization and intermolecular interactions between $[C_{12}MIM][Cl]$ and ibuprofen, the formation of $[C_{12}MIM][Ibu]$ complexes was observed.⁴⁹⁶

Liquid-in-oil microemulsions containing double-chain surface-active ILs were described. $[C_1C_3Pyr][NTf_2]$ or $[C_6MIM][NTf_2]$ was used as polar core, whereas 1-butyl-3-methylimidazolium 1,4-bis(2-ethylhexyl) sulfosuccinate or *N,N*-dimethylethanolammonium 1,4-bis(2-ethylhexyl) sulfosuccinate was used as surfactant. These microemulsions demonstrated high thermostability, and the size of aggregates could be regulated by selection of appropriate polar constituents.^{539,540}

Similarly to polyion complex micelles formed by interactions between ionic copolymers and macromolecules of opposite charge, which are studied as potential drug carriers capable of targeted release of their load,^{541–543} polymeric ILs also find potential application in drug delivery. Nanoparticles from poly(ionic liquid-*co-N*-isopropylacrylamide) with deoxycholic acid were assembled via electrostatic interactions; by using doxorubicin as model drug, drug release and cytotoxic activity of the system were demonstrated.⁵⁴⁴ An IL-based hybrid material (ionogel) containing $[C_4MIM][Ibu]$ was suggested to be used as a novel drug delivery system.^{545,546} Other reported applications of gelled ILs include a self-polymerizing IL gel prepared from choline formate and 2-hydroxyethyl methacrylate; from this gel, curcumin-loaded nanoparticles were obtained.^{547,548} The synthesis of a polyethylenimine-based ionic liquid colloid for possible environmental and medical applications was described.⁵⁴⁹ A pH-sensitive ionogel from choline polyacrylate (polymeric IL) was efficiently employed for prolonged delivery of 5-fluorouracil.⁵⁵⁰ A temperature-sensitive *N*-isopropyl acrylamide-based copolymer doped with 1-butyl-3-vinylimidazolium bromide demonstrated high potential for drug entrapping and release, and low cytotoxicity.⁵⁵¹

Hexafluorophosphate and chloride salts of 1-(4-vinylbenzyl)-3-methylimidazolium and 1-(4-vinylbenzyl)-4-(dimethylamino)-pyridinium were used as monomers for synthesis of positively charged polymers, into which the anionic drug naproxen was loaded. Presumably, such complexes may pass acidic and neutral environment to be released in the intestine.^{552,553} Similarly, positively charged silica nanoparticles modified by pH-sensitive ILs were suggested to be employed for targeted delivery of naproxen⁵⁵⁴ and methotrexate.^{555,556} Positively charged silica nanoparticles modified with 4-acetyl-*N*-butylpyridinium hexafluorophosphate or 4-acetyl-*N*-propenylpyridinium hexafluorophosphate targeted mitochondria and inhibited proliferation of tumor cells in vitro.^{557,558} Gold

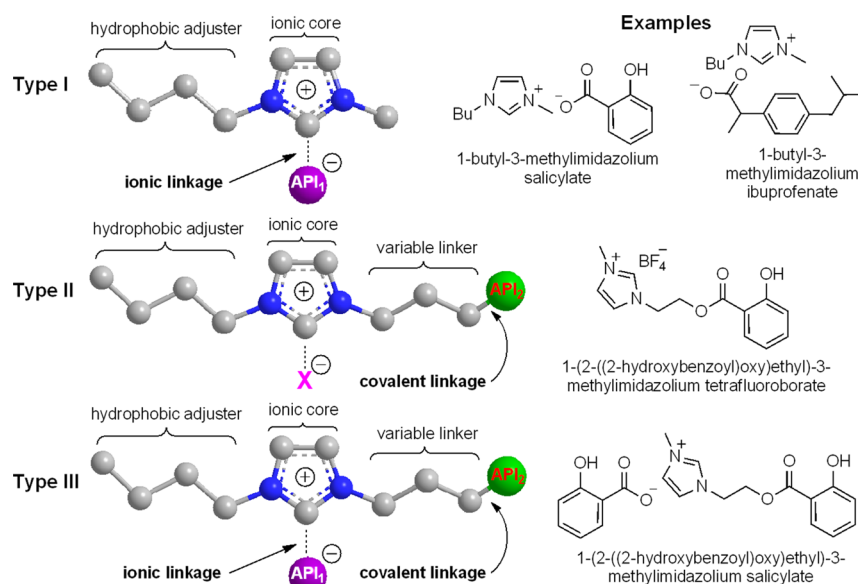


Figure 20. API-ILs containing ionic API as anion (Type I), covalently linked API in the cation (Type II), or combining both options (Type III); note that in the case of Type II, covalent binding to the anion is also possible (omitted for clarity). Figure reproduced with permission from ref 322. Copyright 2015 American Chemical Society.

nanoparticles grown on single-walled carbon nanotubes coated with 1-allyl-3-(3-mercaptopropyl)imidazolium chloride possessed low cytotoxicity and were able to enter lysosomes and to kill tumor cells by heat emission after laser irradiation.⁵⁵⁹

Another promising application of ILs in drug development is adjuvant components in crystallization, and Table 1 shows examples of such usage. Potential employment of ILs for cooling crystallization was suggested by investigating solubility of 12 API, including ibuprofen, acetylsalicylic acid, cyclosporine, and itraconazole, in $[\text{C}_2\text{MIM}][\text{NTf}_2]$. Ten of the API proved to be soluble in the IL at their melting point, whereas ibuprofen formed an immiscible phase. Products of cooling crystallization of acetaminophen in $[\text{C}_2\text{MIM}][\text{NTf}_2]$ were of substantial purity.⁵⁶⁰ Adjusting solubility of acetaminophen via manipulating hydrogen-bonding interactions between the drug and IL was proposed to be employed for drug crystallization.⁵⁶¹ ILs were applied for design of drug polymorphs via drowning-out crystallization: usage of $[\text{AEIM}][\text{BF}_4]$ led to the formation of new polymorphs of adefovir dipivoxil and thermostabilized the drug in the IL–water mixture.⁵⁶² $[\text{AEIM}][\text{BF}_4]$ was also used as antisolvent for studying polymorphic transformations of clopidogrel bisulfate.⁵⁶³ In another study, $[\text{AEIM}][\text{BF}_4]$ and $[\text{C}_4\text{C}_1\text{MIM}][\text{BF}_4]$ were employed as solvent and antisolvent, respectively, in antisolvent crystallization of adefovir dipivoxil. The authors demonstrated that different combinations of ILs produced different polymorphs.⁵⁶⁴ When $[\text{AEIM}][\text{BF}_4]$ and $[\text{C}_4\text{C}_1\text{MIM}][\text{BF}_4]$ were used in the solvent-mediated phase transformation method, a metastable polymorph of adefovir dipivoxil was transformed into a stable one.⁵⁶⁵ 1-Ethyl-3-methylimidazolium methyl phosphonate was employed for preparation of ultrafine particles of rifampicin, which showed good solubility in the IL. Addition of the rifampicin solution in the IL to an aqueous antisolvent produced amorphous submicrometer particles of the drug.⁵⁶⁶

In the end of the section, we would like to mention several interesting examples of indirect application of ILs in drug delivery. Thus, IL was employed as coating/drug carrier in the paclitaxel-coated balloon catheter, a probable alternative for

treatment of coronary and peripheral artery disease. Cetylpyridinium salicylate was used as matrix for drug transfer enhancement. This IL was suggested to meet all of the demands required for good performance of the balloon catheter: it formed a thin, smooth layer on the surface, provided efficient drug incorporation into the coating, and possessed high but postponed solubility in water.^{567–569}

Another interesting example is imidazolium poly(ionic liquid) membranes with the L-tryptophan anion; these membranes demonstrated high antimicrobial activity and induced no crucial hemolysis or cytotoxicity toward human cells.⁵⁷⁰

Polydopamine nanoparticles loaded with $[\text{C}_4\text{MIM}][\text{PF}_6]$ and doxorubicin were used as a successful chemotherapeutic and microwave thermal therapeutic agent for treatment of tumors in mice. Of note, the formulation induced no significant tissue toxicity. In this case, IL was employed as microwave sensitizer.⁵⁷¹

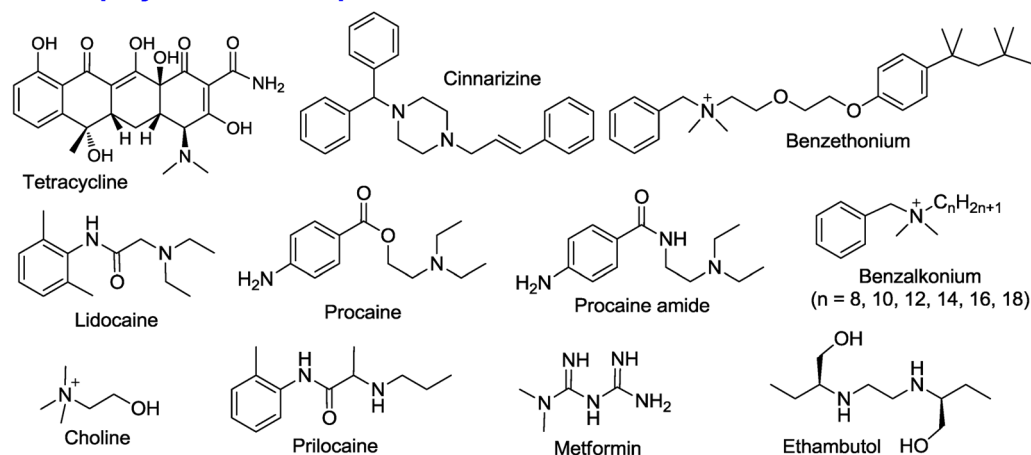
The IL-type gemini surfactant 1,4-bis(3-dodecylimidazolium-1-yl) butane bromide was shown to interact with gramicidin vesicles, therefore triggering a significant change in the conformation of the latter and suggesting a possibility of modulation of drug properties via interactions with such ionic surfactants.⁵⁷²

Finally, noncovalent complexes of cholinium-based amino acid ILs with hemocyanin from *Rapana thomasiana* demonstrated significantly improved selective activity toward tumor cells in vitro. The presence of IL caused rearrangements in the secondary structure of the protein.⁵⁷³ According to several reports, ILs impacted the structure of various amyloid peptides via interaction with specific amino acid residues; both promoting and inhibiting effects were observed.^{574–576}

3.4. Active Pharmaceutical Ingredient-Ionic Liquids (API-ILs)

3.4.1. Solubility and Physicochemical Behavior. As discussed in section 3.1, turning a drug into a salt is a common way to increase its solubility;⁴⁶⁹ therefore, because ionic liquids are liquid salts, turning a drug into an ionic liquid is an obvious method to improve its bioavailability and to relieve the problem of polymorphs. Moreover, it has been suggested that using active pharmaceutical ingredients in an ionic liquid form would allow

API employed in cationic part of ILs



API employed in anionic part of ILs

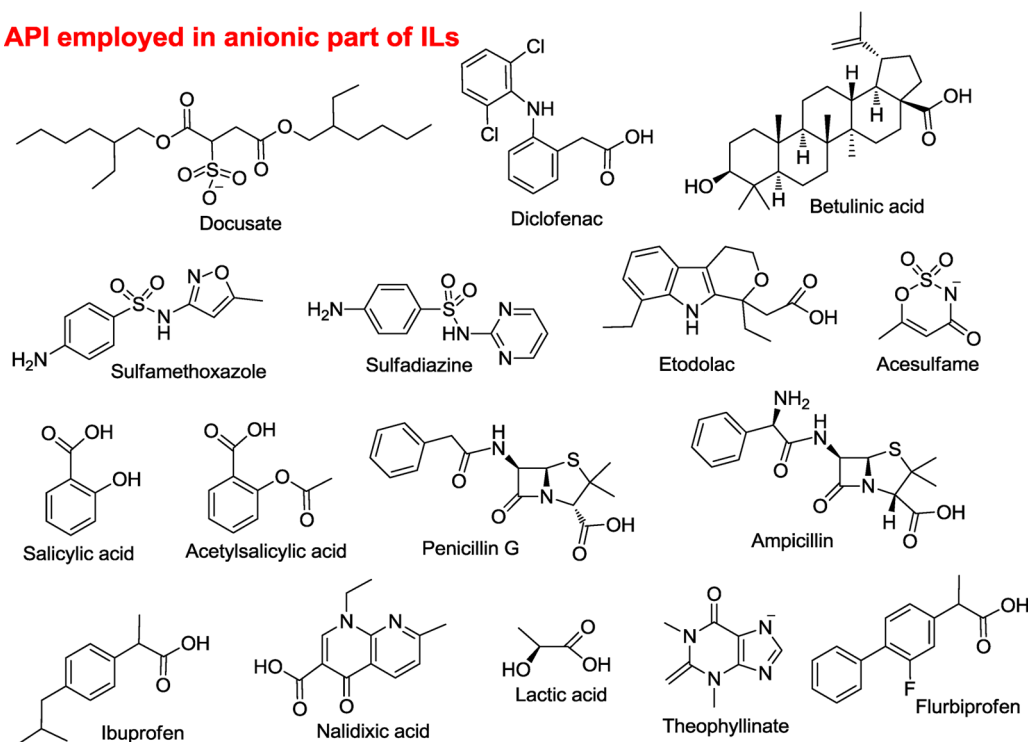


Figure 21. Examples of active pharmaceutical ingredients used in anions and cations of API-ILs (see Table 2 for details).

fine-tuning their physicochemical or biological properties, such as lipophilicity or toxicity.^{106,218,223,225,228} Self-assembled API-IL vesicles were proposed to be an efficient drug delivery system, where IL acted both as carrier and as drug.⁵⁷⁹ For neutral API, obtaining prodrugs via functionalization with ionizable groups was offered.⁵⁸⁰ API-ILs were called “the third evolution” of ionic liquids.²¹²

There are three ways to introduce API into IL systems (Figure 20): (1) Type I, via ionic binding (using API as anion or cation); (2) Type II, via covalent linkage; and (3) Type III, by using both ways to produce API-ILs with dual activities, where similar or different APIs may be combined in one IL.^{228,322} At the moment, the vast majority of the available API-ILs belongs to Type I, that is, contains readily ionizable API moieties, which can be used as IL anions or cations (Figure 21).

In 1998, the first ILs derived from a bioactive substance (miconazole) were reported.⁵⁸¹ Since then, many ILs bearing active pharmaceutical ingredients in the cation or anion have

been obtained (Figure 21). The preparation of various API-IL compositions with lidocaine, docusate, ibuprofen, procaine, and other drugs, including dual-active API-ILs, was patented.⁵⁸² An essential summary for the API-ILs investigated so far is presented in Table 2. The most studied physicochemical properties of API-ILs are thermal behavior and solubility in various media, including water, as well as surface activity and aggregation behavior.

As it has been anticipated, API-ILs are characterized by significantly lower temperatures of glass transition and liquid–liquid transition, in comparison with the corresponding parent drugs. Thus, ranitidinium docusate, a combination of a histamine H_2 receptor antagonist (ranitidine) and a laxative (docusate), is liquid at $T > 29\text{ }^\circ\text{C}$ ($T_g -12\text{ }^\circ\text{C}$), whereas T_g values of lidocainium docusate and didecyltrimethylammonium ibuprofen are -29 and -63 (-73) $^\circ\text{C}$, respectively.^{212,583} T_g of ranitidinium ibuprofenate and ranitidinium sulfacetamide was -12 and $25\text{ }^\circ\text{C}$, respectively; no melting point was found for

Table 2. Current Data on API-ILs and Their Properties^a

API-IL	studied physicochemical properties	studied biomedical properties	ref
[Tet][Doc]	thermal stability; water solubility; octanol–water and liposome–water partition coefficients	not studied	603
[(C ₁) ₄ N][Na]	basic characterization	antibacterial activity	623
[(C ₁) ₃ C ₂ N][Na]	basic characterization	antibacterial activity	623
[(C ₁) ₃ C ₃ N][Na]	basic characterization	antibacterial activity	623
[(C ₁) ₃ C ₁₀ N][Na]	basic characterization	antibacterial activity	623
[(C ₁) ₃ C ₁₂ N][Na]	basic characterization	antibacterial activity	623
[(C ₁) ₃ C ₁₆ N][Na], valproate	thermal stability; thermal properties; surfactant properties; viscosity; conductivity; solubility in various solvents and simulated biological fluid	antibacterial activity	601,623
[Cl ₁₀], [Na], pyrazinate, niflumate, 4-amino-salicylate, [Ibu], [Amp], ketoprofenate, naproxenate, sulfasalazine, betulinic acid - glycine, betulinic acid, [Sal], [Sac], [Acesulf]	solubility in various solvents and simulated biological fluids; thermal behavior; partition properties	in vitro cytotoxicity; antibacterial activity; HSA binding; metabolic stability; exposure kinetics; cell layer permeability; HIV-1 protease inhibition activity	583,585,589,596,599,604,624–626,640–642
[2-(methacryloyloxy)ethyl]trimethylammonium salicylate	copolymers with methyl methacrylate were synthesized, and their size distribution, morphology, and thermal properties were studied	not studied	611
[(C ₁₀) ₂ (C ₁) ₂ N]: [Ibu], [Acesulf], [Sulf], trans-cinnamate, Colawet MA-80, Fast Green FCF, piperacillin, penicillin G, [Sal], 2-(2,6-dichlorophenyl)amino benzenoacetate, N-[4-[(2-amino-1,4-dihydro-4-oxo-6-pteridinyl)methyl]amino]benzoyl]-L-glutamate, (S)-6-methoxy- α -methyl-2-naphthaleneacetate, [Bz], mandelate, 2-acetoxybenzoate, nicotinate, [Doc], [Lac], L-proline	thermal behavior; thermal properties	antibacterial activity; antifungal activity; feeding deterrent activity; acute oral toxicity in rats; skin irritation in rabbits	212,247,295,582,583,614,615
[(C ₁₁) ₂ (C ₁) ₂ N]: [Sac], [Acesulf]	thermal properties	not studied	582
[(C ₉) ₃ C ₁ N][Na]	basic characterization	antibacterial activity	623
[(C ₈) ₃ C ₁ N][Na]	basic characterization	antibacterial activity	623
[(C ₂) ₄ N]: [Amp], [Sal]	thermal properties; solubility in various solvents; partition properties	not studied	589,604,624,625,643
[(HOC ₂) ₃ NH][Sal]	crystal structure	antifungal activity	616
(2-hydroxyethyl)dimethylundecyloxymethylammonium, (2-acetoxyethyl)beptyloxymethyl-dimethylammonium, (2-acetoxyethyl)dodecyloxymethyl-dimethylammonium, (2-hydroxyethyl)cyclododecyloxymethyl-dimethylammonium benzoates	thermal properties	not studied	582
[tuammonium]heptane: [Sal], [Bz], [Bz-Bz], [Bz-Sal], gentisate	thermal properties; ionic conductivity; viscosity	membrane transport	608,609
cocodi(2-hydroxyethyl)methylammonium, oleyldi(2-hydroxyethyl)methylammonium, polyoxyethylene (15) (hydrogenated tallow)methylammonium, oleyltrimethylammonium, cocotrimethylammonium, dicotrimethylammonium, di(hydrogenated tallow alkyl)dimethylammonium, allylbenzyl-dimethylammonium, alkylbenzyl-dimethylammonium, dibenzyl-dimethylammonium, cyclohexyl-dimethylammonium, cyclohexyl-dodecyl-dimethylammonium theophyllinates	thermal properties; surface activity	feeding deterrent activity; antifungal activity; antibacterial activity	617,627
[Im][Sal]	basic characterization	in vitro cytotoxicity; toxicity toward <i>Artemia salina</i>	622
[C ₂ MIM][Na]	basic characterization	antibacterial activity	623
[C ₂ MIM]: [Sal], [Amp], [Na]	thermal properties; solubility in various solvents; partition properties; surfactant properties	ecotoxicity study (carboxylesterase assay, <i>Vibrio fischeri</i> assay); antibacterial activity; in vitro cytotoxicity; HSA binding	322,589,604,618,623–625,633

Table 2. continued

API-IL	studied physicochemical properties	studied biomedical properties	ref
[HOC ₂ MIM]: [Amp], [Ibu]	thermal properties; solubility in various solvents; partition properties; conductivity	antibacterial activity; in vitro cytotoxicity	589,598,604,624,625
[C ₂ OC ₂ MIM][Sal]	basic characterization	in vitro cytotoxicity; toxicity toward <i>Artemia salina</i>	622
[HOC ₃ MIM][Sal]	basic characterization	in vitro cytotoxicity; toxicity toward <i>Artemia salina</i>	622
[SalOC ₂ MIM]: [BF ₄], [Cl], [Sal]	thermal properties; water solubility	in vitro cytotoxicity	322
[SalOC ₃ MIM][Cl]	thermal properties; water solubility	in vitro cytotoxicity	322
[C ₆ MIM]: [Sal], chloramphenicol, sulfadiazine, sulfamethoxazol, fosfomycin, [Dic], [Ibu], [Na], [Sac], [Acesulf]	DFT studies; water solubility; thermal properties; viscosity; hydrolytic stability; corrosion test; friction and wear test; surface activity; conductivity; aggregation behavior; gelation; degradation	in vitro cytotoxicity; antibacterial activity; release kinetics; in vitro release toward <i>Artemia salina</i>	322,545,590,592–594,606,622,623,642,644–649
[HOC ₂ OC ₂ MIM][Sal]	basic characterization	in vitro cytotoxicity; toxicity toward <i>Artemia salina</i>	622
[C ₃ MIM][Na]	basic characterization	antibacterial activity	623
[C ₆ MIM]: [Sal], [Ibu]	water solubility; thermal properties; viscosity; hydrolytic stability; corrosion test; friction and wear test; surface activity; conductivity; aggregation behavior	in vitro cytotoxicity	322,592–594,623
[C ₈ MIM]: sulfadiazine, sulfamethoxazol, fosfomycin, [ASal], [Ibu], [Na]	thermal properties; viscosity; hydrolytic stability; corrosion test; friction and wear test; surface activity; electrical conductivity; aggregation behavior; self-encapsulation into mesoporous silica materials	antibacterial activity	590,592–594,623,650
[C ₁₆ MIM]: [Amp], sulfadiazine, sulfamethoxazol, fosfomycin	water solubility; thermal properties	antibacterial activity	277,590
[C ₁₆ C ₁ MIM][Amp]	water solubility; thermal properties	antibacterial activity	277
methimazole (1-methyl-2-(prop-2-en-1-yl)sulfanyl)-1H-imidazol-3-ium bromide	crystal structure	not studied	651
[C ₂ Py][Doc]	studies on incorporation into medical-grade PVC	antimicrobial activity	607
[C ₄ Py]: [Sal], [ASal], [Amp], [Sac], [Acesulf], [Sulf], sulfathiazole, Colawet MA-80, Fast Green FCF, piperacillin, penicillin G, valproate	thermal properties; solubility in various solvents; partition properties; crystal structure; surfactant properties	ecotoxicity study (carboxylesterase assay, <i>Vibrio fischeri</i> assay); antibacterial activity; in vitro cytotoxicity; HSA binding	247,277,296,567–569,582,587,589,604,618,624,625,628,633
[C ₈ OC ₂ HOPy]: [Sac], [Acesulf]	thermal properties; crystal structure	not studied	247
[C ₄ C ₂ Py][Na]	basic characterization	antibacterial activity	623
[HOC ₂ Py]: [Bz], [Bz-Bz], [Sal], [Bz-Sal], gentisate	thermal properties; ionic conductivity; viscosity	transmembrane transport	609
N-(2,3'-epoxypropyl)-N-methyl-2-oxopyrrolidinium salicylate	density; speed of sound	not studied	652
[C ₂ C ₁ Pip][Na]	basic characterization	antibacterial activity	623
[C ₁₀ C ₁ Pip][Theophyl]	thermal properties; surface activity	feeding deterrent activity; antifungal activity; antibacterial activity	617
[C ₁₂ C ₁ Pip]: [Theophyl], [ASal]	thermal properties; surface activity; aggregation behavior; viscosity	feeding deterrent activity; antifungal activity; antibacterial activity; release of acetylsalicylate	595,617
[C ₂ C ₁ Mor][Na]	basic characterization	antibacterial activity	623
[C ₁ (C ₄) ₃][Na]	basic characterization	antibacterial activity	623

Table 2. continued

API-IL	studied physicochemical properties	studied biomedical properties	ref
[C ₁ (C ₈) ₂ P][Na]	basic characterization	antibacterial activity	623
[(C ₉) ₄ P]: [Dic], [Ibu], [ASa], [Sal], [N-(7-isopropyl-6-(2-methyl-2H-pyrazol-3-yl)-2,4-dioxo-1,4-dihydro-2H-quinazolin-3-yl)-methanesulfonamide, keto-profenate, naproxenate, sulfadiazine, sulfamethoxazole, tolbutamide]	dissolution kinetics; saturation concentration; thermal properties	in vitro release from mesoporous support; release profiles; transmembrane transport; in vitro permeability through cell monolayer	586,588,597,605,610
[HOC ₂ (C ₉) ₃ P][Doc]	studies on incorporation into medical-grade PVC	antimicrobial activity	607
[(C ₉) ₃ C ₁₄ P]: [Sal], [Doc], [NTF], [Amp]	thermal properties; solubility in various solvents; partition properties	ecotoxicity study (carboxylesterase assay, <i>Vibrio fischeri</i> assay); antibacterial activity; in vitro cytotoxicity	589,604,624,625,633
hexetidinium : [Sal], [ASa]	thermal properties	not studied	587
tramadolium : [Sal], [ASa]	thermal properties; crystal structure	not studied	587
[Lid]: [Ibu], [Sal], [N-(oligomeric ions), [ASa], [Doc], [Dic], [Sac], [Acesulf], [Flurb], [Fluf], [HCl], [Lac], [Etodolac]	thermal properties, and behavior; water solubility; degree of ionicity; conductivity; simulated interactions in solution; molecular dynamics	in vitro release from mesoporous support and poly(lactic membranes; anti-noiception activity; neuritic outgrowth inhibition; local anesthesia; transmembrane transport; <i>Phase III clinical trials</i> ([Lid][Etodolac])	212,582,583,587,600,605,606,610,619,629,634,636,653–655
[Pril]: [Sal], [Doc], [Dic], [Sac], [Acesulf], [Flurb], [Fluf]	degree of ionicity	not studied	634
[Bupri]: [Sal], [Doc], [Dic], [Sac], [Acesulf], [Flurb], [Fluf]	degree of ionicity	not studied	634
[Proc]: [Sal], [ASa], [OAc], [HCl]	thermal properties; crystal structure; study on IL hydrates; structural dynamics in the glassy state	not studied	587,613,656
[ProcA]: [Sal], [ASa], HCl-(ZnCl ₂) ₂ , [HCl]	thermal properties; conductivity	not studied	587,620,657
[Metf]: [Sal], [Doc], [Dic], [Sac], [Acesulf], [Flurb], [Fluf]	degree of ionicity	not studied	634
ethambutol dibenzoate	first example of polymorphism in API-IL	not studied	612
ethambutol : adipate, HCl-(ZnCl ₂) ₂ , hydrochloride nalidixate, [Lac]	thermal properties	antimicrobial activity	296,620,658
acetaminophen prodrugs paired with the docusate anion	thermal properties; hydrolysis study	not studied	580
[Benzetf]: [Doc], [NTF], [Sal], [ASa], [Sac], HCl-(ZnCl ₂) ₂	thermal properties	ecotoxicity study (carboxylesterase assay, <i>Vibrio fischeri</i> assay); antibacterial activity	587,620,633,635
[Benzalk]: [Sal], [ASa], [Ibu], [Sulf], Colawet MA-80, [Sac], [Acesulf], [Theophyl], betulinic acid–glycine, betulinic acid, <i>trans</i> -cinnamate, Fast Green FCF, piperacillin, penicillin G, [Bz], sulfathiazole, [Doc], thimerosal, [Lac], 1-proline, mandelate, [NO ₃]	thermal properties; surfactant properties; partition properties	ecotoxicity study (carboxylesterase assay, <i>Vibrio fischeri</i> assay); antibacterial activity; antifungal activity; feeding deterrent activity; HSA binding; in vitro cytotoxicity; HIV-1 protease inhibition activity	218,247,295,582,587,599,614,615,617,618,626,633
miconazole derivatives paired with [PF ₆]	thermal properties	not studied	581
propantelinium : [Acesulf], <i>p</i> -toluenesulfonate	thermal properties	not studied	635
ranitidinium : [Doc], HCl-(ZnCl ₂) ₂ , [Ibu], [Sulf]	thermal properties; water solubility	antimicrobial activity; antioxidant activity; cytotoxicity	212,582,584,620
phenoxybenzamine-HCl -(ZnCl ₂) ₂	thermal properties	not studied	620
homatropine-HCl -(ZnCl ₂) ₂	thermal properties	not studied	620
nicaardipine-HCl -(ZnCl ₂) ₂	thermal properties	not studied	620
imipramine-HCl -(ZnCl ₂) ₂	thermal properties	not studied	620
Rhodamine-6G : tetraphenylborate, bis(perfluoroethylsulfonyl)imide	colloidal stability data	cytotoxicity toward tumor cells	621
cinnarizine : [C ₁₈ SO ₄], [C ₁₂ SO ₄], [C ₁₈ SO ₄], 7-ethyl-2-methyl-4-undecylsulfate, oleate, stearate, [NTF]	thermal properties; solubility; dispersion	plasma concentration	591

Table 2. continued

API-IL	studied physicochemical properties	studied biomedical properties	ref
<i>halofantrine</i> : [C ₁₀ SO ₄], [C ₁₂ SO ₄], oleate, [NTf ₂]	thermal properties; solubility; dispersion	not studied	591
<i>itraconazole</i> : [C ₁₂ SO ₄], 7-ethyl-2-methyl-4-undecylsulfate, [Doc]	thermal properties; solubility; dispersion	plasma concentration	591
[bromohexinium][Ibu]	basic characterization	transmembrane transport	608
[ephedrinium][Ibu]	thermal properties	transmembrane transport	610
verapamil hydrochloride	thermal behavior; conductivity; entropic models	not studied	659,660
[carvedol][H ₂ PO ₄]	structural dynamics in the glassy state	not studied	656
[amitriptyline][C ₁₂ SO ₄]	surface activity; aggregation behavior	hemolytic toxicity; drug release	579
[mepenzolate][Doc]	thermal properties	not studied	582
domiphen mandelate	basic characterization	antibacterial activity; antifungal activity	615
proline ethylester ibuprofenate	basic characterization	skin permeation	631
<i>ketokonazole</i> : citrate, tartrate	thermal properties; viscosity; solubility;	not studied	602
<i>diphenhydraminium</i> : [Ibu], [Doc]	thermal properties; water solubility; ionicity	antimicrobial activity; antioxidant activity; cytotoxicity	584
<i>glycinium, ethylglycinium</i> : [Doc]	thermal properties; water solubility; ionicity	antimicrobial activity; antioxidant activity; cytotoxicity	584

^aFor convenience, API-ILs are classified by cations, and dual-active API-ILs are shown in bold.

these API-ILs. Similarly, diphenhydraminium docusate, glycinium docusate, and ethylglycinium docusate possessed no melting point, and their T_g values were below 0 °C.⁵⁸⁴ T_m of cholinium nalidixate is -4.7 °C (205 °C for nalidixate).⁵⁸⁵ Tetrabutylphosphonium ibuprofenate, tetrabutylphosphonium ketoprofenate, and tetrabutylphosphonium naproxenate are liquid at room temperature.⁵⁸⁶ A wide panel of salicylate- or acetylsalicylate-containing dual-active API-ILs with tetrabutylphosphonium, cetylpyridinium, benzalkonium, benzethonium, lidocainium, procainium, and other counterions demonstrated relatively low T_g and T_m , together with acceptable thermostability.⁵⁸⁷ Lidocainium salicylate and tetrabutylphosphonium salicylate were used for the preparation of oligomeric ILs with various thermal properties.⁵⁸⁸ Ampicillin-bearing ILs with ammonium, pyridinium and imidazolium as cations, as well as 1-alkyl-3-methylimidazolium ILs bearing sulfadiazine, sulfamethoxazol, acetylsalicylate, and other API as anion, possessed low T_g .^{589,590} The same was observed for API-ILs combining cinnarizine, halofantrine, and itraconazole with decylsulfate, dodecylsulfate, oleate, and other anions.⁵⁹¹ 1-Alkyl-3-methylimidazolium ibuprofenates showed high thermal and hydrolytic stability, together with good tribological properties, and were proposed to be used as lubricants.⁵⁹² Surfactant properties of these ILs suggested the possibility to modulate their aggregation via changing the alkyl side chain length.^{593,594} 1-Butyl-3-methylimidazolium ibuprofenate was employed for the preparation of ionogels, which were shown to be a promising drug delivery system.⁵⁴⁵ Similarly, studies on aggregation behavior of 1-dodecyl-1-methylpiperidinium acetylsalicylate and anion release from gels formed by this IL suggested its possible pharmaceutical application. Interestingly, small [C₁₂C₁Pip]-[ASal] aggregates transformed into gel-like wide micellar networks upon addition of sodium salicylate.⁵⁹⁵

The prodrug-IL concept was tested on a series of acetaminophen-based prodrugs with the docusate anion. The obtained prodrug-ILs demonstrated low T_g and underwent fast hydrolysis in simulated body fluids.⁵⁸⁰

Another anticipated advantage of API-ILs is improved solubility, in comparison with parent API. Thus, in the case of cholinium nalidixate, water solubility was almost 5000 times higher than that of the parent drug, whereas in the case of cholinium niflumate it was 56 000 times higher.⁵⁸⁵ Cholinium sulfasalazin demonstrated 4000-fold improved saline solubility and increased exposure, in comparison with the parent sulfalazine.⁵⁹⁶ Tetrabutylphosphonium API-ILs showed improved dissolution rates, as compared to the parent drugs diclofenac, ibuprofen, naproxen, and others.^{586,597} 1-(2-Hydroxyethyl)-3-methylimidazolium ibuprofenate showed very high water solubility (150 000 times higher than the pure ibuprofen),⁵⁹⁸ whereas 1-alkyl-3-methylimidazolium salicylates were significantly more water-soluble than the pure salicylic acid.³²² Water solubility of ionic derivatives of betulinic acid was improved up to 100 times, in comparison with the parent drug.⁵⁹⁹ Both lidocaine and etodolac in the form of lidocainium etodolac showed higher water solubility than either drug alone.⁶⁰⁰ Trimethylhexadecylammonium valproate demonstrated enhanced solubility in the simulated gastric fluid.⁶⁰¹ Ketokonazole citrate and tartrate showed improved solubility in the phosphate buffer, in comparison with the pure ketoconazole (antifungal drug) or its physical mixtures with citric or tartaric acid.⁶⁰²

The solubility of API-IL can be modulated by varying the counterion hydrophilicity/hydrophobicity. Thus, tetracycline

docusate demonstrated significantly lower water solubility than tetracycline hydrochloride or sodium docusate, but showed preferential partitioning toward a lipophilic phase.⁶⁰³ Ampicillin-bearing ILs were miscible with various solvents, depending on the cation.^{589,604} API-ILs combining cinnarizine, halofantrine, and itraconazole with decylsulfate, dodecylsulfate, oleate, and other anions demonstrated improved solubility in lipid-based emulsions.⁵⁹¹

Tetrabutylphosphonium ibuprofenate and lidocainium ibuprofenate were stable when immobilized on mesoporous silica, and were readily released from the support.⁶⁰⁵ Lidocainium ibuprofenate also acted as plasticizer in poly(L-lactic acid) membranes.⁶⁰⁶ 1-Ethylpyridinium docusate and tri-*n*-butyl(2-hydroxyethyl)phosphonium docusate were employed as both plasticizers and antimicrobial agents for medical-grade polyvinyl chlorides.⁶⁰⁷

Several API-ILs demonstrated rapid transport through a model membrane, possibly in the form of hydrogen-bonded clusters (see also section 1.2).⁶⁰⁸ The authors suggested that lower ionicity may be an advantage for API-ILs and may facilitate their trans-membrane transfer.⁶⁰⁹ This suggestion was confirmed by the study of simultaneous transport of lidocaine and ibuprofen through the membrane, where the formation of a hydrogen-bonded complex of the two API occurred in the solution.⁶¹⁰

Aggregate structures formed by ILs with surface-active API were studied by the example of 1-alkyl-3-methylimidazolium ibuprofenate. In the case of the cations with short alkyl chains, the micelles contained mainly the ibuprofenate anion; upon increasing the alkyl chain length, the amount of imidazolium cations in the micelles increased until aggregates with stoichiometric composition were formed; these aggregates interacted with each other giving globular conglomerates.^{593,594} Similarly to common ILs, API-ILs were suggested to be used for preparation of polymers with the ability to exchange biologically active ions ([2-(methacryloyloxy)ethyl]trimethylammonium salicylate was used as a building block of such a copolymer).⁶¹¹

API-ILs look to be a very promising pharmaceutical strategy; nevertheless, some drawbacks of the API-ILs concept should also be mentioned. The API-IL strategy cannot be considered the ultimate solution of the polymorphism problem, because polymorphs of API-ILs have been described. Ethambutol dibenzoate was found to form three polymorphs, which demonstrated close T_m (93–96 °C), but different thermal stability.⁶¹² High hygroscopicity of many ILs also may be a problem. Thus, procainium acetate formed a dihydrate, which underwent irreversible crystallization.⁶¹³ Crystallization and glass transition of 1-(2-hydroxyethyl)-3-methylimidazolium ibuprofenate showed high sensitivity to the water content.⁵⁹⁸ These findings highlight the importance of studying the formation of polymorphs and hydrates even in the case of API-ILs.

3.4.2. Biomedical Activity. Despite numerous reports on the preparation and physicochemical properties of API-ILs, their biomedical activity is significantly less studied, and the available data have been obtained mostly using in vitro models (see Table 2). In this section, we describe the current status of the knowledge on biomedical properties of API-ILs. Most studies available so far are dedicated to either physicochemical or biological properties of API-ILs, and only about two dozen works address both aspects (see, e.g., refs 212,247,277,322,545, 579,584,585,591,595–597,599,600,606,607,609,610,614–621). Usually these studies concern surface properties and the

enhancement of solubility of API-ILs, with respect to their biological activity or ability to penetrate biological membranes.

Increased water solubility suggests that API-ILs should demonstrate higher bioavailability than poorly soluble parent drugs. However, effective doses of many conventional solid drugs are very low, and their main issue is rather polymorphism, which is difficult to control. Moreover, the impact of incorporating API into IL is not always obvious. Therefore, an ideal API-IL should be liquid, so not to form polymorphs, and should retain the level of activity exhibited by the parent drug.

The influence of the drug transition from the solid to ionic liquid form was studied for several APIs. Thus, the introduction of salicylic acid into an imidazolium-type IL via an ionic or covalent linkage did not disturb the cytotoxic activity of the API: API-ILs of all three types (see Figure 20) exhibited cytotoxicity similar to that of the pure drug.³²² As expected, oxygenation of the alkyl side chain of the imidazolium cation led to significant reduction of toxicity exhibited by salicylate-containing ILs toward cell cultures and the brine shrimp *Artemia salina*.⁶²²

Usage of cholinium as counterion did not perturb the cytotoxicity of nalidixic, niflumic, and pyrazinoic acids.⁵⁸⁵ When studying ILs with the nalidixate anion, no improvement of antimicrobial activity against *Salmonella* was observed; however, according to this study, ILs with different cations showed different modes of toxic action.⁶²³ Imidazolium- and cetylpyridinium-based ILs bearing ampicillin demonstrated significantly lower water solubility than sodium ampicillin, but their antibacterial activity was improved.^{277,624} Moreover, these API-ILs also showed pronounced inhibitory activity toward tumor cell lines, but not normal fibroblasts, in vitro.⁶²⁵ Ionic derivatives of betulinic acid, a plant compound possessing antitumor properties, demonstrated enhanced water solubility, together with enhanced inhibition of HIV-1 protease and cytotoxicity against various tumor cell lines.^{599,626} 1-Alkyl-3-methylimidazolium ILs bearing sulfadiazine, sulfamethoxazol, acetylsalicylate, and other API as anion, as well as the chlorometallate-based IL benzethonium-Cl-(ZnCl₂)₂, possessed antibacterial and antibiofilm activity.^{590,620} Surface-active theophyllinate-containing ILs showed a promising fungicidal and bactericidal potential.^{617,627} Triethanolammonium salicylate inhibited growth of the fungus *Rhizopus oryzae*.⁶¹⁶ Ranitidinium ibuprofenate and diphenhydraminium ibuprofenate demonstrated activity against various *Candida* species, and the union of ethylglycine and docusate, which did not possess antibacterial activity in their pure forms, produced ethylglycinium docusate, which demonstrated activity against penicillin- and methicillin-resistant *Staphylococcus aureus*.⁵⁸⁴

Using rhodamin 6G-based IL-like compounds, a cooperative effect of both ions on antitumor activity was demonstrated.⁶²¹ Employment of cetylpyridinium salicylate, which itself possessed antibacterial activity, as dispersing agent for silver nanoparticles allowed one to produce nanoformulations with a broader antibacterial effect.⁶²⁸

Unsurprisingly, biological activity of API-ILs turned out to depend on the ion structure. Thus, the cation nature influenced antibacterial activity of dual-active API-ILs combining an antibacterial agent (didecyldimethylammonium, benzalkonium, cetylpyridinium, or 3-hydroxy-1-octyloxymethylpyridinium) with an artificial sweetener (acesulfame or saccharinate).²⁴⁷ Several dual-active API-ILs were tested against bacterial species involved in biofouling. Ethambutol hydrochloride nalidixate and cetrimonium nalidixate demonstrated excellent antibacterial properties; however, when ethambutol was combined with the

lactate anion, the resulting IL showed weak antibacterial activity, possibly because the bacteria utilized lactate as source of carbon.²⁹⁶ On the other hand, didecyldimethylammonium and benzalkonium D,L- and L-lactates were recognized as effective antimicrobial agents.⁶¹⁴

Lidocainium docusate provided a longer and more pronounced analgesic effect than lidocainium hydrochloride,²¹² possibly due to association of the ions in the aqueous media, which led to a synergistic impact on the activity;⁶²⁹ lidocainium ibuprofen was also efficiently applied for dermal anesthesia in rats.⁶¹⁹ Lidocainium etodolac demonstrated higher skin permeation than etodolac alone, due to improved lipophilicity/hydrophobicity of the drug.⁶⁰⁰ Tetrabutylphosphonium N-[7-isopropyl-6-(2-methyl-2H-pyrazol-3-yl)-2,4-dioxo-1,4-dihydro-2H-quinazolin-3-yl]-methanesulfonamide penetrated a cell monolayer more efficiently than the free drug (α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor antagonist).⁵⁹⁷ For API-ILs combining cinnarizine, halofantrine, and itraconazole with decylsulfate, dodecylsulfate, oleate, and other anions, enhanced drug absorption was observed in rats.⁵⁹¹ An external preparation containing fatty acid-based ILs for enhanced transdermal penetration has been patented.⁶³⁰ Implying biocompatible nature of amino acids, proline ethylester was suggested to be used as counterion to ibuprofenate; the resulting IL, proline ethylester-ibuprofenate, showed significantly improved transdermal permeation, as compared to a saturated ibuprofen solution.⁶³¹

According to an interesting example of antimicrobial peptides conjugated with one or multiple instances of imidazolium cations, the presence of these cations produced synergistic effects, which correlated with the cation number.⁶³²

In addition to biomedical properties of API-ILs, their environmental impact also should not be forgotten. Bioassays based on inhibition of human carboxylesterase 2 and *Vibrio fischeri* were suggested to assess the ecotoxicity of selected API-ILs bearing 1-ethyl-3-methylimidazolium, benzethonium, benzalkonium, cetylpyridinium, or trihexyltetradecylphosphonium as cation and salicylate, docusate, or bis(trifluoromethylsulfonyl)amide as anion. According to this study, 1-ethyl-3-methylimidazolium salicylate may be considered practically harmless, benzalkonium salicylate and cetylpyridinium salicylate were slightly toxic, whereas benzethonium salicylate and benzethonium docusate demonstrated moderate aquatotoxicity.⁶³³ Didecyldimethylammonium saccharinate and didecyldimethylammonium acesulfame exhibited significant oral toxicity and skin irritation.²⁴⁷

3.4.3. Combining Dual Activities within One Molecule.

The idea of combining two (or more) activities within one drug molecule seems very tempting.²²⁸ The role of the API-IL counterion should not necessarily be limited by preclusion of crystallization; it can also possess its own biomedical activity, thus providing API-IL with dual function or giving some new synergistic active combination not achievable with two separate APIs. We should note that it is not always easy to distinguish API-ILs with one activity from those with two or even more; nevertheless, we tentatively show most apparent examples of dual-active API-ILs in bold in Table 2. Undoubtedly, the targeted combining of ions, which possess several desired activities, within one IL has been attracting more attention lately.^{296,582,587,618,620,634,635} Here, we discuss several interesting examples.

The synthesis and behavior of dual-active API-ILs have been reported.^{582,587} Antibacterial cations, such as didecyldimethy-

lammonium, benzalkonium, and others, were united with the "sweet" anions saccharinate and acesulfame. Some of these dual-active API-ILs demonstrated improved antimicrobial activity. $[(C_{10})_2(C_1)_2N][Sac]$ and $[(C_{10})_2(C_1)_2N][Acesulf]$ also exhibited good deterrent activity against insects, but caused oral toxicity and skin irritation.²⁴⁷

Several attempts to combine the antibiotic ampicillin with the antiseptic cetylpyridinium have been made. The resulting API-IL $[C_{16}Py][Amp]$ demonstrated significantly higher activity against several Gram-positive and Gram-negative bacterial strains, as compared to the parent $[Na][Amp]$ or $[C_{16}Py][Cl]$.^{277,624} Among the API-ILs tested, $[C_{16}Py][Amp]$ possessed the highest potential for reversion of resistance of ampicillin-resistant clinical strains.⁶²⁴ Interestingly, $[C_{16}Py][Amp]$ also inhibited growth of some tumor cell lines more efficiently than $[Na][Amp]$.⁶²⁵

In search for agents against microbial corrosion, the antituberculous drug ethambutol and the antiseptic cetrimonium were combined with the antibiotic nalidixic acid. The resulting ethambutol hydrochloride nalidixate and cetrimonium nalidixate demonstrated enhanced antibacterial activity against bacteria found in marine biofilms on steel constructions.²⁹⁶

Lewis-acid ILs containing metal chlorides are an interesting example of an attractive pharmaceutical formulation. When the antiseptic benzethonium was combined with $ZnCl_2$, which also possessed antimicrobial properties, the resulting IL benzethonium-Cl-($ZnCl_2$)₂ demonstrated improved antibacterial activity, in comparison with benzethonium chloride.⁶²⁰

The combination of the antibiotic tetracycline with the laxative docusate produced an API-IL, which showed rather poor water solubility, in comparison with tetracycline hydrochloride and sodium docusate. However, $[Tet][Doc]$ demonstrated a higher partition coefficient in liposomes, possibly due to more intense interactions with the phosphatidylcholine bilayer, suggesting better pharmacokinetics.⁶⁰³ When docusate was used as counteranion for lidocaine, lidocainium docusate possessed higher bioavailability;²¹² this effect may be explained by improved membrane permeability due to formation of ionic associates near the cell membrane.⁶²⁹ Similarly, lidocaine and ibuprofen were shown to pass the membrane together, as a hydrogen-bonded complex,⁶¹⁰ and were effectively applied as local anesthetic.⁶¹⁹

Etodolac is a nonsteroidal anti-inflammatory drug commonly used for relieving pain and inflammation caused by rheumatoid arthritis and osteoarthritis. To improve its transdermal penetration, etodolac was combined with the local anesthetic lidocaine. The resulting API-IL lidocainium etodolac demonstrated significantly higher saturation solubility in water than etodolac alone; solubility of lidocaine in the IL form was also improved, in comparison with the parent drug. As part of API-IL, etodolac was transferred through the skin more efficiently; however, the opposite was observed for lidocaine.⁶⁰⁰ Of note, lidocainium etodolac has recently entered clinical trials, which are described in the following section.

3.4.4. Clinical Application. It seems that at the moment, lidocainium etodolac^{600,636} is the only API-IL that has reached clinical trials. In 2013, the results of the phase I trial of the MRX-7EAT (Etoreat) Etodolac-Lidocaine Topical Patch were published by IL Pharma Inc. (MEDRx).⁶³⁷ Usage of etodolac in an IL form was found to increase its skin absorption, whereas adverse effects were mild to moderate. The phase II/III trials on efficacy and safety of MRX-7EAT in the pain treatment have been conducted.⁶³⁸ Several trials on using MRX-7EAT in the treatment of ankle sprains (NCT01198834), low back pain

Table 3. Application of ILs in the Synthesis of Drugs and Their Intermediates

compound	activity	IL	role of IL	ref
(<i>R,S</i>)-ibuprofen	NSAID ^{a†}	[C ₆ C ₁ Py][BF ₄]	media for enzymatic enantioselective esterification or hydrolytic resolution	666,667,714,715
iodoquinol	antiprotozoal drug	[C ₆ C ₁ Py][Cl ₂]	iodinating reagent	668
clioquinol	antifungal drug	[C ₄ C ₁ Py][Cl ₂]	iodinating reagent	668
pravastoline	NSAID	[C ₄ MIM][PF ₆]	media	665
(<i>R</i>)-modafinil	wakefulness-promoting agent	[C ₂ MIM][Br]	media for enzymatic enantioselective esterification	716
stavudine	anti-HIV [†] drug	[C ₁ OC ₂ MIM][Ms], [C ₁ OC ₂ MIM][TFa], [C ₄ MIM][TFa]	media	670
brivudine	anti-HSV [†] drug	[C ₁ OC ₂ MIM][Ms], [C ₁ OC ₂ MIM][TFa], [C ₄ MIM][TFa]	media	670
trifuridine	anti-HSV drug	[C ₁ OC ₂ MIM][Ms], [C ₁ OC ₂ MIM][TFa], [C ₄ MIM][TFa]	media	670
(<i>S</i>)-naproxen	NSAID	[C ₄ MIM][BF ₄]	media	664
tioconazole	antifungal drug	[AlkMIM][Br]	catalyst	717
hydrocortisone	steroid hormone	[C ₁ MIM][BF ₄], [C ₂ MIM][BF ₄], [C ₄ MIM][PF ₆], [(C ₁) ₄ N][BF ₄]	catalyst	718
hydrazinyl phthalazines	antimalarial agents	[C ₁ C ₁ MIM][C ₂ SO ₄]	dehydrating agent	687
ciclesonide	anti-asthmatic, anti-allergic drug	acidic ILs	catalyst, media	719
α -tocopherol succinate	vitamin E ester	[C ₃ MIM][NO ₃]	catalyst, media	720
modafinil and its derivatives	stimulant for treatment of sleep disorders	[C ₄ MIM][PF ₆]	media	721
isoxazolines	precursor of antimicrobial agents	[C ₄ MIM][BF ₄]	media	722
1-[(<i>E</i>)-3-methyl-4-benzenesulfonyl-3-methylbut-2-enyl]-2,3,4,5-tetramethoxy-6-methylbenzene	coenzyme Q ₁₀ intermediate	[C ₂ Py][BF ₄], [MCl _m] (MCl _m = AlCl ₃ , FeCl ₃ , ZnCl ₂ , SnCl ₄ , SnCl ₂ , CuCl)	catalyst	723
(<i>S</i>)-3-chloro-1-phenyl-1-propanol	precursor of antidepressant drugs	[C ₄ MIM][NTF ₅]	IL phase, whole-cell catalysis	724
(<i>R</i>)-phenylacetylcarbinol	precursor of (1 <i>R</i> ,2 <i>S</i>)ephedrine and (1 <i>S</i> ,2 <i>S</i>)pseudoephedrine	[C ₄ MIM][PF ₆]	IL phase, whole-cell catalysis	725
3-phenylglycidol	precursor of various drugs, e.g., tomozetone and reboxetine	[C ₄ MIM][NTF ₅]	media	726
(<i>R,S</i>)-1-chloro-3-(3,4-difluorophenoxy)-2-propanol	intermediate in the synthesis of (<i>S</i>)-labeledzole	[C ₄ MIM][PF ₆], [C ₄ MIM][BF ₄]	media for enzymatic enantioselective esterification	727
thiazolidine derivatives, including rhodamine ones	antimicrobial agents, intermediates for preparation of antidiabetic drugs, other activities	[(C ₁) ₂ (C ₁) ₂ G][Lac], [(C ₁) ₂ (C ₁) ₂ G][OAc], [(C ₁) ₂ (C ₁) ₂ G][TFa], [(C ₁) ₂ (C ₁) ₂ G][P ₁], [(C ₁) ₂ (C ₁) ₂ G][P ₂], [(C ₁) ₂ (C ₁) ₂ G][P ₃], [(C ₁) ₂ (C ₁) ₂ G][P ₄], [(C ₁) ₂ (C ₁) ₂ G][P ₅], [(C ₂) ₃ NH][H ₂ SO ₄]	catalyst, media	685,686,712,713,728
11 α -hydroxy-16 α ,17-epoxyprogesterone	intermediate in the synthesis of steroidal drugs	[C ₄ MIM][PF ₆]	media for enzymatic enantioselective esterification	729
(<i>R</i>)-1-trimethylsilylethanol	key synthon for various silicon-containing drugs	[C ₄ MIM][PF ₆]	IL phase, whole-cell catalysis	730
dronedarone intermediate	antitachycardic arrhythmia drug	various imidazolium and pyridinium ILs	media	731
5-(α -haloacetyl)-8-substituted oxy-(1 <i>H</i>)-quinolin-2-ones	intermediates of 5-[(<i>R</i>)-2-(<i>S</i>)-6-diethyl-indan-2-ylamino]-1-hydroxy-ethyl]-8-hydroxy-(1 <i>H</i>)-quinolinone-2-one salts (bronchodilators, anti-asthmatic drugs)	various imidazolium ILs	media	732

^{a†}NSAID, nonsteroidal anti-inflammatory drug; HIV, human immunodeficiency virus; HSV, herpes simplex virus.

(NCT01968005), acute tendonitis and bursitis (NCT01161615, NCT01506154), and shoulder pain (NCT01506154) have been completed. In March 2016, a phase III trial in the treatment of acute delayed onset muscle soreness was announced (NCT02695381).⁶³⁹ At the end of November 2016, MEDRx declared termination of the development of Etoreat due to unsatisfactory results of the trials.

4. CHEMICAL APPLICATIONS OF IONIC LIQUIDS IN PHARMACY

4.1. Ionic Liquids in Drug Synthesis

Ionic liquids are used in numerous chemical processes including fascinating application in the synthesis of pharmaceutical substances and drugs. ILs are employed in synthesis of heterocyclic molecules, such as imidazoles, furans, oxazoles, thiazoles, quinolines, and others, which are exploited in biology and medicine.⁶⁶¹ Because of their unique nature, ILs can simultaneously play dual roles of media and catalysts, thus allowing reduced volumes of volatile solvents and reduced quantities (or even elimination) of metal catalysts that may leach into the environment. ILs can also influence the selectivity of enzymes used in biotechnological processes. Previous excellent reviews cover these topical issues.^{14,15,219,662,663} In this section, we discuss important issues of the IL employment in drug synthesis and point out on interactions between ILs and proteins, which may be responsible for the unique properties demonstrated by pharmaceutically significant enzymes in the presence of ILs.

Because of their excellent solvent characteristics, ILs are often employed as reaction media and catalysts. Of note, upon dissolution in ILs, many substances display new behavior, which differs from that in molecular liquids. This phenomenon may be explained by intense intermolecular interactions between the solute and IL resulting in the formation of nanostructures (see section 1.2). In the case of uncharged solutes, the solute molecules are suggested to distribute in the polar or nonpolar nanodomains existing in ILs; small, charged solutes are supposed to undergo complete dissociation, and the ions are supposed to allocate in the polar IL nanodomains. According to several reports, transition metal ions form coordination complexes with IL anions, leading to local changes in the IL structure. Unfortunately, the case of bulkier charged solutes is significantly less studied.⁶⁷

In 1997, hydrogenation of 2-arylacrylic acids in the [C₄MIM]-[BF₄]⁻ media was published. The authors employed the procedure for producing the nonsteroidal anti-inflammatory drug (NSAID) (S)-naproxen.⁶⁶⁴ In 2000, the first high-yield synthesis of a drug in an ionic liquid media was reported: the NSAID pravastatin was prepared by using [C₄MIM][PF₆]⁻ as media.⁶⁶⁵ Thus, the age of ionic liquids in the drug synthesis has begun.

In the last 15 years, many research groups have reported the employment of ILs in various chemical processes related to the production of pharmaceuticals, their precursors or intermediates, and other agents with proven or potential biological activity. In most cases, ILs are used as reaction media, which often also acts as catalyst, and the most popular ILs are salts of imidazolium (see Table 3). So far, the synthesis of several NSAIDs,^{664,666–668} antiviral,^{669–671} antimicrobial,^{668,672–686} antimalarial,^{687,688} and antitumor^{669,689–701} agents, as well as cholinesterase inhibitors^{702–705} and radiolabeled molecular imaging and therapy agents,⁷⁰⁶ have been described. Table 3 shows important

examples of advanced synthesis of drugs molecules using IL media. There are also many reports on the synthesis of molecules, which can be used as precursors for potential drugs with various activities and contain sulfonamide,⁶⁷² lactam,⁶⁷⁵ imidazole,^{707,708} pyrazolone,⁷⁰⁹ thiazole,^{710,711} or thiazolidine^{712,713} cores.

One of the most interesting and promising discoveries concerns the employment of ILs for controlling the stability and selectivity of enzymes. ILs have recommended themselves as “chaperons”, which interact both with water and with the protein surface in the solution. Usage of IL additives provides stabilization and enhanced activity of enzymes,^{47,733} allows controlled, reversible folding/unfolding of proteins, and reduces protein aggregation.^{96,734–737} Imidazolium-based cations with short alkyl side chains mostly stabilize the protein structure and preserve its activity.⁷³⁴ However, the behavior of large, multidomain proteins in the IL media can be rather complex, and making general assumptions and conclusions on the basis of the existing data is a difficult task.^{734,738}

In many cases, the IL impact on a protein structure seems to be related to interactions between ILs and the aqueous environment of a protein. Because hydration energies of charged molecules are typically rather large, the relocation of water bound to them should produce considerable energetic effects. Water possesses a high dielectric constant, which renders it an efficient solvent for ions and biological polyelectrolytes, such as proteins and DNA. In the case of proteins, surface water molecules interact preferentially with charged amino acids, as well as with other polar groups. Protein folding is known to be mediated by aqueous solvating: water hydrates the protein backbone and “guides” it toward a final active structure. At the protein surface, water molecules gather in clusters, which are governed by the hydrogen-bonding capacity and structure of the protein; some of these molecules occupy specific sites and are highly important for correct protein function. Such water molecules can be seen as a rightful part of the secondary structure of a protein, whereas ordered water molecules between donor and acceptor sites can promote proton and electron transfer. Being a nucleophile and proton donor, water can serve as a reagent in the active site of a protein. Moreover, hydrogen-bond networks are suggested to provide dielectric shielding between protein molecules, thus promoting directed electrostatic interactions between their hydrophilic groups.^{739–742}

Most ILs, except highly hydrophobic ones, contain some water. Moreover, water can be easily adsorbed by ILs from humid air. According to molecular dynamics simulations, water molecules, which are present in IL, accumulate near charged electrodes, and their amount increases upon increasing the surface charge density.⁷⁴³ This observation agrees with experimental studies, according to which water molecules form hydrogen-bond networks at the electrode interface⁷⁴⁴ and interact with IL, therefore diminishing the interactions between the electrode surface and cations and anions.^{745,746} Of note, the mechanisms of surface charging are suggested to influence the water impact on IL structuring significantly; this impact is supposedly related to balances between interactions between ions, surface, and water, which can vary depending on their combination.⁷⁴⁷ Interestingly, water–IL systems demonstrate electrochemical anomalies, which depend on the amount of water. Thus, the ethylammonium nitrate–water system demonstrated electrochemical instabilities in the intermediate state between IL and bulk water (70–90% water in the system).⁷⁴⁸ All

of these data support the idea of water being a mediator of at least some effects imposed by ILs on proteins.

The ability of ILs to stabilize protein molecules may be generally systemized similarly to small ions of the traditional salts. IL ions can be divided into kosmotropic (structuring) ions, which stabilize protein molecules, and chaotropic (destructing) ions, which destabilize protein molecules. Kosmotropes are intensively hydrated ions with pronounced salting-out activity; chaotropes are less hydrated ions with weak salting-out activity toward protein molecules. Thus, IL cations can be ascribed to chaotropic ions. However, due to the complex structure of IL ions and their capability to participate in various types of interactions, there are some difficulties in the unambiguous classification of IL ions to kosmotropic or chaotropic groups.^{354,734}

According to molecular dynamics simulations, α -chymotrypsin⁷⁴⁹ and lipases from *Candida* spp.⁷⁵⁰ demonstrated higher stability in aqueous solutions of the $[\text{C}_4\text{MIM}]^+$ ILs, as compared to water. The ILs stripped nonlocalized water molecules from the protein surface, and the effect correlated with the solubility and hydrophobicity of the anions; thus, smaller anions with higher water solubility, such as chloride, were able to enter the protein core and caused its destabilization.^{749,750}

A combined simulation and experimental study on the interaction of cytochrome *c* with ethylammonium nitrate showed that the IL formed a tight shell around the solvated protein supporting its renaturation.⁷⁵¹ On the contrary, ammonium-based ILs turned out to be powerful destabilizers of heme proteins. The length of the cation alkyl chains correlated with the effect: shorter chains imposed stronger destabilization, possibly due to stronger interactions with the hydrophobic regions of the proteins, which, in turn, might be related to the IL viscosity.⁷⁵² Similarly, aqueous $[\text{C}_2\text{MIM}][\text{OAc}]$ led to denaturation of a β -hairpin peptide.⁷⁵³ Aqueous solutions of $[\text{C}_2\text{MIM}][\text{OAc}]$ and $[\text{C}_3\text{MIM}][\text{C}_2\text{SO}_4]$ interfered with the activity of xylanase from *Trichoderma longibrachiatum*, however, not due to denaturation of the enzyme but due to hindrance of its dynamic motion and kinetic trapping of the IL cations in its binding pocket.⁷⁵⁴

The existence of a hydration layer was crucial for the enzyme stabilization; thus, the structure of cutinase from the fungus *Fusarium solani* was dependent on the water amount in IL. $[\text{C}_4\text{MIM}][\text{PF}_6]$ removed water from the protein surface, but the remaining water molecules were enough for retention of the active structure; contrariwise, $[\text{C}_4\text{MIM}][\text{NO}_3]$ promoted more extensive water removal, which led to the enzyme inhibition. Strong interactions between $[\text{NO}_3]^-$ and hydrogen donor groups of cutinase were supposed to be the main cause of the structural destabilization.⁷⁵⁵

As was shown by classical MD simulation, cations of $[\text{C}_3\text{MIM}][\text{Br}]$ may replace water molecules in the solvation layer of lysozyme. This replacement caused the reduction of the size of the protein molecule (the hydrodynamic radius decreased from 18 Å in water to 11 Å in the water/IL mixture) and accelerated its conformational dynamics. The overall radius of the area of preferential solvation of the protein molecule by the IL cation was 42 Å.⁷⁵⁶

An experimental study on the stability of protein dispersions in aqueous solutions of $[\text{C}_n\text{MIM}][\text{Cl}]$ ($n = 6, 8$) suggested that the proteins preferentially bound to the IL via electrostatic interactions between charged areas of the protein molecule and the imidazolium part and chloride anion of the IL; the formation of hydrogen bonds and van der Waals interactions was also possible. Therefore, ILs penetrated the first hydration level of the

protein and replaced it with an IL bilayer, which enhanced the stability of a nonaggregated protein fraction (monomers and dimers). The protein monomers subsequently adhered to each other due to overlapping of hydrophobic tails of the bound IL, which led to the formation of an aggregated fraction (bulky protein clusters). The presence of IL inhibited the pH-dependent protonation/deprotonation occurring in the protein dispersions in water, but had little impact on the protein integrity.⁷⁵⁷ The authors suggested potential application of ILs as aggregation inhibitors for protein solutions (Figure 22).⁷⁵⁸

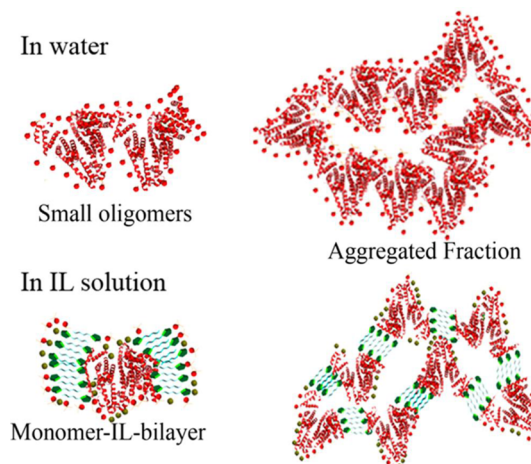


Figure 22. BSA aggregation in aqueous solutions in the presence/absence of $[\text{C}_3\text{MIM}][\text{Cl}]$. Reproduced with permission from ref 757. Copyright 2012 American Chemical Society.

Interestingly, positive supercharging of the surface of endoglucanase E1 from *Acidothermus cellulolyticus* assisted the maintenance of its wild-type activity in $[\text{C}_2\text{MIM}][\text{OAc}]$ and other ILs, which otherwise inhibited the enzyme.⁷⁵⁹ According to a theoretical study, $[\text{C}_2\text{MIM}][\text{OAc}]$ demonstrated preferential solvation of positively charged solutes, possibly because of higher packing density in the first solvation shell formed by acetate ions; this result highlights the dominance of electrostatic interactions between the first solvent shell and the solute.⁷⁶⁰

According to the existing reports, the intact hydration layer is not always necessary for the protein integrity. Thus, anhydrous ILs were shown to solubilize dry proteins by forming surfactant nanostructures,⁷⁶¹ and ionization properties of IL were suggested to be responsible for the enzyme activation (Figure 23).⁷⁶²

According to a theoretical study on lipase B from *Candida antarctica* (CAL-B), the enzyme formed Coulomb interactions with the IL anions and van der Waals interactions with the cations; the ions, which formed strong hydrogen bonds with the enzyme, promoted conformational changes in the protein, leading to its denaturation. The butyl group of the $[\text{C}_4\text{MIM}]^+$ cation was able to penetrate into the active site of the lipase, possibly inhibiting its activity.⁷⁶³ In the consecutive study, the authors investigated the process of CAL-B destabilization by ILs and defined two modes of the IL action: destabilization of the enzyme surface occurred mostly due to strong Coulomb interactions with the IL anions and polar cation groups, whereas destabilization of the enzyme core was due to conformational changes induced by long alkyl chains of ILs; in the latter case, these changes opened the access of IL ions into CAL-B.⁷⁶⁴ Similar results were obtained in an experimental and theoretical study of interactions between bovine serum albumin (BSA) and

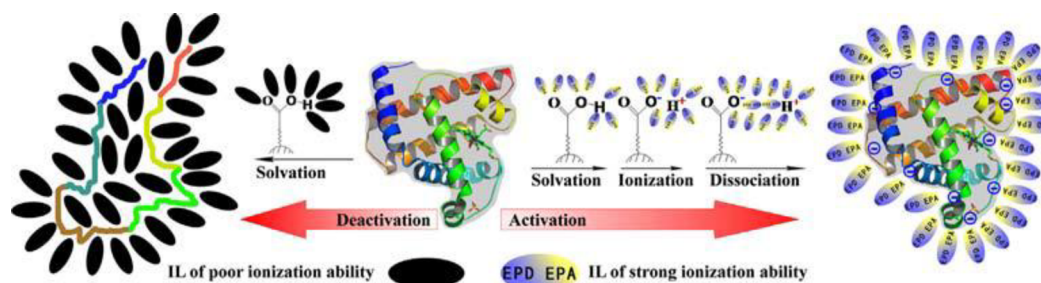


Figure 23. Enzyme activation/deactivation in ionic liquids. Reproduced with permission from ref 762. Copyright 2016 Elsevier.

imidazolium ILs: ILs led to unfolding of the protein. The imidazolium moiety entered the BSA subdomains and formed contacts with hydrophobic amino acids. The process depended on the anionic moiety of ILs: thus, the presence of the $[\text{NO}_3]^-$ anion resulted in significantly stronger interactions between the IL and BSA, in comparison with the $[\text{Cl}]^-$ anion.⁷⁶⁵ A study on the interaction between BSA and $[\text{C}_n\text{MIM}][\text{Br}]$ ($n = 4, 6, 8, 10$) demonstrated that in the case of $[\text{C}_{10}\text{MIM}][\text{Br}]$, hydrophobic interactions were predominant, whereas in the case of $[\text{C}_4\text{MIM}][\text{Br}]$, $[\text{C}_6\text{MIM}][\text{Br}]$, and $[\text{C}_8\text{MIM}][\text{Br}]$, hydrogen bonds and van der Waals interactions played the major role. Accordingly, $[\text{C}_{10}\text{MIM}][\text{Br}]$ caused significant changes in the secondary structure of BSA.⁷⁶⁶

Molecular dynamics modeling of the *Candida rugosa* lipase 1 in $[\text{C}_4\text{MIM}][\text{PF}_6]$ and $[\text{C}_4\text{MIM}][\text{NO}_3]$ demonstrated that even at 310 and 375 K the ILs significantly decelerated the protein dynamics and detained the system near its starting structure. The interactions between the enzyme surface and the solvent were dominated by the IL anion, possibly due to a broader spatial distribution of positively charged residues of the protein and limited mobility of the bulky cation.⁷⁶⁷ According to a subsequent MD investigation of the native and modified *C. rugosa* lipase in the aqueous ionic liquids $[\text{C}_4\text{MIM}][\text{Cl}]$ and $[\text{C}_2\text{MIM}][\text{C}_2\text{SO}_4]$, there was no indication of structural destabilization of either modified or unmodified enzyme on the 50 ns time scale. In the modified lipase, the surface lysine residues were randomly mutated to glutamate, which effectively decreased the net surface charge. Changes in the surface characteristics of the protein were shown to impact the close-range ion solvation shells; thus, a decrease in the local concentration of anions and an increase in the bound cations or water molecules were observed due to the increase of the number of negatively charged residues. The solvent reorganization was suggested to be crucial for the protection of the enzyme from smaller anions, which could permeate its surface and disrupt its structure.⁷⁶⁸ Therefore, targeted modification of enzymes can be an effective way to regulate their tolerance toward IL action and their biopharmaceutical activity.

According to MD simulation, $[\text{C}_1\text{C}_4\text{Pyr}][\text{NTf}_2]$ could dramatically decrease the mobility of a 20-residue tryptophan (Trp) cage mini-protein. The unfolded Trp cage folded in water at 298 K in less than 500 ns of simulation; on the contrary, it demonstrated very limited mobility in IL at the same temperature, possibly due to higher IL viscosity. Nevertheless, the folded Trp cage in IL closely resembled that in water. Similar theoretical results were obtained for amino-acid-based ILs.⁷⁶⁹ Complete substitution of water by organic cations and anions further hampered the conformational flexibility of the Trp cage, suggesting that such ILs could protect proteins from thermal denaturation. Comparison of the Trp cage solution in the amino-acid IL and in water showed that the amino acid anions

dominated in the first coordination sphere of positively charged sites of the Trp cage, whereas water molecules dominated in the first coordination sphere of negatively charged sites of the Trp cage.⁷⁷⁰ ILs also selectively promoted folding of various secondary structures of proteins,⁷⁷¹ implying possible application of ILs as stabilizers.

All of these data also suggest the possibility of modulating the structure and activity of an enzyme by tuning the structure of an IL supplement. Thus, cocosalkylpentaethoxymethylammonium methyl sulfate was found to induce changes in the secondary and tertiary structures of β -galactosidase from *Thermus thermophilus*; according to experimental and theoretical studies, the presence of IL led to increased flexibility of the enzyme and subsequent modifications in its activity and regioselectivity.⁷⁷² Similarly, the presence of $[\text{C}_4\text{MIM}][\text{PF}_6]$ changed the substrate specificity of β -galactosidase from *Aspergillus oryzae*.⁷⁷³ $[\text{C}_8\text{MIM}][\text{Cl}]$ and $[\text{C}_{10}\text{MIM}][\text{Cl}]$ stimulated the activity of the firefly luciferase: the imidazolium ring of the ILs blocked the entrance into the AMP pocket of the enzyme; the ILs acted as competitive inhibitors of AMP and luciferin, which resulted in low-concentration hormetic stimulation. For $[\text{C}_6\text{MIM}][\text{Cl}]$ and $[\text{C}_{12}\text{MIM}][\text{Cl}]$, the effect was less pronounced, whereas $[\text{C}_2\text{MIM}][\text{Cl}]$ and $[\text{C}_4\text{MIM}][\text{Cl}]$ demonstrated no hormesis.⁷⁷⁴ To overcome inhibitive effects of ILs on biocatalytically demanded enzymes, targeted protein engineering for diminishing unfavorable interactions between ILs and proteins was proposed.^{775,776}

$[\text{C}_4\text{MIM}][\text{PF}_6]$ provided a suitable environment for kinetic resolution of (*R,S*)-ibuprofen by native lipases from *Candida rugosa* and *Aspergillus*,^{666,667} whereas $[\text{C}_8\text{C}_2\text{Py}][\text{BF}_4]$ allowed efficient production of (*S*)-ibuprofen from racemic ibuprofen ethyl ester by thermostable esterase (EST10) from *Thermotoga maritima*.⁷¹⁵ The ability of ILs to affect the enzyme regioselectivity was exploited in lipase-catalyzed synthesis of esters of nucleoside drugs (derivatives of ribavirin, cytarabine, and inosine), where 10% supplement of $[\text{C}_4\text{MIM}][\text{BF}_4]$ accelerated the reaction and led to significantly higher yields.⁶⁶⁹ $[\text{C}_4\text{MIM}][\text{NTf}_2]$ turned out to be a promising solvent for *trans*-esterification of methyl caffeate by lipase B from *Candida antarctica*; the obtained esters demonstrated antiproliferative activity;⁶⁹⁴ this enzyme also effectively synthesized caffeic acid phenylethyl ester in $[\text{C}_2\text{MIM}][\text{NTf}_2]$.⁷⁷⁷ $[\text{C}_4\text{MIM}][\text{PF}_6]$ and $[\text{C}_4\text{MIM}][\text{BF}_4]$ showed good results when used as cosolvents in *trans*-esterification of (*R,S*)-1-chloro-3-(3,4-difluorophenoxy)-2-propanol, which was an intermediate in the synthesis of the neuroprotector (*S*)-lubeuzole, by lipases from *Pseudomonas aeruginosa*.⁷²⁷ $[\text{C}_2\text{MIM}][\text{Br}]$ facilitated enantioselective production of (*R*)-modafinil by chloroperoxidase from *Caldariomyces fumago*.⁷¹⁶

Recently, a water-soluble artificial enzyme demonstrated a significantly enhanced ability to promote a dephosphorylation reaction, when bearing an ionic imidazolium tag. The

Table 4. Application of ILs in the Preparation of Drug Delivery Systems

drug delivery system	IL	role of IL	ref
microporous poly(lactic acid) scaffolds	[C ₄ MIM][OTf], [C ₄ MIM][Cl], [C ₄ MIM][BF ₄], [C ₄ MIM][SbF ₆]	microporosity enhancers	784
polylactic acid and polycaprolactone membranes and microspheres	[(C ₈) ₃ C ₁ N][Cl], [C ₄ MIM][NTf ₂], [C ₄ MIM][PF ₆], [C ₄ MIM][BF ₄], [C ₄ MIM][OTf], [C ₄ MIM][Cl], [C ₄ MIM][SbF ₆]	additive for microsphere preparation	785–788
cellulose-biopolymer composite hydrogel	[C ₂ MIM][OAc]	solvent	793
cellulose-graft-poly(L-lactide)	[C ₄ MIM][Cl], [AlkMIM][Cl]	solvent	794,814
cellulose/SWCNT complex	[C ₄ MIM][Br]	solvent	795
cellulose-nanohydroxyapatite composite scaffolds	[C ₄ MIM][Cl]	solvent	796
cellulose, keratin, and chitosan composite materials	[C ₄ MIM][Cl]	solvent	800
chitosan-based nanocarriers	[C ₄ MIM][Cl]	solvent	801
linoleic acid-grafted chitosan micelles	[C ₄ MIM][OAc]	solvent	802
biopolymeric chitosan Schiff bases with salicylidene IL brushes	IL-based salicylaldehyde	functionalization agent	803
benzylpyrazolyl coumarin scaffolds	[C ₂ MIM][Br]	catalyst and reaction media	782
fullerenes	[C ₄ MIM][BF ₄]	fullerene dispersion enhancer	781
curcumin-loaded platinum-phytase nanospheres	[C ₄ MIM][BF ₄]	media for self-assembly of enzyme nanospheres	783
silica particles	[C ₄ MIM][BF ₄]	solvent	805
Ag-NPs	[HOC ₂ C ₁₄ IM][Br], [(HO) ₂ C ₃ C ₁₄ IM][Cl], [HOC ₂ C ₁ C ₁₄ IM][Cl], [HOC ₂ C ₁ C ₁₆ IM][Cl], [C ₁₆ Py][Sal]	dispersing agent	628,812
starch nanoparticles	[C ₁₆ MIM][Br] + [C ₈ MIM][OAc], [C ₁₆ MIM][Br], [C ₄ MIM][PF ₆], [C ₈ MIM][NTf ₂] + [C ₁₆ MIM][Br]	microemulsion cross-linking reaction	806–809
polyphosphate nanogel	[C ₄ MIM][BF ₄]	solvent	789
magnetic mesoporous CoNi@Au nanorods	[C ₄ MIM][PF ₆]	IL-in-water microemulsion as electrochemical media	811
Au-NPs	bis-imidazolium amphiphiles	ligands acting as transfer agents and NP stabilizers	810

phenomenon was explained in the terms of IL effects: the imidazolium cation was supposedly responsible for the substrate activation.¹³⁵

Another area of IL usage in pharmaceutical synthesis is closely related to the previous one and also exploits unique IL abilities to affect enzymatic activity and selectivity. In whole-cell biosynthetic processes, ILs are used as components of ionic liquid/water solvent systems to overcome low water solubility and toxicity of substrates and products.^{778–780} Thus, [C₄MIM][NTf₂] was used for promoting *Escherichia coli*, which expressed reductase from *Saccharomyces cerevisiae* and glucose dehydrogenase from *Bacillus subtilis*, to enantioselectively convert 3-chloro-1-phenyl-1-propanone to (*S*)-3-chloro-1-phenyl-1-propanol, which was a building block for the synthesis of antidepressant drugs.⁷²⁴ A biphasic system consisting of [C₄MIM][PF₆] and buffer was employed for enantioselective reduction of acetyltrimethylsilane to (*R*)-1-trimethylsilylethanol, a precursor of silicon-containing drugs, by *Candida parapsilosis*. Both the substrate and the product were toxic for the immobilized cells, but the IL system allowed separating the dangerous substances from the biocatalyst.⁷³⁰ Another example of successful employment of an IL system containing [C₄MIM][PF₆] in the whole-cell biocatalysis is hydroxylation of 16 α ,17-epoxyprogesterone, an important stage in the synthesis of steroidal drugs, by *Rhizopus nigricans*.⁷²⁹

There are also numerous reports on using ILs for production of diverse drug delivery systems. According to Table 4, which includes examples of current IL applications in preparation of drug delivery systems, in this case, imidazolium ILs also seem to be the substances of choice and are mostly used as media and dispersion agents. Thus, [C₄MIM][BF₄]-water mixtures allowed effective dispersion of hydrophobic C₆₀ fullerenes, which showed high biomedical potential.⁷⁸¹ Benzylpyrazolyl coumarin scaffolds, which presumably possessed inhibitory

activity toward the cyclooxygenase COX-II, were synthesized by using [C₅MIM][Br] as catalyst and reaction media.⁷⁸² The enzyme phytase self-assembled into active nanospheres in [C₄MIM][BF₄], and these nanospheres were subsequently used for production of platinum-decorated, curcumin-loaded phytase nanospheres, which the authors considered “multi-functional protein-based drug delivery vehicles”.⁷⁸³

[C₄MIM][OTf], [C₄MIM][Cl], [C₄MIM][BF₄], and [C₄MIM][SbF₆] were employed for developing tissue-compatible microporous poly(lactic acid) scaffolds with enhanced cell adhesion properties and drug loading capacity.⁷⁸⁴ Highly porous polylactic and polycaprolactone membranes and microspheres were prepared using [(C₈)₃C₁N][Cl] or [C₄MIM] ILs with various anions.^{785,786} The polycaprolactone microspheres were subsequently modified with water-soluble carbon nanotubes and paclitaxel or NGF-containing gelatin to obtain model drug delivery systems.^{787,788} IL miniemulsions were employed for the synthesis of a biocompatible branched polyphosphate nanogel.⁷⁸⁹

There are several patents on cellulose dissolution and processing in ILs,^{790–792} and ILs turned out to be helpful in the preparation of cellulose-based polymers with possible biomedical applications (see examples in Table 4). Thus, dissolution of cellulose in [C₂MIM][OAc] with subsequent addition of lipase from *Candida rugosa* allowed efficient entrapping of the enzyme in the cellulose hydrogel.⁷⁹³ Microcrystalline cellulose-graft-poly(L-lactide) copolymers were synthesized in [C₄MIM][Cl]; micelles of these copolymers demonstrated high efficiency in drug loading.⁷⁹⁴ A biocompatible complex of cellulose with single-walled carbon nanotubes was prepared in [C₄MIM][Br].⁷⁹⁵ Employment of [C₄MIM][Cl] allowed better dispersion of hydroxyapatite in cellulose when preparing cellulose-nanohydroxyapatite composite scaffolds for drug encapsulation.⁷⁹⁶

It should be mentioned that the solubility of natural polymers, such as cellulose, in ILs is a very important topic. Thus, cellulose is poorly soluble in water and common organic solvents; application of ILs allows one to realize the chemical conversion of plant biomass, which has been considered a rather difficult task before.^{34,36,797,798} Similarly, imidazolium-based ILs turned out to be appropriate solvents for chitin.⁷⁹⁹

Cellulose, keratin, and chitosan composite materials suitable for controlled drug release,⁸⁰⁰ as well as temperature- and pH-sensitive chitosan-based nanocarriers,⁸⁰¹ were prepared by using $[C_4MIM][Cl]$ as solvent. Micelles of linoleic acid-modified chitosan were produced in $[C_4MIM][OAc]$.⁸⁰² Recently, biopolymeric chitosan Schiff bases with salicylidene IL brushes complexed with Ag(I) were suggested to be used for the development of potent anticancer and antimicrobial agents.⁸⁰³

The first theoretical study of cellulose dissolution in ILs by classical MD was carried out for $[C_2MIM][OAc]$ as solvent. The energy of the interaction between the polysaccharide chain and IL was higher than that for either water or methanol. The anion formed strong hydrogen bonds with hydroxyl groups of cellulose, whereas some cations established hydrophobic interactions with the polysaccharide, confirming the hypothesis that the cation was important for the dissolution of cellulose in IL.⁸⁰⁴

ILs were also used for the preparation of various biomedical nanoparticles from silica,⁸⁰⁵ starch,^{806–809} silver,^{93,94} gold,⁸¹⁰ and cobalt–nickel–gold⁸¹¹ (see Table 4). In the case of silver, the particle size and dispersivity of nanoparticles depended on the IL structure.⁸¹² Of note, Ag nanoparticles with enhanced antimicrobial activity were produced using $[C_{16}Py][Sal]$ as dispersing agent.⁶²⁸

At the end of this section, we would like to mention a recent report on the application of ILs for skin tissue engineering. $[C_4MIM][OAc]$ was proposed to be used as solvent for muga fibroin from cocoons of the silkworm *Antheraea assama*. The resulting solution was subsequently used for electrospinning of nanofibrous mats, which could become a promising alternative to xenotransplants or allotransplants.⁸¹³

4.2. Ionic Liquids in Biomedical Analytics

Because of their unique and flexible properties, ionic liquids find application in numerous fields, including those related to chemical analysis. ILs are employed in electrophoresis,^{80,81,815} extraction,^{81,85,87,488–490,816,817} gas and liquid chromatography,^{80,81,818,819} UV–visible, IR, Raman, and fluorescence spectroscopy,^{80,81} NMR,^{81,164,165,167–169,820} and mass spectrometry,^{80,81,170,821,822} as well as in various sensor systems (Figure 24).^{80–82,823–825} Several comprehensive reviews have been published on the topic recently.^{80–82,818,825} Therefore, in this section we discuss only some representative examples of successful application of ILs in the detection of natural and pharmaceutical substances.

In one of the first reports on the application of ILs in analytical techniques, an imidazolium IL-coated capillary was used for the determination of sildenafil and its metabolite in the human serum by capillary zone electrophoresis–ion-trap mass spectrometry. The IL coating provided improved resolution and precluded absorption of the substances on the capillary wall.⁸¹⁵ Usage of imidazolium-based ILs as mobile phase additives in the reversed-phase liquid chromatography enhanced separation of basic drugs by alleviating the issue of broad chromatographic peaks, which arose through interactions of the positively charged drugs with free silanol groups of the reverse phase.^{826,827} In addition, employment of chiral ILs as electrolyte additives during

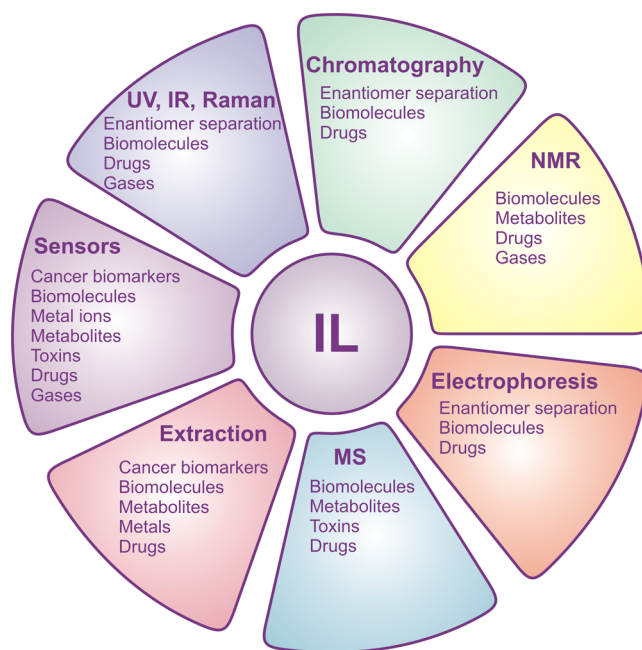


Figure 24. Graphical summary of application of ionic liquids in biomedical analytics.

electrophoresis allowed separating enantiomers of various active pharmaceutical ingredients.^{828–830} Similarly, an IL-functionalized β -dextrin was successfully used as chiral selector in capillary electrophoresis.⁸³¹ A fluorescence technique based on interactions of enantiomers with optically active ILs was proposed for establishing enantiomeric composition of drugs.⁸³²

Excellent extraction properties of ILs have stimulated the development of numerous IL-based extraction systems for chemical and pharmaceutical analysis. There are three possible mechanisms of extraction: the solute can move between the phases in the form of supramolecular associates; partial mixing of the solvents may lead to transition of both solute and solvent molecules from one phase into the other and therefore to the nonuniformity of the phase boundary; and pure molecular extraction can occur, with solute molecules passing the boundary individually (Figure 25). For extraction of peptides from IL to the organic phase, the third mechanism (molecular diffusion) was found to be preferable.⁴⁸⁸

The IL-based microextraction combined with capillary electrophoresis was employed for the detection of analytes in

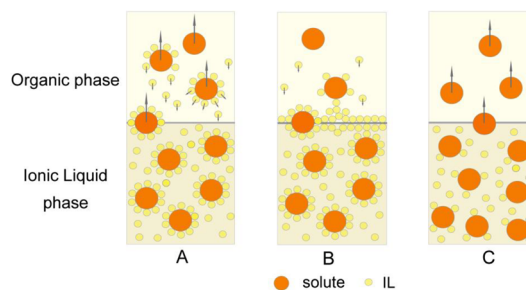


Figure 25. Possible mechanisms of solute transition from IL to organic phase: (A) Solute molecules move as supramolecular associates; (B) partial mixing of the phases occurs; and (C) solute molecules pass the boundary individually by molecular diffusion. Reproduced with permission from ref 488. Copyright 2015 American Chemical Society.

biological samples and provided up to 1000-fold sensitivity enhancement.⁸³³ Microextraction coupled with gas chromatography and mass spectrometry was proposed to be used for the determination of cancer biomarkers in urea samples. The suggested method was fully automated and allowed high-throughput analysis.⁸³⁴ The IL-based microextraction coupled with liquid chromatography and UV detection was developed for analysis of the levels of the drugs nifurtimox and benzimidazole in the plasma; the method demonstrated very low detection limits and required low amounts of sample.⁸³⁵

Among other analytical methods, NMR and mass spectrometry are highly demanded.^{164,165,168,170} Thus, UV-light absorbing IL-based matrices were efficiently used in matrix-assisted desorption/ionization mass spectrometry (MALDI-MS) for biosensing bacterial toxins without preliminary separation and purification.⁸³⁶

As in many other scientific and industrial areas, the tunability of ILs has won them an important place in the field of electrochemical sensors. ILs are characterized by wide electrochemical windows and good electrochemical stability, which make them promising electrolytes.^{82,825} However, ionic conductivity of ILs strongly depends on their chemical structure and viscosity; thus, imidazolium ILs demonstrate higher conductivity than pyrrolidinium or tetraalkylammonium ones. Therefore, IL-based electrolytes do not necessarily perform better than conventional ones, and the favorable combination of the electrode structure and material is the key to success.⁸²⁵ Nevertheless, ILs are broadly employed in various electrochemical biosensors, which are based on carbon nanotubes (CNT), metal nanomaterial, graphene, carbon paste, and other materials.^{82,823,825}

The IL-containing electrodes are used for the detection of low-level target molecules in biological fluids. A multiwall carbon nanotube (MWCNT)-ionic liquid electrode doped with gold nanoparticles and antibody was applied as immunosensor for human serum albumin.⁸³⁷ An MWCNT-IL-modified electrode was used for developing an immunosensor for the tumor marker prostate specific antigen (PSA) in the serum; the immunosensor was produced by a simple one-step method and showed high sensitivity and reasonable accuracy.⁸³⁸ An electrochemical immunosensor based on IL-doped graphene/gold nanoparticles with alkaline phosphatase as enhancer was used for the detection of the cancer biomarker APE1.⁸³⁹ A carbon IL paste electrode with a graphene/MWCNT composite film successfully detected carbamazepine in the presence of paracetamol in tablets or urine samples.⁸⁴⁰ A nanocomposite from IL-functionalized graphene and gold nanoparticles was used in an electrochemical immunosensor with the detection range from 1 fg mL⁻¹ to 100 ng mL⁻¹; the sensor was efficiently applied for analysis of human serum samples;⁸⁴¹ similarly, an immunosensor based on IL-functionalized reduced graphene oxide and gold nanoparticles was proposed to be employed for simultaneous detection of multiple analytes.⁸⁴² A polymerized IL film-oxide electrode containing ZnCdHgSe quantum dots was decorated with antibody against the serum tumor marker neuron-specific enolase and demonstrated the wide detection range from 1 pg mL⁻¹ to 100 ng mL⁻¹.⁸⁴³

In the end of the section, important reports on employing ILs in DNA studies should be also mentioned. It is well-established knowledge that highly charged counterions have an impact on DNA compaction.⁸⁴⁴ Therefore, it is unsurprising that ILs can influence the DNA structure. Multicharged ions may possibly form by hydrogen bonding in ILs, which overcomes the

electrostatic repulsion between similarly charged ions. Iron-containing magnetic hydrophobic ILs were found to perform excellently as solvents in efficient extraction of single- and double-stranded DNA molecules, as well as bacterial plasmid DNA. The obtained results suggested the possibility of selective extraction due to IL structural fine-tuning.⁸⁴⁵ Of note, a [C₄MIM][PF₆]-containing extraction system was proposed to be used for direct DNA quantification,⁸⁴⁶ whereas bis-(dialkylamino)cyclopropenium chloride-based poly(ILs) formed complexes with plasmid DNA and were proposed to be used for efficient transfection of cells.⁸⁴⁷

ILs of various classes maintained the native double-strand DNA B structure; the cations mostly formed electrostatic interactions with the DNA phosphate groups, whereas the anions established hydrogen bonds with the nucleobases.^{848–851} Hydrated choline dihydrogen phosphate stabilized the formation of a DNA triplex due to the binding of the choline cations to the third DNA strand.⁸⁵² DNA stored in IL showed enhanced resistance against nuclease-induced degradation,^{853,854} and the IL guanidinium tris(pentafluoroethyl)trifluorophosphate was shown to induce DNA compaction.⁸⁵⁵ The authors suggest that multicharged micellar aggregates of the IL are involved in this process: the positively charged part of the IL neutralizes the DNA negative charge (phosphate groups), whereas the anions participate in the formations of micelles; consequently, the DNA strands are attracted to the micellar surface (Figure 26).⁸⁵⁵ [C_nMIM][Cl] (*n* = 8, 12, 16) intercalated into double-stranded DNA, and the binding increased with the increasing alkyl side chain.⁸⁵⁶

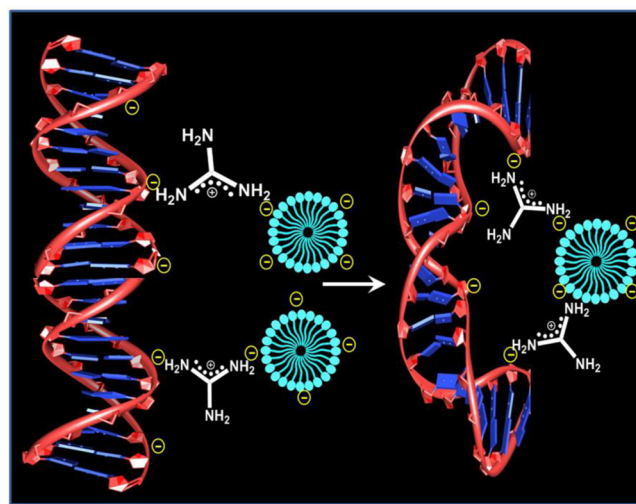


Figure 26. Possible mechanism of ionic liquid-driven DNA compaction. The positively charged part of IL neutralizes the DNA phosphate groups, whereas the anions form micelles to which the DNA strands are subsequently attracted. Reproduced with permission from ref 855. Copyright 2015 Nature Publishing Group.

Molecular dynamics simulations, docking studies, fluorescence, and electrochemical techniques suggest that, similarly to inorganic ions, IL cations interact both with the minor and with the major grooves of DNA. This interaction is mostly realized via the Coulomb attraction of cations to the diffuse negative charge of DNA, but hydrogen bonding, hydration, and dispersion forces also may be important (Figure 27).³⁵⁴

For [C₁MIM][Cl], [C₁MIM][PF₆], and [C₁MIM][NTf₂], it was established that larger anions possessed higher ability to

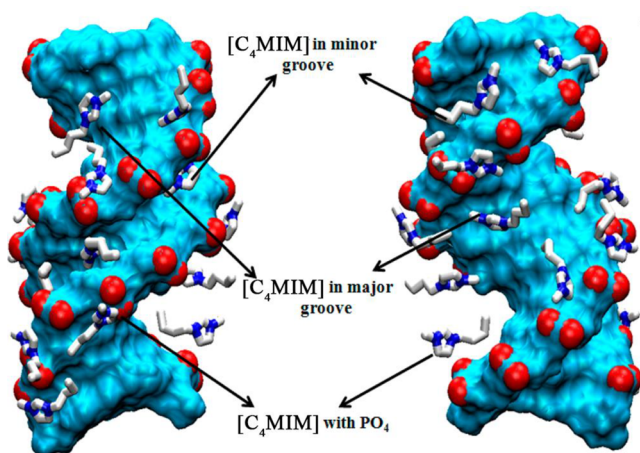


Figure 27. Interaction of $[C_4MIM]$ cation with DNA backbone and grooves. Reproduced with permission from ref 849. Copyright 2012 American Chemical Society.

associate with π -systems of organic molecules ($[Cl]^- < [PF_6]^- < [NTf_2]^-$).¹⁹³ Evidence of $[Cl]^- \cdots \pi$ interactions was found within $[C_4MIM][Cl]$ by DFT and ab initio methods; it should be noted that to produce correct theoretical results, the method must be capable of accurate description of the dispersion interaction.¹⁸⁷ Thus, bulky organic ions of ILs may interact with DNA by dispersion forces, which are especially pronounced in large aromatic molecular systems⁸⁵⁷ and are particularly important for DNA–substrate complexes⁸⁵⁸ and for stabilization of the double-helical DNA structure.⁸⁵⁹

5. CONCLUSIONS

During the last decades, the field of drug delivery has undergone dramatic changes. Medicine has turned from oral and transdermal formulations to sophisticated medium-sensitive carriers and nanoparticles for targeted delivery (Figure 28).⁸⁶⁰ Accordingly, pharmaceutical formulations have changed from powders to diffusions in polymers to micelles and nanoparticles.^{860,861}

Ionic interactions have been undoubtedly shown to take an important part in the formation and function of various drug carriers. As a leading example, block ionomer complexes, or polyion complex micelles, were designed as a class of carriers for charged molecules.^{541–543} These complexes are formed via interactions between ionic block copolymers and macromolecules of opposite charge (DNA, proteins, etc.). Block

ionomer complexes are characterized by high efficiency of loading, ease of preparation, and preservation of biomolecule activity, and are suggested to be used for overcoming the blood–brain barrier and delivering active substances into the brain.^{542,543}

Another promising property of ionic carriers is the possibility of targeted release of their cargo. Biomolecules can leave the carrier not only via diffusion or carrier degradation, but also due to a pH shift or displacement by environmental counterions. Thus, anionic 5'-triphosphates of nucleoside analogues left the cationic nanogel upon interaction of the latter with cellular membranes, whereas cytochrome *c* exited the anionic gel upon acidification or addition of Ca^{2+} ions (Figure 29).⁵⁴¹

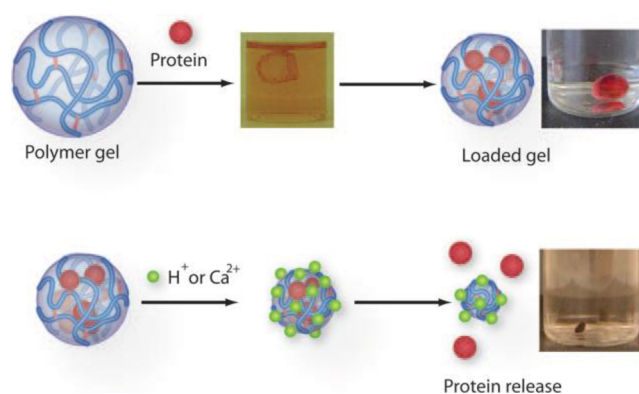


Figure 29. Loading and release of cytochrome *c* from anionic gel. Cytochrome *c* spontaneously forms a complex with anionic chains of the gel; upon acidification or addition of calcium ions, the protein is released due to gel protonation or competitive binding. Reproduced with permission from ref 541. Copyright 2009 Wiley-VCH Verlag GmbH & Co. KGaA.

In view of state-of-the-art tendencies in the development of new drugs, the following advantages and opportunities of ILs in the field of pharmaceuticals and medicine can be summarized on the basis of the literature discussed in this Review:

(I) It is a straightforward and cost-efficient way to generate highly tunable, diverse libraries of biologically active compounds with a virtually unlimited number of combinations of anions and cations.

(II) ILs exhibit control of ion formation in solution and adjustment of solvation properties in water and biological fluids to provide a flexible approach for alleviation of solubility,

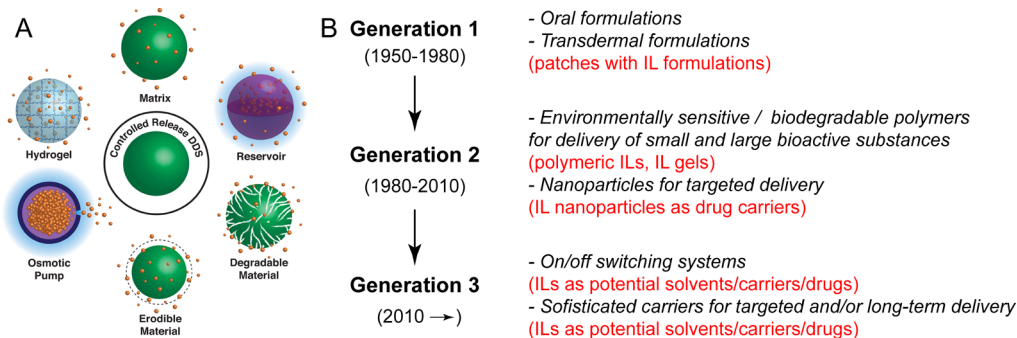


Figure 28. Evolution of controlled drug delivery systems: from oral and transdermal formulations to “clever” environmentally sensitive carriers. Part (A) reproduced with permission from ref 861 (Copyright 2016 American Chemical Society), and part (B) (in adapted form) reproduced with permission from ref 860 (Copyright 2014 Elsevier).

bioavailability, and polymorphism limitations of conventional drugs.

(III) ILs allow easy incorporation of readily developed ionic cores into existing drug molecules, as well as design of novel drugs and biologically active molecules.

(IV) It is a powerful API-IL concept to fine-tune hydrophobic/hydrophilic properties, nature of ionic core, covalent/ionic binding, linker size, and characteristics via simple organic synthesis procedures (Figure 21).

(V) ILs show intrinsically tunable structural organization (section 1.1) reflected at the nanoscale and microscale levels (section 1.2) with access to various types of biological activity (sections 2 and 3).

(VI) Application of ILs allows one to advance (bio)synthetic processes and to optimize the cost of production/analytcs of pharmaceutical compounds (section 4).

The active development of the field has raised a number of important questions. Thus, the following challenges can be formulated upon the analysis of the current literature:

(I) Mechanisms of action of many biologically active compounds with ionic nature are poorly understood and require more detailed studies.

(II) Systematic comparative studies on different types of IL-based drug development systems are currently missing, because most of the reported results are focused on particular types of IL systems.

(III) Most of the studied systems utilize only a combination of ions (section 3) and Type I of the API-IL concept (Figure 21), whereas powerful but more complicated systems remain a future challenge.

(IV) The studies on the relationship between the molecular structure and nano-/microscale organization and translation of molecular properties into self-organized structures are only at the early stage of development.

(V) The fundamental nature of the systems involving formation, organization, and chemical reactivity of ions in the liquid phase urgently appeals for the development of new experimental and theoretical methods to gain the necessary understanding.

Research activity in the area of ionic liquids has reached an astonishing level with an enormous number of studies (Figure 30). The number of publications and the growth trend show a behavior similar to that of the mature life-science fields, such as genomics and proteomics, which have a principal influence on the progress in pharmaceuticals and medicine.^{862–864}

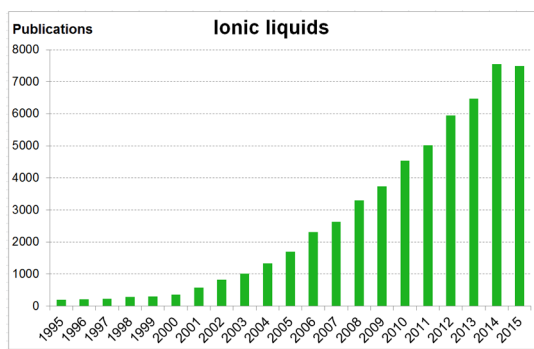


Figure 30. Number of publications on the subject of ionic liquids and molten salts (according to the Web of Science database, <http://www.webofknowledge.com/>).

Ionomics, the studies of ions in liquids, is the key point of the ionic liquid rise, which has already made a valuable contribution to synthesis, catalysis, and material science.⁸⁶⁵ Ionomics should cover not only the pool of ions in biological systems, but also a broad range of studies on their properties and applications, which vary from classical chemical reactions to nanotechnology and drug delivery systems. Now it is clearly established that ionic interactions and ions themselves play important roles in building and maintaining structures of biological molecules and artificial materials, in enzymatic reactions and various chemical processes. The time is ripe now for fundamental insight into more complex chemical phenomena involving ILs and for their application in pharmaceuticals and medicine.

As highlighted in the recent literature, ILs are anticipated to find several applications in drug delivery systems, from well-known conventional formulations to state-of-the-art carriers. The main subject of this Review deals with the rapidly developing area of in-depth studies on biological activity of ILs, and our aim is to attract the attention of a broad audience of readers to the challenges and opportunities of this fascinating interdisciplinary research. The emerging applications of ionic liquids in pharmaceutical science and drug development represent a collaborative effort between chemical, biological, and medical scientists, which will certainly uncover even more striking possibilities in the near future.

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Biographies

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Evgeniy Gordeev completed his M.Sc. (2004) and Ph.D. in theoretical modeling and computations chemistry (2007) at the Lomonosov Moscow State Academy of Fine Chemical Technologies. Since 2011, he has been a researcher at the Zelinsky Institute of Organic Chemistry Russian Academy of Sciences. His research interests include theoretical studies, molecular modeling, reaction mechanisms, and scientific visualization.

Valentine Ananikov received his M.Sc. degree in 1996 (biochemistry), Ph.D. degree in 1999 (organic chemistry and catalysis), Habilitation in 2003, and in 2005 he was appointed Professor and Laboratory Head of the Zelinsky Institute of Organic Chemistry, Russian Academy of Sciences. In 2008 he was elected as a Member of the Russian Academy of Sciences. In 2012 he became Professor of the Chemistry Department of Moscow State University. He was a recipient of the Russian State Prize for Outstanding Achievements in Science and Technology (2004), an Award of the Science Support Foundation (2005), a Medal of the Russian Academy of Sciences (2000), Liebig Lecturer by German Chemical Society (2010), the Balandin Prize for outstanding achievements in the field of catalysis (2010), Organometallics Distinguished

Author Award Lectureship by American Chemical Society (2016), and the Hitachi High-Technologies Award in Appreciation for Novel Approach and Outstanding Contribution to Setting New Standards for Electron Microscopy Applications in Chemistry (2016). His research interests are focused on mechanistic studies, catalysis, ionic liquids, and molecular complexity.

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ABBREVIATIONS

Cations of ILs

[Im]	imidazolium
[AlkMIM]	1-alkyl-3-methylimidazolium
[AMIM]	1-allyl-3-methylimidazolium
[AEIM]	1-allyl-3-ethylimidazolium
[C ₁ MIM]	1,3-dimethylimidazolium
[C ₁ C ₁ MIM]	1,2,3-trimethylimidazolium
[C ₂ MIM]	1-ethyl-3-methylimidazolium
[C ₄ MIM]	1-butyl-3-methylimidazolium
[C ₄ C ₁ MIM]	1-butyl-2,3-dimethylimidazolium
[C ₅ MIM]	1-pentyl-3-methylimidazolium
[C ₆ MIM]	1-hexyl-3-methylimidazolium
[C ₈ MIM]	1-octyl-3-methylimidazolium
[C ₁₀ MIM]	1-decyl-3-methylimidazolium
[C ₁₂ MIM]	1-dodecyl-3-methylimidazolium
[C ₁₄ MIM]	1-tetradecyl-3-methylimidazolium
[C ₁₆ MIM]	1-hexadecyl-3-methylimidazolium
[C ₁₆ C ₁ MIM]	1-hexadecyl-2,3-dimethylimidazolium
[HOC ₂ MIM]	1-(2-hydroxyethyl)-3-methylimidazolium
[C ₁ OC ₂ MIM]	1-methoxyethyl-3-methylimidazolium
[C ₂ OC ₂ MIM]	1-ethoxyethyl-3-methylimidazolium
[HOC ₂ OC ₂ MIM]	1-(4-hydroxy-2-oxybutyl)-3-methylimidazolium
[HOC ₃ MIM]	1-(3-hydroxypropyl)-3-methylimidazolium
[HOC ₂ C ₁₄ IM]	1-(2-hydroxyethyl)-3-tetradecylimidazolium
[((HO) ₂ C ₃)C ₁₄ IM]	1-(2',3'-dihydroxy)propyl-3-tetradecylimidazolium
[HOC ₂ C ₁ C ₁₄ IM]	1-(2-hydroxyethyl)-2-methyl-3-tetradecylimidazolium
[HOC ₂ C ₁ C ₁₆ IM]	1-hexadecyl-3-(2-hydroxyethyl)-2-methylimidazolium
[SalOC ₂ MIM]	1-(2-((2-hydroxybenzoyl)oxy)ethyl)-3-methylimidazolium
[SalOC ₃ MIM]	1-(3-((2-hydroxybenzoyl)oxy)propyl)-3-methylimidazolium
[C ₂ Py]	1-ethylpyridinium
[C ₁₆ Py]	1-hexadecylpyridinium, 1-cetylpyridinium
[C ₄ C ₁ Py]	1-butyl-3-methylpyridinium
[C ₈ C ₁ Py]	1-octyl-3-methylpyridinium
[C ₆ C ₆ OCOPy]	1-hexyl-3-hexyloxycarbonylpyridinium
[C ₈ OC ₁ HOPy]	3-hydroxy-1-octyloxymethylpyridinium
[C ₁ C ₃ Pyr]	<i>N</i> -methyl- <i>N</i> -propylpyrrolidinium
[C ₁ C ₄ Pyr]	<i>N</i> -methyl- <i>N</i> -butylpyrrolidinium
[HOC ₂ Pyr]	<i>N</i> -(2-hydroxyethyl)-pyrrolidinium
[C ₂ C ₁ Pip]	1-ethyl-1-methylpiperidinium
[C ₁₀ C ₁ Pip]	1-decyl-1-methylpiperidinium
[C ₁₂ C ₁ Pip]	1-dodecyl-1-methylpiperidinium
[C ₂ C ₁ Mor]	1-ethyl-1-methylmorpholinium

[C ₁₀ C ₁ Mor]	1-decyl-1-methylmorpholinium
[(C ₁) ₄ N]	tetramethylammonium
[(C ₂) ₄ N]	tetraethylammonium
[(C ₄) ₄ N]	tetrabutylammonium
[(C ₁) ₃ C ₄ N]	trimethylbutylammonium
[(C ₁) ₃ C ₈ N]	trimethyloctylammonium
[(C ₁) ₃ C ₁₀ N]	trimethyldecylammonium
[(C ₁) ₃ C ₁₂ N]	trimethyldodecylammonium
[(C ₁) ₃ C ₁₆ N]	trimethylhexadecylammonium, cetyltrimethylammonium
[(C ₄) ₃ C ₁ N]	tributylmethylammonium
[(C ₈) ₃ C ₁ N]	trioctylmethylammonium
[(C ₁₀) ₂ (C ₁) ₂ N]	didecyl dimethylammonium
[(C ₁₁) ₂ (C ₁) ₂ N]	diundecyl dimethylammonium
[Cho]	trimethylethanolammonium, cholinium
[(C ₂) ₃ NH]	triethylammonium
[(HOC ₂) ₃ NH]	triethanolammonium
[(C ₄) ₃ C ₁ P]	tributylmethylphosphonium
[(C ₈) ₃ C ₁ P]	trioctylmethylphosphonium
[(C ₄) ₄ P]	tetrabutylphosphonium
[(C ₆) ₃ C ₁₄ P]	trihexyl(tetradecyl)phosphonium
[HOC ₂ (C ₄) ₃ P]	tributyl(2-hydroxyethyl)phosphonium
[(C ₁) ₂ (C ₁) ₂ G]	1,1,3,3-tetramethylguanidine
[HDBU]	1,8-diazabicyclo[5.4.0]undec-7-en-8-ium
[Benzalk]	benzalkonium
[Benzeth]	benzethonium
[Bupiv]	bupivacainium
[Lid]	lidocainium
[Metf]	metformin
[Pril]	prilocainium
[Proc]	procainium
[ProcA]	procainium amide
[Tet]	tetracycline

Anions of ILs

[Acesulf]	acesulfame
[Amp]	ampicillin
[ASal]	acetylsalicylate
[BF ₄]	tetrafluoroborate
[Br]	bromide
[Bz]	benzoic acid
[HSO ₄]	hydrogensulfate
[C ₁ SO ₄]	methyl sulfate
[C ₂ SO ₄]	ethyl sulfate
[C ₈ SO ₄]	octyl sulfate
[C ₁₀ SO ₄]	decyl sulfate
[C ₁₂ SO ₄]	dodecyl (lauryl) sulfate
[C ₁₈ SO ₄]	octadecyl sulfate
[Dic]	diclofenac
[DMP]	dimethylphosphate
[Doc]	docusate
[Fluf]	flufenamate
[Flurb]	flurbiprofenate
[Ibu]	ibuprofenate
[IBut]	isobutanoate
[ICl ₂]	dichloroiodate
[Lac]	lactate
[Ms]	methanesulfonate
[N(CN) ₂]	dicyanamide
[Nal]	nalidixate
[NO ₃]	nitrate
[NTf ₂]	bis(trifluoromethylsulfonyl)amide
[OAc]	acetate

[OTf]	trifluoromethanesulfonate
[PF ₆]	hexafluorophosphate
[Pro]	propanoate
[Sac]	saccharinate
[Sal]	salicylate
[Sulf]	sulfacetamide
[TFa]	trifluoroacetate
[Theophyl]	theophyllinate
[Trp]	tryptophan

Other

IL	ionic liquid
API-IL	active pharmaceutical ingredient-ionic liquid
BSA	bovine serum albumin
CNT	carbon nanotubes
CPMD	Car-Parinello molecular dynamics
DFT	density functional theory
GED	gas-phase electron diffraction
MD	molecular dynamics
MWCNT	multiwall carbon nanotubes
NGF	nerve growth factor
NSAID	nonsteroidal anti-inflammatory drug
PSA	prostate specific antigen
SANS	small-angle neutron scattering
SWCNT	single-wall carbon nanotubes
TSIL	task-specific ionic liquid

REFERENCES

- (1) Wilkes, J. S.; Zaworotko, M. J. Air and Water Stable 1-Ethyl-3-methylimidazolium Based Ionic Liquids. *J. Chem. Soc., Chem. Commun.* **1992**, 965–967.
- (2) Welton, T. Room-Temperature Ionic Liquids. Solvents for Synthesis and Catalysis. *Chem. Rev.* **1999**, *99*, 2071–2084.
- (3) Wasserscheid, P.; Keim, W. Ionic Liquids—New “Solutions” for Transition Metal Catalysis. *Angew. Chem., Int. Ed.* **2000**, *39*, 3772–3789.
- (4) Huddleston, J. G.; Visser, A. E.; Reichert, W. M.; Willauer, H. D.; Broker, G. A.; Rogers, R. D. Characterization and Comparison of Hydrophilic and Hydrophobic Room Temperature Ionic Liquids Incorporating the Imidazolium Cation. *Green Chem.* **2001**, *3*, 156–164.
- (5) Sheldon, R. Catalytic Reactions in Ionic Liquids. *Chem. Commun.* **2001**, 2399–2407.
- (6) Dupont, J.; de Souza, R. F.; Suarez, P. A. Z. Ionic Liquid (Molten Salt) Phase Organometallic Catalysis. *Chem. Rev.* **2002**, *102*, 3667–3692.
- (7) Zhao, D.; Wu, M.; Kou, Y.; Min, E. Ionic Liquids: Applications in Catalysis. *Catal. Today* **2002**, *74*, 157–189.
- (8) Welton, T. Ionic Liquids in Catalysis. *Coord. Chem. Rev.* **2004**, *248*, 2459–2477.
- (9) Părvulescu, V. I.; Hardacre, C. Catalysis in Ionic Liquids. *Chem. Rev.* **2007**, *107*, 2615–2665.
- (10) van Rantwijk, F.; Sheldon, R. A. Biocatalysis in Ionic Liquids. *Chem. Rev.* **2007**, *107*, 2757–2785.
- (11) Plechkova, N. V.; Seddon, K. R. Applications of Ionic Liquids in the Chemical Industry. *Chem. Soc. Rev.* **2008**, *37*, 123–150.
- (12) Weingartner, H. Understanding Ionic Liquids at the Molecular Level: Facts, Problems, and Controversies. *Angew. Chem., Int. Ed.* **2008**, *47*, 654–670.
- (13) Olivier-Bourbigou, H.; Magna, L.; Morvan, D. Ionic Liquids and Catalysis: Recent Progress from Knowledge to Applications. *Appl. Catal., A* **2010**, *373*, 1–56.
- (14) Hallett, J. P.; Welton, T. Room-Temperature Ionic Liquids: Solvents for Synthesis and Catalysis. 2. *Chem. Rev.* **2011**, *111*, 3508–3576.
- (15) Zhang, Q.; Zhang, S.; Deng, Y. Recent Advances in Ionic Liquid Catalysis. *Green Chem.* **2011**, *13*, 2619–2637.
- (16) Domínguez de María, P. “Nonsolvent” Applications of Ionic Liquids in Biotransformations and Organocatalysis. *Angew. Chem., Int. Ed.* **2008**, *47*, 6960–6968.
- (17) Haumann, M.; Riisager, A. Hydroformylation in Room Temperature Ionic Liquids (RTILs): Catalyst and Process Developments. *Chem. Rev.* **2008**, *108*, 1474–1497.
- (18) Ni, B.; Headley, A. D. Ionic-Liquid-Supported (ILS) Catalysts for Asymmetric Organic Synthesis. *Chem. - Eur. J.* **2010**, *16*, 4426–4436.
- (19) Sawant, A. D.; Raut, D. G.; Darvatkar, N. B.; Salunkhe, M. M. Recent Developments of Task-Specific Ionic Liquids in Organic Synthesis. *Green Chem. Lett. Rev.* **2011**, *4*, 41–54.
- (20) *Ionic Liquids in Biotransformations and Organocatalysis: Solvents and Beyond*; Domínguez de María, P., Ed.; John Wiley & Sons, Inc.: Hoboken, NJ, 2012.
- (21) *Catalysis in Ionic Liquids: From Catalyst Synthesis to Application*; Hardacre, C., Părvulescu, V., Eds.; The Royal Society of Chemistry: Cambridge, 2014.
- (22) *Ionic Liquids (ILs) in Organometallic Catalysis*; Dupont, J., Kollár, L., Eds.; Springer-Verlag: Berlin-Heidelberg, 2015.
- (23) *Environmentally Friendly Syntheses Using Ionic Liquids*; Dupont, J., Itoh, T., Lozano, P., Malhotra, S. V., Eds.; CRC Press: Boca Raton-London-New York, 2015.
- (24) *Green Solvents II: Properties and Applications of Ionic Liquids*; Mohammad, A., Inamuddin, Eds.; Springer-Verlag: Berlin-Heidelberg, 2012.
- (25) Zein El Abedin, S.; Endres, F. Electrodeposition of Metals and Semiconductors in Air- and Water-Stable Ionic Liquids. *ChemPhysChem* **2006**, *7*, 58–61.
- (26) MacFarlane, D. R.; Forsyth, M.; Howlett, P. C.; Pringle, J. M.; Sun, J.; Annat, G.; Neil, W.; Izgorodina, E. I. Ionic Liquids in Electrochemical Devices and Processes: Managing Interfacial Electrochemistry. *Acc. Chem. Res.* **2007**, *40*, 1165–1173.
- (27) Fedorov, M. V.; Kornyshev, A. A. Towards Understanding the Structure and Capacitance of Electrical Double Layer in Ionic Liquids. *Electrochim. Acta* **2008**, *53*, 6835–6840.
- (28) Hapiot, P.; Lagrost, C. Electrochemical Reactivity in Room-Temperature Ionic Liquids. *Chem. Rev.* **2008**, *108*, 2238–2264.
- (29) Armand, M.; Endres, F.; MacFarlane, D. R.; Ohno, H.; Scrosati, B. Ionic-Liquid Materials for the Electrochemical Challenges of the Future. *Nat. Mater.* **2009**, *8*, 621–629.
- (30) Fedorov, M. V.; Kornyshev, A. A. Ionic Liquids at Electrified Interfaces. *Chem. Rev.* **2014**, *114*, 2978–3036.
- (31) *Electrochemistry in Ionic Liquids. Vol. 1: Fundamentals*; Torriero, A. A. J., Ed.; Springer-Verlag: Berlin-Heidelberg, 2015.
- (32) *Electrochemistry in Ionic Liquids. Vol. 2: Applications*; Torriero, A. A. J., Ed.; Springer-Verlag: Berlin-Heidelberg, 2015.
- (33) Pinkert, A.; Marsh, K. N.; Pang, S.; Staiger, M. P. Ionic Liquids and Their Interaction with Cellulose. *Chem. Rev.* **2009**, *109*, 6712–6728.
- (34) Zakrzewska, M. E.; Bogel-Lukasik, E.; Bogel-Lukasik, R. Ionic Liquid-Mediated Formation of 5-Hydroxymethylfurfural – a Promising Biomass-Derived Building Block. *Chem. Rev.* **2011**, *111*, 397–417.
- (35) Patel, D. D.; Lee, J. M. Applications of Ionic Liquids. *Chem. Rec.* **2012**, *12*, 329–355.
- (36) Wang, H.; Gurau, G.; Rogers, R. D. Ionic Liquid Processing of Cellulose. *Chem. Soc. Rev.* **2012**, *41*, 1519–1537.
- (37) Passos, H.; Freire, M. G.; Coutinho, J. A. Ionic Liquid Solutions as Extractive Solvents for Value-Added Compounds from Biomass. *Green Chem.* **2014**, *16*, 4786–4815.
- (38) da Costa Lopes, A. M.; Bogel-Lukasik, R. Acidic Ionic Liquids as Sustainable Approach of Cellulose and Lignocellulosic Biomass Conversion without Additional Catalysts. *ChemSusChem* **2015**, *8*, 947–965.
- (39) Kuchenbuch, A.; Giernoth, R. Ionic Liquids Beyond Simple Solvents: Glimpses at the State of the Art in Organic Chemistry. *ChemistryOpen* **2015**, *4*, 677–681.
- (40) Amarasekara, A. S. Acidic Ionic Liquids. *Chem. Rev.* **2016**, *116*, 6133–6183.

- (41) Mai, N. L.; Kim, C. K.; Park, B.; Park, H.-J.; Lee, S. H.; Koo, Y.-M. Prediction of Cellulose Dissolution in Ionic Liquids Using Molecular Descriptors Based QSAR Model. *J. Mol. Liq.* **2016**, *215*, 541–548.
- (42) Sun, X.; Luo, H.; Dai, S. Ionic Liquids-Based Extraction: a Promising Strategy for the Advanced Nuclear Fuel Cycle. *Chem. Rev.* **2012**, *112*, 2100–2128.
- (43) Zhang, Q.; Shreeve, J. M. Energetic Ionic Liquids as Explosives and Propellant Fuels: a New Journey of Ionic Liquid Chemistry. *Chem. Rev.* **2014**, *114*, 10527–10574.
- (44) Goossens, K.; Lava, K.; Bielawski, C. W.; Binnemans, K. Ionic Liquid Crystals: Versatile Materials. *Chem. Rev.* **2016**, *116*, 4643–4807.
- (45) *Ionic Liquids in Synthesis*; Wasserscheid, P., Welton, T., Eds.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, 2008.
- (46) *Ionic Liquid Applications: Pharmaceuticals, Therapeutics, and Biotechnology*; Malhotra, S. V., Ed.; American Chemical Society: Washington, DC, 2010.
- (47) Lee, S. H.; Doan, T. T. N.; Ha, S. H.; Chang, W.-J.; Koo, Y.-M. Influence of Ionic Liquids as Additives on Sol–Gel Immobilized Lipase. *J. Mol. Catal. B: Enzym.* **2007**, *47*, 129–134.
- (48) Attri, P.; Venkatesu, P.; Kumar, A. Activity and Stability of Alpha-Chymotrypsin in Biocompatible Ionic Liquids: Enzyme Refolding by Triethyl Ammonium Acetate. *Phys. Chem. Chem. Phys.* **2011**, *13*, 2788–2796.
- (49) Jeong, S.; Ha, S. H.; Han, S.-H.; Lim, M.-C.; Kim, S. M.; Kim, Y.-R.; Koo, Y.-M.; So, J.-S.; Jeon, T.-J. Elucidation of Molecular Interactions between Lipid Membranes and Ionic Liquids Using Model Cell Membranes. *Soft Matter* **2012**, *8*, 5501–5506.
- (50) Kumar, A.; Venkatesu, P. Overview of the Stability of Alpha-Chymotrypsin in Different Solvent Media. *Chem. Rev.* **2012**, *112*, 4283–4307.
- (51) Plaquevent, J. C.; Levillain, J.; Guillen, F.; Malhiac, C.; Gaumont, A. C. Ionic Liquids: New Targets and Media for Alpha-Amino Acid and Peptide Chemistry. *Chem. Rev.* **2008**, *108*, 5035–5060.
- (52) Lei, Z.; Chen, B.; Li, C.; Liu, H. Predictive Molecular Thermodynamic Models for Liquid Solvents, Solid Salts, Polymers, and Ionic Liquids. *Chem. Rev.* **2008**, *108*, 1419–1455.
- (53) Dean, P. M.; Pringle, J. M.; MacFarlane, D. R. Structural Analysis of Low Melting Organic Salts: Perspectives on Ionic Liquids. *Phys. Chem. Chem. Phys.* **2010**, *12*, 9144–9153.
- (54) Dupont, J.; Scholten, J. D. On the Structural and Surface Properties of Transition-Metal Nanoparticles in Ionic Liquids. *Chem. Soc. Rev.* **2010**, *39*, 1780–1804.
- (55) Giernoth, R. Task-Specific Ionic Liquids. *Angew. Chem., Int. Ed.* **2010**, *49*, 2834–2839.
- (56) Petkovic, M.; Seddon, K. R.; Rebelo, L. P.; Silva Pereira, C. Ionic Liquids: a Pathway to Environmental Acceptability. *Chem. Soc. Rev.* **2011**, *40*, 1383–1403.
- (57) Tang, S.; Baker, G. A.; Zhao, H. Ether- and Alcohol-Functionalized Task-Specific Ionic Liquids: Attractive Properties and Applications. *Chem. Soc. Rev.* **2012**, *41*, 4030–4066.
- (58) Chatel, G.; MacFarlane, D. R. Ionic Liquids and Ultrasound in Combination: Synergies and Challenges. *Chem. Soc. Rev.* **2014**, *43*, 8132–8149.
- (59) Lei, Z.; Dai, C.; Chen, B. Gas Solubility in Ionic Liquids. *Chem. Rev.* **2014**, *114*, 1289–1326.
- (60) Lei, Z.; Dai, C.; Zhu, J.; Chen, B. Extractive Distillation with Ionic Liquids: A Review. *AIChE J.* **2014**, *60*, 3312–3329.
- (61) Smiglak, M.; Pringle, J. M.; Lu, X.; Han, L.; Zhang, S.; Gao, H.; MacFarlane, D. R.; Rogers, R. D. Ionic Liquids for Energy, Materials, and Medicine. *Chem. Commun.* **2014**, *50*, 9228–9250.
- (62) *Structures and Interactions of Ionic Liquids*; Zhang, S., Wang, J., Zhao, Q., Zhou, Q., Eds.; Springer-Verlag: Berlin-Heidelberg, 2014.
- (63) Zhang, S.; Lu, X.; Zhou, Q.; Li, X.; Zhang, X.; Li, S. *Ionic Liquids: Physicochemical Properties*; Elsevier: Oxford, 2009.
- (64) Greaves, T. L.; Drummond, C. J. Protic Ionic Liquids: Properties and Applications. *Chem. Rev.* **2008**, *108*, 206–237.
- (65) Rupp, A. B. A.; Krossing, I. Ionic Liquids with Weakly Coordinating $[M^{III}(OR^F)_4]^-$ Anions. *Acc. Chem. Res.* **2015**, *48*, 2537–2546.
- (66) Podgoršek, A.; Jacquemin, J.; Pádua, A. A.; Costa Gomes, M. F. Mixing Enthalpy for Binary Mixtures Containing Ionic Liquids. *Chem. Rev.* **2016**, *116*, 6075–6106.
- (67) Hayes, R.; Warr, G. G.; Atkin, R. Structure and Nanostructure in Ionic Liquids. *Chem. Rev.* **2015**, *115*, 6357–6426.
- (68) Dong, K.; Zhang, S. J. Hydrogen Bonds: A Structural Insight into Ionic Liquids. *Chem. - Eur. J.* **2012**, *18*, 2748–2761.
- (69) Giacalone, F.; Gruttadauria, M. Covalently Supported Ionic Liquid Phases: An Advanced Class of Recyclable Catalytic Systems. *ChemCatChem* **2016**, *8*, 664–684.
- (70) Jain, N.; Kumar, A.; Chauhan, S.; Chauhan, S. M. S. Chemical and Biochemical Transformations in Ionic Liquids. *Tetrahedron* **2005**, *61*, 1015–1060.
- (71) Zhao, H.; Malhotra, S. V. Applications of Ionic Liquids in Organic Synthesis. *Aldrichimica Acta* **2002**, *35*, 75–83.
- (72) Gu, Y.; Li, G. Ionic Liquids-Based Catalysis with Solids: State of the Art. *Adv. Synth. Catal.* **2009**, *351*, 817–847.
- (73) Yue, C.; Fang, D.; Liu, L.; Yi, T.-F. Synthesis and Application of Task-Specific Ionic Liquids Used as Catalysts and/or Solvents in Organic Unit Reactions. *J. Mol. Liq.* **2011**, *163*, 99–121.
- (74) *Ionic Liquids in Separation Technology*; De Los Rios, A. P.; Fernandez, F. J. H., Eds.; Elsevier: Oxford, 2014.
- (75) *Ionic Liquids for Better Separation Processes*; Rodríguez, H., Ed.; Springer-Verlag: Berlin-Heidelberg, 2016.
- (76) Vidal, L.; Riekkola, M. L.; Canals, A. Ionic Liquid-Modified Materials for Solid-Phase Extraction and Separation: a Review. *Anal. Chim. Acta* **2012**, *715*, 19–41.
- (77) Cowan, M. G.; Gin, D. L.; Noble, R. D. Poly(ionic liquid)/Ionic Liquid Ion-Gels with High “Free” Ionic Liquid Content: Platform Membrane Materials for CO₂/Light Gas Separations. *Acc. Chem. Res.* **2016**, *49*, 724–732.
- (78) Liu, H.; Liu, Y.; Li, J. Ionic Liquids in Surface Electrochemistry. *Phys. Chem. Chem. Phys.* **2010**, *12*, 1685–1697.
- (79) Su, Y. Z.; Fu, Y. C.; Wei, Y. M.; Yan, J. W.; Mao, B. W. The Electrode/Ionic Liquid Interface: Electric Double Layer and Metal Electrodeposition. *ChemPhysChem* **2010**, *11*, 2764–2778.
- (80) Koel, M. Ionic Liquids in Chemical Analysis. *Crit. Rev. Anal. Chem.* **2005**, *35*, 177–192.
- (81) Sun, P.; Armstrong, D. W. Ionic Liquids in Analytical Chemistry. *Anal. Chim. Acta* **2010**, *661*, 1–16.
- (82) Shiddiky, M. J.; Torriero, A. A. Application of Ionic Liquids in Electrochemical Sensing Systems. *Biosens. Bioelectron.* **2011**, *26*, 1775–1787.
- (83) *Ionic Liquids in Chemical Analysis*; Koel, M., Ed.; CRC Press: Boca Raton-London-New York, 2009.
- (84) Pereira, J. F. B.; Lima, Á. S.; Freire, M. G.; Coutinho, J. A. P. Ionic Liquids as Adjuvants for the Tailored Extraction of Biomolecules in Aqueous Biphasic Systems. *Green Chem.* **2010**, *12*, 1661–1669.
- (85) Poole, C. F.; Poole, S. K. Extraction of Organic Compounds with Room Temperature Ionic Liquids. *J. Chromatogr. A* **2010**, *1217*, 2268–2286.
- (86) Ho, T. D.; Canestraro, A. J.; Anderson, J. L. Ionic Liquids in Solid-Phase Microextraction: a Review. *Anal. Chim. Acta* **2011**, *695*, 18–43.
- (87) Freire, M. G.; Claudio, A. F.; Araujo, J. M.; Coutinho, J. A.; Marrucho, I. M.; Canongia Lopes, J. N.; Rebelo, L. P. Aqueous Biphasic Systems: a Boost Brought about by Using Ionic Liquids. *Chem. Soc. Rev.* **2012**, *41*, 4966–4995.
- (88) *Application, Purification, and Recovery of Ionic Liquids*; Kuzmina, O., Hallett, J., Eds.; Elsevier: Oxford, 2016.
- (89) Mai, N. L.; Ahn, K.; Koo, Y.-M. Methods for Recovery of Ionic Liquids — A Review. *Process Biochem. (Oxford, U. K.)* **2014**, *49*, 872–881.
- (90) Wishart, J. F. Energy Applications of Ionic Liquids. *Energy Environ. Sci.* **2009**, *2*, 956–961.
- (91) Moniruzzaman, M.; Kamiya, N.; Goto, M. Activation and Stabilization of Enzymes in Ionic Liquids. *Org. Biomol. Chem.* **2010**, *8*, 2887–2899.

- (92) Park, S.; Kazlauskas, R. J. Biocatalysis in Ionic Liquids – Advantages Beyond Green Technology. *Curr. Opin. Biotechnol.* **2003**, *14*, 432–437.
- (93) Pinto, P. C. A. G.; Saraiva, M. L. M. F. S.; Lima, J. L. F. C. Oxidoreductase Behavior in Ionic Liquids: a Review. *Anal. Sci.* **2008**, *24*, 1231–1238.
- (94) Quijano, G.; Couvert, A.; Amrane, A. Ionic Liquids: Applications and Future Trends in Bioreactor Technology. *Bioresour. Technol.* **2010**, *101*, 8923–8930.
- (95) Naushad, M.; Alothman, Z. A.; Khan, A. B.; Ali, M. Effect of Ionic Liquid on Activity, Stability, and Structure of Enzymes: a Review. *Int. J. Biol. Macromol.* **2012**, *51*, 555–560.
- (96) Weingärtner, H.; Cabrele, C.; Herrmann, C. How Ionic Liquids Can Help to Stabilize Native Proteins. *Phys. Chem. Chem. Phys.* **2012**, *14*, 415–426.
- (97) Patel, R.; Kumari, M.; Khan, A. B. Recent Advances in the Applications of Ionic Liquids in Protein Stability and Activity: A Review. *Appl. Biochem. Biotechnol.* **2014**, *172*, 3701–3720.
- (98) *Ionic Liquids IV: Not Just Solvents Anymore*; Brennecke, J. F., Rogers, R. D., Seddon, K. R., Eds.; American Chemical Society: Washington, DC, 2007.
- (99) *Ionic Liquids: From Knowledge to Application*; Plechkova, N. V., Rogers, R. D., Seddon, K. R., Eds.; American Chemical Society: Washington, DC, 2010.
- (100) *Ionic Liquids UnCOILed: Critical Expert Overviews*; Seddon, K. R., Plechkova, N. V., Eds.; John Wiley & Sons, Inc.: Hoboken, NJ, 2013.
- (101) *Ionic Liquids Further UnCOILed: Critical Expert Overviews*; Plechkova, N. V., Seddon, K. R., Eds.; John Wiley & Sons, Inc.: Hoboken, NJ, 2014.
- (102) *Ionic Liquids Completely UnCOILed: Critical Expert Overviews*; Plechkova, N. V., Seddon, K. R., Eds.; John Wiley & Sons, Inc.: Hoboken, NJ, 2016.
- (103) Jordan, A.; Gathergood, N. Biodegradation of Ionic Liquids – a Critical Review. *Chem. Soc. Rev.* **2015**, *44*, 8200–8237.
- (104) Deetlefs, M.; Faselow, M.; Seddon, K. R. Ionic Liquids: the View from Mount Improbable. *RSC Adv.* **2016**, *6*, 4280–4288.
- (105) Heckenbach, M. E.; Romero, F. N.; Green, M. D.; Halden, R. U. Meta-Analysis of Ionic Liquid Literature and Toxicology. *Chemosphere* **2016**, *150*, 266–274.
- (106) Shamshina, J. L.; Kelley, S. P.; Gurau, G.; Rogers, R. D. Develop Ionic Liquid Drugs. *Nature* **2015**, *528*, 188–189.
- (107) Lewis, G. N. The Atom and the Molecule. *J. Am. Chem. Soc.* **1916**, *38*, 762–785.
- (108) Pauling, L. The Nature of the Chemical Bond. III. The Transition from One Extreme Bond Type to Another. *J. Am. Chem. Soc.* **1932**, *54*, 988–1003.
- (109) Pauling, L. *The Nature of the Chemical Bond and the Structure of Molecules and Crystals: an Introduction to Modern Structural Chemistry*, 3rd ed.; Cornell University Press: Ithaca, NY, 1960.
- (110) Bader, R. F. W. *Atoms in Molecules: A Quantum Theory*; Clarendon Press: Oxford, 1990.
- (111) Gillespie, R. J.; Popelier, P. L. A. *Chemical Bonding and Molecular Geometry*; Oxford University Press: New York, 2001.
- (112) Popelier, P. L. A. *Atoms in Molecules: An Introduction*; Prentice Hall: New York, 2000.
- (113) Atkins, P.; de Paula, J. *Atkins' Physical Chemistry*, 8th ed.; W.H. Freeman and Company: New York, 2006.
- (114) Reichardt, C.; Welton, T. *Solvents and Solvent Effects in Organic Chemistry*, 4th ed.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, 2011.
- (115) Holloczki, O.; Malberg, F.; Welton, T.; Kirchner, B. On the Origin of Ionicity in Ionic Liquids. Ion Pairing versus Charge Transfer. *Phys. Chem. Chem. Phys.* **2014**, *16*, 16880–16890.
- (116) Chen, X.; Regan, C. K.; Craig, S. L.; Krenske, E. H.; Houk, K. N.; Jorgensen, W. L.; Brauman, J. I. Steric and Solvation Effects in Ionic S_N2 Reactions. *J. Am. Chem. Soc.* **2009**, *131*, 16162–16170.
- (117) Gahlmann, A.; Park, S. T.; Zewail, A. H. Structure of Isolated Biomolecules by Electron Diffraction-Laser Desorption: Uracil and Guanine. *J. Am. Chem. Soc.* **2009**, *131*, 2806–2808.
- (118) Chandra, A. K.; Nguyen, M. T.; Uchamaru, T.; Zeegers-Huyskens, T. DFT Study of the Interaction between Guanine and Water. *J. Mol. Struct.* **2000**, *555*, 61–66.
- (119) Guille, K.; Clegg, W. Anhydrous Guanine: a Synchrotron Study. *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.* **2006**, *62*, o515–o517.
- (120) McCaffrey, P. D.; Mawhorter, R. J.; Turner, A. R.; Brain, P. T.; Rankin, D. W. H. Accurate Equilibrium Structures Obtained from Gas-Phase Electron Diffraction Data: Sodium Chloride. *J. Phys. Chem. A* **2007**, *111*, 6103–6114.
- (121) Bouazizi, S.; Nasr, S.; Jaidane, N.; Bellissent-Funel, M. C. Local Order in Aqueous NaCl Solutions and Pure Water: X-ray Scattering and Molecular Dynamics Simulations Study. *J. Phys. Chem. B* **2006**, *110*, 23515–23523.
- (122) Sirdeshmukh, D. B.; Sirdeshmukh, L.; Subhadra, K. G. *Alkali Halides. A Handbook of Physical Properties*; Springer-Verlag: Berlin-Heidelberg, 2001.
- (123) Friant-Michel, P.; Ruiz-Lopez, M. F. Glycine Dimers: Structure, Stability, and Medium Effects. *ChemPhysChem* **2010**, *11*, 3499–3504.
- (124) Hamad, S.; Hughes, C. E.; Catlow, C. R.; Harris, K. D. Clustering of Glycine Molecules in Aqueous Solution Studied by Molecular Dynamics Simulation. *J. Phys. Chem. B* **2008**, *112*, 7280–7288.
- (125) Huang, J.; Stringfellow, T. C.; Yu, L. Glycine Exists Mainly as Monomers, not Dimers, in Supersaturated Aqueous Solutions: Implications for Understanding Its Crystallization and Polymorphism. *J. Am. Chem. Soc.* **2008**, *130*, 13973–13980.
- (126) Yani, Y.; Chow, P. S.; Tan, R. B. H. Glycine Open Dimers in Solution: New Insights into α -Glycine Nucleation and Growth. *Cryst. Growth Des.* **2012**, *12*, 4771–4778.
- (127) Jönsson, P. G.; Kvik, Å. Precision Neutron Diffraction Structure Determination of Protein and Nucleic Acid Components. III. The Crystal and Molecular Structure of the Amino Acid α -Glycine. *Acta Crystallogr., Sect. B: Struct. Crystallogr. Cryst. Chem.* **1972**, *28*, 1827–1833.
- (128) Hughes, C. E.; Hamad, S.; Harris, K. D. M.; Catlow, C. R. A.; Griffiths, P. C. A Multi-Technique Approach for Probing the Evolution of Structural Properties during Crystallization of Organic Materials from Solution. *Faraday Discuss.* **2007**, *136*, 71–89.
- (129) Renuka Devi, K.; Srinivasan, K. A Novel Approach to Understand the Nucleation Kinetics of α and γ Polymorphs of Glycine from Aqueous Solution in the Presence of a Selective Additive through Charge Compensation Mechanism. *CrystEngComm* **2014**, *16*, 707–722.
- (130) Capito, R. M.; Azevedo, H. S.; Velichko, Y. S.; Mata, A.; Stupp, S. I. Self-Assembly of Large and Small Molecules into Hierarchically Ordered Sacs and Membranes. *Science* **2008**, *319*, 1812–1816.
- (131) Gröhn, F. Soft Matter Nanoparticles with Various Shapes and Functionalities Can Form through Electrostatic Self-Assembly. *Soft Matter* **2010**, *6*, 4296–4302.
- (132) Lemmers, M.; Sprakel, J.; Voets, I. K.; van der Gucht, J.; Cohen Stuart, M. A. Multiresponsive Reversible Gels Based on Charge-Driven Assembly. *Angew. Chem., Int. Ed.* **2010**, *49*, 708–711.
- (133) Willerich, I.; Gröhn, F. Photoswitchable Nanoassemblies by Electrostatic Self-Assembly. *Angew. Chem., Int. Ed.* **2010**, *49*, 8104–8108.
- (134) Hosseinkhani, H.; Hong, P. D.; Yu, D. S. Self-Assembled Proteins and Peptides for Regenerative Medicine. *Chem. Rev.* **2013**, *113*, 4837–4861.
- (135) Ferreira, J. G.; Ramos, L. M.; de Oliveira, A. L.; Orth, E. S.; Neto, B. A. An Ionically Tagged Water-Soluble Artificial Enzyme Promotes the Dephosphorylation Reaction with Nitroimidazole: Enhanced Ionic Liquid Effect and Mechanism. *J. Org. Chem.* **2015**, *80*, 5979–5983.
- (136) Key, H. M.; Dydio, P.; Clark, D. S.; Hartwig, J. F. Abiological Catalysis by Artificial Haem Proteins Containing Noble Metals in Place of Iron. *Nature* **2016**, *534*, 534–537.
- (137) Vriezema, D. M.; Comellas Aragones, M.; Elemans, J. A.; Cornelissen, J. J.; Rowan, A. E.; Nolte, R. J. Self-Assembled Nanoreactors. *Chem. Rev.* **2005**, *105*, 1445–1489.
- (138) Soni, S. K.; Ramanathan, R.; Coloe, P. J.; Bansal, V.; Bhargava, S. K. Self-Assembled Enzyme Capsules in Ionic Liquid [BMIM][BF₄] as

Templating Nanoreactors for Hollow Silica Nanocontainers. *Langmuir* **2010**, *26*, 16020–16024.

(139) Barclay, T. G.; Constantopoulos, K.; Matisons, J. Nanotubes Self-Assembled from Amphiphilic Molecules via Helical Intermediates. *Chem. Rev.* **2014**, *114*, 10217–10291.

(140) Spruijt, E.; van den Berg, S. A.; Cohen Stuart, M. A.; van der Gucht, J. Direct Measurement of the Strength of Single Ionic Bonds between Hydrated Charges. *ACS Nano* **2012**, *6*, 5297–5303.

(141) Knipe, P. C.; Thompson, S.; Hamilton, A. D. Ion-Mediated Conformational Switches. *Chem. Sci.* **2015**, *6*, 1630–1639.

(142) Pyle, A. M. Ribozymes: A Distinct Class of Metalloenzymes. *Science* **1993**, *261*, 709–714.

(143) Sigel, R. K.; Pyle, A. M. Alternative Roles for Metal Ions in Enzyme Catalysis and the Implications for Ribozyme Chemistry. *Chem. Rev.* **2007**, *107*, 97–113.

(144) Anzellotti, A. I.; Farrell, N. P. Zinc Metalloproteins as Medicinal Targets. *Chem. Soc. Rev.* **2008**, *37*, 1629–1651.

(145) Ding, F.; O'Donnell, J.; Xu, Q.; Kang, N.; Goldman, N.; Nedergaard, M. Changes in the Composition of Brain Interstitial Ions Control the Sleep-Wake Cycle. *Science* **2016**, *352*, 550–555.

(146) Kasianowicz, J. J. Introduction to Ion Channels and Disease. *Chem. Rev.* **2012**, *112*, 6215–6217.

(147) Zaydman, M. A.; Silva, J. R.; Cui, J. Ion Channel Associated Diseases: Overview of Molecular Mechanisms. *Chem. Rev.* **2012**, *112*, 6319–6333.

(148) Nadler, M. J.; Hermosura, M. C.; Inabe, K.; Perraud, A. L.; Zhu, Q.; Stokes, A. J.; Kurosaki, T.; Kinet, J. P.; Penner, R.; Scharenberg, A. M.; et al. LTRPC7 is a Mg-ATP-Regulated Divalent Cation Channel Required for Cell Viability. *Nature* **2001**, *411*, 590–595.

(149) Runnels, L. W.; Yue, L.; Clapham, D. E. TRP-PLIK, a Bifunctional Protein with Kinase and Ion Channel Activities. *Science* **2001**, *291*, 1043–1047.

(150) Jin, J.; Wu, L. J.; Jun, J.; Cheng, X.; Xu, H.; Andrews, N. C.; Clapham, D. E. The Channel Kinase, TRPM7, is Required for Early Embryonic Development. *Proc. Natl. Acad. Sci. U. S. A.* **2012**, *109*, E225–E233.

(151) Sah, R.; Mesirca, P.; Van den Boogert, M.; Rosen, J.; Mably, J.; Mangoni, M. E.; Clapham, D. E. Ion Channel-Kinase TRPM7 is Required for Maintaining Cardiac Automaticity. *Proc. Natl. Acad. Sci. U. S. A.* **2013**, *110*, E3037–E3046.

(152) *Mammalian Transient Receptor Potential (TRP) Cation Channels*. Vol. I; Nilius, B., Flockerzi, V., Eds.; Springer-Verlag: Berlin-Heidelberg, 2014.

(153) *Mammalian Transient Receptor Potential (TRP) Cation Channels*. Vol. II; Nilius, B., Flockerzi, V., Eds.; Springer-Verlag: Berlin-Heidelberg, 2014.

(154) Maffeo, C.; Bhattacharya, S.; Yoo, J.; Wells, D.; Aksimentiev, A. Modeling and Simulation of Ion Channels. *Chem. Rev.* **2012**, *112*, 6250–6284.

(155) Robertson, J. W.; Kasianowicz, J. J.; Banerjee, S. Analytical Approaches for Studying Transporters, Channels and Porins. *Chem. Rev.* **2012**, *112*, 6227–6249.

(156) Reiner, J. E.; Balijepalli, A.; Robertson, J. W.; Campbell, J.; Suehle, J.; Kasianowicz, J. J. Disease Detection and Management via Single Nanopore-Based Sensors. *Chem. Rev.* **2012**, *112*, 6431–6451.

(157) Holcman, D.; Yuste, R. The New Nanophysiology: Regulation of Ionic Flow in Neuronal Subcompartments. *Nat. Rev. Neurosci.* **2015**, *16*, 685–692.

(158) Liu, Q.; Xiao, K.; Wen, L.; Lu, H.; Liu, Y.; Kong, X. Y.; Xie, G.; Zhang, Z.; Bo, Z.; Jiang, L. Engineered Ionic Gates for Ion Conduction Based on Sodium and Potassium Activated Nanochannels. *J. Am. Chem. Soc.* **2015**, *137*, 11976–11983.

(159) Jain, T.; Raseria, B. C.; Guerrero, R. J.; Boutillier, M. S.; O'Hern, S. C.; Idrobo, J. C.; Karnik, R. Heterogeneous Sub-Continuum Ionic Transport in Statistically Isolated Graphene Nanopores. *Nat. Nanotechnol.* **2015**, *10*, 1053–1057.

(160) Soberats, B.; Yoshio, M.; Ichikawa, T.; Zeng, X.; Ohno, H.; Ungar, G.; Kato, T. Ionic Switch Induced by a Rectangular-Hexagonal

Phase Transition in Benzenammonium Columnar Liquid Crystals. *J. Am. Chem. Soc.* **2015**, *137*, 13212–13215.

(161) Landolt, H. P.; Holst, S. C. Ionic Control of Sleep and Wakefulness. *Science* **2016**, *352*, 517–518.

(162) Xie, J.; Hase, W. L. Rethinking the S_N2 reaction. *Science* **2016**, *352*, 32–33.

(163) Holbrey, J. D.; Seddon, K. R. Ionic Liquids. *Clean Technol. Environ. Policy* **1999**, *1*, 223–236.

(164) Ananikov, V. P. Characterization of Molecular Systems and Monitoring of Chemical Reactions in Ionic Liquids by Nuclear Magnetic Resonance Spectroscopy. *Chem. Rev.* **2011**, *111*, 418–454.

(165) Khokhlova, E. A.; Kachala, V. V.; Ananikov, V. P. The First Molecular Level Monitoring of Carbohydrate Conversion to 5-Hydroxymethylfurfural in Ionic Liquids. B_2O_3 - an Efficient Dual-Function Metal-Free Promoter for Environmentally Benign Applications. *ChemSusChem* **2012**, *5*, 783–789.

(166) Seitkalieva, M. M.; Grachev, A. A.; Egorova, K. S.; Ananikov, V. P. Nanoscale Organization of Ionic Liquids and Their Interaction with Peptides Probed by ^{13}C NMR Spectroscopy. *Tetrahedron* **2014**, *70*, 6075–6081.

(167) Bankmann, D.; Giernoth, R. Magnetic Resonance Spectroscopy in Ionic Liquids. *Prog. Nucl. Magn. Reson. Spectrosc.* **2007**, *51*, 63–90.

(168) Giernoth, R. NMR Spectroscopy in Ionic Liquids. In *Ionic Liquids*; Kirchner, B., Ed.; Springer-Verlag: Berlin-Heidelberg, 2010; Vol. 290, pp 263–283.

(169) Giernoth, R.; Bankmann, D.; Schlörer, N. High Performance NMR in Ionic Liquids. *Green Chem.* **2005**, *7*, 279–282.

(170) Khemchyan, L. L.; Khokhlova, E. A.; Seitkalieva, M. M.; Ananikov, V. P. Efficient Sustainable Tool for Monitoring Chemical Reactions and Structure Determination in Ionic Liquids by ESI-MS. *ChemistryOpen* **2013**, *2*, 208–214.

(171) Joshi, M. D.; Anderson, J. L. Recent Advances of Ionic Liquids in Separation Science and Mass Spectrometry. *RSC Adv.* **2012**, *2*, 5470–5484.

(172) Alfassi, Z. B.; Huie, R. E.; Milman, B. L.; Neta, P. Electrospray Ionization Mass Spectrometry of Ionic Liquids and Determination of Their Solubility in Water. *Anal. Bioanal. Chem.* **2003**, *377*, 159–164.

(173) Jackson, G. P.; Duckworth, D. C. Electrospray Mass Spectrometry of Undiluted Ionic Liquids. *Chem. Commun.* **2004**, 522–523.

(174) Smiglak, M.; Metlen, A.; Rogers, R. D. The Second Evolution of Ionic Liquids: from Solvents and Separations to Advanced Materials – Energetic Examples from the Ionic Liquid Cookbook. *Acc. Chem. Res.* **2007**, *40*, 1182–1192.

(175) Tran, C. D. Ionic Liquids for and by Analytical Spectroscopy. *Anal. Lett.* **2007**, *40*, 2447–2464.

(176) Maton, C.; De Vos, N.; Stevens, C. V. Ionic Liquid Thermal Stabilities: Decomposition Mechanisms and Analysis Tools. *Chem. Soc. Rev.* **2013**, *42*, 5963–5977.

(177) Berg, R. W. Raman Spectroscopy and Ab-Initio Model Calculations on Ionic Liquids. *Monatsh. Chem.* **2007**, *138*, 1045–1075.

(178) Lei, Z.; Dai, C.; Liu, X.; Xiao, L.; Chen, B. Extension of the UNIFAC Model for Ionic Liquids. *Ind. Eng. Chem. Res.* **2012**, *51*, 12135–12144.

(179) Kirchner, B.; Hollóczki, O.; Canongia Lopes, J. N.; Pádua, A. A. H. Multiresolution Calculation of Ionic Liquids. *Wiley Interdiscip. Rev.: Comput. Mol. Sci.* **2015**, *5*, 202–214.

(180) Sprenger, K. G.; Pfäendner, J. Using Molecular Simulation to Study Biocatalysis in Ionic Liquids. *Methods in Enzymology*; Elsevier: Oxford, 2016; Vol. 577, pp 419–441.

(181) Wilkes, J. S. A Short History of Ionic Liquids—from Molten Salts to Neoteric Solvents. *Green Chem.* **2002**, *4*, 73–80.

(182) Hardacre, C. Application of Exafs to Molten Salts and Ionic Liquid Technology. *Annu. Rev. Mater. Res.* **2005**, *35*, 29–49.

(183) Hardacre, C.; Holbrey, J. D.; Nieuwenhuyzen, M.; Youngs, T. G. Structure and Solvation in Ionic Liquids. *Acc. Chem. Res.* **2007**, *40*, 1146–1155.

- (184) Tariq, M.; Freire, M. G.; Saramago, B.; Coutinho, J. A.; Lopes, J. N.; Rebelo, L. P. Surface Tension of Ionic Liquids and Ionic Liquid Solutions. *Chem. Soc. Rev.* **2012**, *41*, 829–868.
- (185) Dong, K.; Zhang, S.; Wang, J. Understanding the Hydrogen Bonds in Ionic Liquids and Their Roles in Properties and Reactions. *Chem. Commun.* **2016**, *52*, 6744–6764.
- (186) Canongia Lopes, J. N.; Pádua, A. A. Nanostructural Organization in Ionic Liquids. *J. Phys. Chem. B* **2006**, *110*, 3330–3335.
- (187) Hunt, P. A.; Gould, I. R. Structural Characterization of the 1-Butyl-3-methylimidazolium Chloride Ion Pair Using Ab Initio Methods. *J. Phys. Chem. A* **2006**, *110*, 2269–2282.
- (188) Russina, O.; De Santis, S.; Gontrani, L. Micro- and Mesoscopic Structural Features of a Bio-Based Choline-Amino Acid Ionic Liquid. *RSC Adv.* **2016**, *6*, 34737–34743.
- (189) Del Pópolo, M. G.; Voth, G. A. On the Structure and Dynamics of Ionic Liquids. *J. Phys. Chem. B* **2004**, *108*, 1744–1752.
- (190) Hayes, R.; Imberti, S.; Warr, G. G.; Atkin, R. Effect of Cation Alkyl Chain Length and Anion Type on Protic Ionic Liquid Nanostructure. *J. Phys. Chem. C* **2014**, *118*, 13998–14008.
- (191) Weinhold, F.; Klein, R. A. Anti-Electrostatic Hydrogen Bonds. *Angew. Chem., Int. Ed.* **2014**, *53*, 11214–11217.
- (192) Knorr, A.; Ludwig, R. Cation-Cation Clusters in Ionic Liquids: Cooperative Hydrogen Bonding Overcomes Like-Charge Repulsion. *Sci. Rep.* **2015**, *5*, 17505.
- (193) Deetlefs, M.; Hardacre, C.; Nieuwenhuyzen, M.; Padua, A. A.; Sheppard, O.; Soper, A. K. Liquid Structure of the Ionic Liquid 1,3-Dimethylimidazolium Bis[(trifluoromethyl)sulfonyl]amide. *J. Phys. Chem. B* **2006**, *110*, 12055–12061.
- (194) Fujii, K.; Soejima, Y.; Kyoshoin, Y.; Fukuda, S.; Kanzaki, R.; Umabayashi, Y.; Yamaguchi, T.; Ishiguro, S.; Takamuku, T. Liquid Structure of Room-Temperature Ionic Liquid, 1-Ethyl-3-methylimidazolium Bis-(trifluoromethanesulfonyl) imide. *J. Phys. Chem. B* **2008**, *112*, 4329–4336.
- (195) Bhargava, B. L.; Balasubramanian, S. Insights into the Structure and Dynamics of a Room-Temperature Ionic Liquid: Ab Initio Molecular Dynamics Simulation Studies of 1-*n*-Butyl-3-methylimidazolium Hexafluorophosphate ([bmim][PF₆]) and the [bmim][PF₆]-CO₂ Mixture. *J. Phys. Chem. B* **2007**, *111*, 4477–4487.
- (196) Bühl, M.; Chaumont, A.; Schurhammer, R.; Wipff, G. Ab Initio Molecular Dynamics of Liquid 1,3-Dimethylimidazolium Chloride. *J. Phys. Chem. B* **2005**, *109*, 18591–18599.
- (197) Del Pópolo, M. G.; Lynden-Bell, R. M.; Kohanoff, J. Ab Initio Molecular Dynamics Simulation of a Room Temperature Ionic Liquid. *J. Phys. Chem. B* **2005**, *109*, 5895–5902.
- (198) Bhargava, B. L.; Balasubramanian, S. Intermolecular Structure and Dynamics in an Ionic Liquid: A Car-Parrinello Molecular Dynamics Simulation Study of 1,3-Dimethylimidazolium Chloride. *Chem. Phys. Lett.* **2006**, *417*, 486–491.
- (199) Hu, Z.; Margulis, C. J. Heterogeneity in a Room-Temperature Ionic Liquid: Persistent Local Environments and the Red-Edge Effect. *Proc. Natl. Acad. Sci. U. S. A.* **2006**, *103*, 831–836.
- (200) Kashin, A. S.; Galkin, K. I.; Khokhlova, E. A.; Ananikov, V. P. Direct Observation of Self-Organized Water-Containing Structures in the Liquid Phase and Their Influence on 5-(Hydroxymethyl)furfural Formation in Ionic Liquids. *Angew. Chem., Int. Ed.* **2016**, *55*, 2161–2166.
- (201) Frost, D. S.; Nofen, E. M.; Dai, L. L. Particle Self-Assembly at Ionic Liquid-Based Interfaces. *Adv. Colloid Interface Sci.* **2014**, *206*, 92–105.
- (202) Dupont, J. On the Solid, Liquid and Solution Structural Organization of Imidazolium Ionic Liquids. *J. Braz. Chem. Soc.* **2004**, *15*, 341–350.
- (203) Schröder, U.; Wadhawan, J. D.; Compton, R. G.; Marken, F.; Suarez, P. A. Z.; Consorti, C. S.; de Souza, R. F.; Dupont, J. Water-Induced Accelerated Ion Diffusion: Voltammetric Studies in 1-Methyl-3-[2,6-(S)-dimethylocten-2-yl]imidazolium Tetrafluoroborate, 1-Butyl-3-methylimidazolium Tetrafluoroborate and Hexafluorophosphate Ionic Liquids. *New J. Chem.* **2000**, *24*, 1009–1015.
- (204) Cammarata, L.; Kazarian, S. G.; Salter, P. A.; Welton, T. Molecular States of Water in Room Temperature Ionic Liquids. *Phys. Chem. Chem. Phys.* **2001**, *3*, 5192–5200.
- (205) Chang, H. C.; Jiang, J. C.; Liou, Y. C.; Hung, C. H.; Lai, T. Y.; Lin, S. H. Effects of Water and Methanol on the Molecular Organization of 1-Butyl-3-methylimidazolium Tetrafluoroborate as Functions of Pressure and Concentration. *J. Chem. Phys.* **2008**, *129*, 044506.
- (206) Ficke, L. E.; Brennecke, J. F. Interactions of Ionic Liquids and Water. *J. Phys. Chem. B* **2010**, *114*, 10496–10501.
- (207) Almásy, L.; Turmine, M.; Perera, A. Structure of Aqueous Solutions of Ionic Liquid 1-Butyl-3-methylimidazolium Tetrafluoroborate by Small-Angle Neutron Scattering. *J. Phys. Chem. B* **2008**, *112*, 2382–2387.
- (208) Gao, J.; Wagner, N. J. Water Nanocluster Formation in the Ionic Liquid 1-Butyl-3-methylimidazolium Tetrafluoroborate ([C₄mim][BF₄])-D₂O Mixtures. *Langmuir* **2016**, *32*, 5078–5084.
- (209) Salma, U.; Ballirano, P.; Usula, M.; Caminiti, R.; Plechkova, N. V.; Seddon, K. R.; Gontrani, L. A New Insight into the Nanostructure of Alkylammonium Alkanoates Based Ionic Liquids in Water. *Phys. Chem. Chem. Phys.* **2016**, *18*, 11497–11502.
- (210) Snyder, P. W.; Lockett, M. R.; Moustakas, D. T.; Whitesides, G. M. Is it the Shape of the Cavity, or the Shape of the Water in the Cavity? *Eur. Phys. J.: Spec. Top.* **2014**, *223*, 853–891.
- (211) Tu, Y.; Peng, F.; Adawy, A.; Men, Y.; Abdelmohsen, L. K.; Wilson, D. A. Mimicking the Cell: Bio-Inspired Functions of Supramolecular Assemblies. *Chem. Rev.* **2016**, *116*, 2023–2078.
- (212) Hough, W. L.; Smiglak, M.; Rodríguez, H.; Swatloski, R. P.; Spear, S. K.; Daly, D. T.; Pernak, J.; Grisel, J. E.; Carliss, R. D.; Soutullo, M. D.; et al. The Third Evolution of Ionic Liquids: Active Pharmaceutical Ingredients. *New J. Chem.* **2007**, *31*, 1429–1436.
- (213) Tavares, A. P. M.; Rodríguez, M.; Macedo, E. A. New Generations of Ionic Liquids Applied to Enzymatic Biocatalysis. In *Ionic Liquids - New Aspects for the Future*; Kadokawa, J.-i., Ed.; InTech, 2013.
- (214) Pucheault, M.; Vaultier, M. Task Specific Ionic Liquids and Task Specific Onium Salts. In *Ionic Liquids*; Kirchner, B., Ed.; Springer-Verlag: Berlin-Heidelberg, 2010; Vol. 290, pp 83–126.
- (215) Li, A.; Tian, Z.; Yan, T.; Jiang, D. E.; Dai, S. Anion-Functionalized Task-Specific Ionic Liquids: Molecular Origin of Change in Viscosity upon CO₂ Capture. *J. Phys. Chem. B* **2014**, *118*, 14880–14887.
- (216) Chaturvedi, D. Recent Developments on Task Specific Ionic Liquids. *Curr. Org. Chem.* **2011**, *15*, 1236–1248.
- (217) Egorova, K. S.; Ananikov, V. P. Toxicity of Ionic Liquids: Eco(cyto)activity as Complicated, but Unavoidable Parameter for Task-Specific Optimization. *ChemSusChem* **2014**, *7*, 336–360.
- (218) Hough, W. L.; Rogers, R. D. Ionic Liquids Then and Now: From Solvents to Materials to Active Pharmaceutical Ingredients. *Bull. Chem. Soc. Jpn.* **2007**, *80*, 2262–2269.
- (219) Pinto, P. C. A. G.; Saraiva, M. L. M. F. S. Ionic Liquids: a Pharmaceutical Perspective. In *Ionic Liquids: Synthesis, Characterization and Applications*; Brooks, A., Ed.; Nova Science Publishers, Inc., 2014.
- (220) Sivapragasam, M.; Moniruzzaman, M.; Goto, M. Recent Advances in Exploiting Ionic Liquids for Biomolecules: Solubility, Stability and Applications. *Biotechnol. J.* **2016**, *11*, 1000–1013.
- (221) Zhao, D.; Liao, Y.; Zhang, Z. Toxicity of Ionic Liquids. *Clean: Soil, Air, Water* **2007**, *35*, 42–48.
- (222) Pham, T. P.; Cho, C. W.; Yun, Y. S. Environmental Fate and Toxicity of Ionic Liquids: a Review. *Water Res.* **2010**, *44*, 352–372.
- (223) Ferraz, R.; Branco, L. C.; Prudêncio, C.; Noronha, J. P.; Petrovski, Ž. Ionic Liquids as Active Pharmaceutical Ingredients. *ChemMedChem* **2011**, *6*, 975–985.
- (224) Moniruzzaman, M.; Goto, M. Ionic Liquids: Future Solvents and Reagents for Pharmaceuticals. *J. Chem. Eng. Jpn.* **2011**, *44*, 370–381.
- (225) Stoimenovski, J.; MacFarlane, D. R.; Bica, K.; Rogers, R. D. Crystalline vs. Ionic Liquid Salt Forms of Active Pharmaceutical Ingredients: a Position Paper. *Pharm. Res.* **2010**, *27*, 521–526.
- (226) Siodmiak, T.; Marszall, M. P.; Proszowska, A. Ionic Liquids: A New Strategy in Pharmaceutical Synthesis. *Mini-Rev. Org. Chem.* **2012**, *9*, 203–208.

- (227) Cojocar, O. A.; Shamshina, J. L.; Rogers, R. D. Review/Preview: Prodrug Ionic Liquids Combining the Prodrug and Ionic Liquid Strategies to Active Pharmaceutical Ingredients. *Chim. Oggi* **2013**, *31*, 24–28.
- (228) Shamshina, J. L.; Barber, P. S.; Rogers, R. D. Ionic Liquids in Drug Delivery. *Expert Opin. Drug Delivery* **2013**, *10*, 1367–1381.
- (229) Marrucho, I. M.; Branco, L. C.; Rebelo, L. P. Ionic Liquids in Pharmaceutical Applications. *Annu. Rev. Chem. Biomol. Eng.* **2014**, *5*, 527–546.
- (230) Balk, A.; Holzgrabe, U.; Meinel, L. 'Pro et Contra' Ionic Liquid Drugs - Challenges and Opportunities for Pharmaceutical Translation. *Eur. J. Pharm. Biopharm.* **2015**, *94*, 291–304.
- (231) Domingos, S.; Andre, V.; Quaresma, S.; Martins, I. C.; Minas da Piedade, M. F.; Duarte, M. T. New Forms of Old Drugs: Improving Without Changing. *J. Pharm. Pharmacol.* **2015**, *67*, 830–846.
- (232) Dias, A. R.; Costa-Rodrigues, J.; Fernandes, M. H.; Ferraz, R.; Prudêncio, C. Anti-Cancer Potential of Ionic Liquids. *ChemMedChem* **2016**, ePub ahead of print; doi: 10.1002/cmdc.201600480.
- (233) Matzke, M.; Stolte, S.; Thiele, K.; Juffernholz, T.; Arning, J.; Ranke, J.; Welz-Biermann, U.; Jastorff, B. The Influence of Anion Species on the Toxicity of 1-Alkyl-3-Methylimidazolium Ionic Liquids Observed in an (Eco)toxicological Test Battery. *Green Chem.* **2007**, *9*, 1198–1207.
- (234) Steudte, S.; Stepnowski, P.; Cho, C. W.; Thöming, J.; Stolte, S. (Eco)toxicity of Fluoro-Organic and Cyano-Based Ionic Liquid Anions. *Chem. Commun.* **2012**, *48*, 9382–9384.
- (235) Ventura, S. P.; Gonçalves, A. M.; Gonçalves, F.; Coutinho, J. A. Assessing the Toxicity on [C₃mim][Tf₂N] to Aquatic Organisms of Different Trophic Levels. *Aquat. Toxicol.* **2010**, *96*, 290–297.
- (236) Kurnia, K. A.; Sintra, T. E.; Neves, C. M.; Shimizu, K.; Canongia Lopes, J. N.; Gonçalves, F.; Ventura, S. P.; Freire, M. G.; Santos, L. M.; Coutinho, J. A. The effect of the cation alkyl chain branching on mutual solubilities with water and toxicities. *Phys. Chem. Chem. Phys.* **2014**, *16*, 19952–19963.
- (237) Wang, Y.; Li, H.; Han, S. A Theoretical Investigation of the Interactions between Water Molecules and Ionic Liquids. *J. Phys. Chem. B* **2006**, *110*, 24646–24651.
- (238) Ding, Z.-D.; Chi, Z.; Gu, W.-X.; Gu, S.-M.; Wang, H.-J. Theoretical and Experimental Investigation of the Interactions between [emim]Ac and Water Molecules. *J. Mol. Struct.* **2012**, *1015*, 147–155.
- (239) Marekha, B. A.; Bria, M.; Moreau, M.; De Waele, I.; Miannay, F.-A.; Smortsova, Y.; Takamuku, T.; Kalugin, O. N.; Kiselev, M.; Idrissi, A. Intermolecular Interactions in Mixtures of 1-n-Butyl-3-methylimidazolium Acetate and Water: Insights from IR, Raman, NMR Spectroscopy and Quantum Chemistry Calculations. *J. Mol. Liq.* **2015**, *210*, 227–237.
- (240) Ohno, H.; Fujita, K.; Kohno, Y. Is Seven the Minimum Number of Water Molecules per Ion Pair for Assured Biological Activity in Ionic Liquid-Water Mixtures? *Phys. Chem. Chem. Phys.* **2015**, *17*, 14454–14460.
- (241) Bernardes, C. E.; Minas da Piedade, M. E.; Canongia Lopes, J. N. The Structure of Aqueous Solutions of a Hydrophilic Ionic Liquid: the Full Concentration Range of 1-Ethyl-3-methylimidazolium Ethylsulfate and Water. *J. Phys. Chem. B* **2011**, *115*, 2067–2074.
- (242) Bingham, R. J.; Ballone, P. Computational Study of Room-Temperature Ionic Liquids Interacting with a POPC Phospholipid Bilayer. *J. Phys. Chem. B* **2012**, *116*, 11205–11216.
- (243) Lim, G. S.; Zidar, J.; Cheong, D. W.; Jaenicke, S.; Klähn, M. Impact of Ionic Liquids in Aqueous Solution on Bacterial Plasma Membranes Studied with Molecular Dynamics Simulations. *J. Phys. Chem. B* **2014**, *118*, 10444–10459.
- (244) Docherty, K. M.; Kulpa, J. C. F. Toxicity and Antimicrobial Activity of Imidazolium and Pyridinium Ionic Liquids. *Green Chem.* **2005**, *7*, 185–189.
- (245) Pernak, J.; Syguda, A.; Mirska, I.; Pernak, A.; Nawrot, J.; Pradzynska, A.; Griffin, S. T.; Rogers, R. D. Choline-Derivative-Based Ionic Liquids. *Chem. - Eur. J.* **2007**, *13*, 6817–6827.
- (246) Dipeolu, O.; Green, E.; Stephens, G. Effects of Water-Miscible Ionic Liquids on Cell Growth and Nitro Reduction Using *Clostridium sporogenes*. *Green Chem.* **2009**, *11*, 397–401.
- (247) Hough-Troutman, W. L.; Smiglak, M.; Griffin, S.; Matthew Reichert, W.; Mirska, I.; Jodynis-Liebert, J.; Adamska, T.; Nawrot, J.; Stasiewicz, M.; Rogers, R. D.; et al. Ionic Liquids with Dual Biological Function: Sweet and Anti-Microbial, Hydrophobic Quaternary Ammonium-Based Salts. *New J. Chem.* **2009**, *33*, 26–33.
- (248) Papaiconomou, N.; Estager, J.; Traore, Y.; Bauduin, P.; Bas, C.; Legeai, S.; Viboud, S.; Draye, M. Synthesis, Physicochemical Properties, and Toxicity Data of New Hydrophobic Ionic Liquids Containing Dimethylpyridinium and Trimethylpyridinium Cations. *J. Chem. Eng. Data* **2010**, *55*, 1971–1979.
- (249) Cornellias, A.; Perez, L.; Comelles, F.; Ribosa, I.; Manresa, A.; Garcia, M. T. Self-Aggregation and Antimicrobial Activity of Imidazolium and Pyridinium Based Ionic Liquids in Aqueous Solution. *J. Colloid Interface Sci.* **2011**, *355*, 164–171.
- (250) Iwai, N.; Nakayama, K.; Kitazume, T. Antibacterial Activities of Imidazolium, Pyrrolidinium and Piperidinium Salts. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 1728–1730.
- (251) Pretti, C.; Renzi, M.; Focardi, S. E.; Giovani, A.; Monni, G.; Melai, B.; Rajamani, S.; Chiappe, C. Acute Toxicity and Biodegradability of N-Alkyl-N-Methylmorpholinium and N-Alkyl-DABCO Based Ionic Liquids. *Ecotoxicol. Environ. Saf.* **2011**, *74*, 748–753.
- (252) Wang, H.; Malhotra, S. V.; Francis, A. J. Toxicity of Various Anions Associated with Methoxyethyl Methyl Imidazolium-Based Ionic Liquids on *Clostridium sp.* *Chemosphere* **2011**, *82*, 1597–1603.
- (253) Ventura, S. P.; Marques, C. S.; Rosatella, A. A.; Afonso, C. A.; Gonçalves, F.; Coutinho, J. A. Toxicity Assessment of Various Ionic Liquid Families towards *Vibrio fischeri* Marine Bacteria. *Ecotoxicol. Environ. Saf.* **2012**, *76*, 162–168.
- (254) Ventura, S. P.; de Barros, R. L.; Sintra, T.; Soares, C. M.; Lima, A. S.; Coutinho, J. A. Simple Screening Method to Identify Toxic/Non-Toxic Ionic Liquids: Agar Diffusion Test Adaptation. *Ecotoxicol. Environ. Saf.* **2012**, *83*, 55–62.
- (255) Ventura, S. P.; e Silva, F. A.; Gonçalves, A. M.; Pereira, J. L.; Gonçalves, F.; Coutinho, J. A. Ecotoxicity Analysis of Cholinium-Based Ionic Liquids to *Vibrio fischeri* Marine Bacteria. *Ecotoxicol. Environ. Saf.* **2014**, *102*, 48–54.
- (256) Anvari, S.; Hajfarajollah, H.; Mokhtarani, B.; Enayati, M.; Sharifi, A.; Mirzaei, M. Antibacterial and Anti-Adhesive Properties of Ionic Liquids with Various Cationic and Anionic Heads toward Pathogenic Bacteria. *J. Mol. Liq.* **2016**, *221*, 685–690.
- (257) Yu, J.; Zhang, S.; Dai, Y.; Lu, X.; Lei, Q.; Fang, W. Antimicrobial Activity and Cytotoxicity of Piperazinium- and Guanidinium-Based Ionic Liquids. *J. Hazard. Mater.* **2016**, *307*, 73–81.
- (258) Lovejoy, K. S.; Davis, L. E.; McClellan, L. M.; Lillo, A. M.; Welsh, J. D.; Schmidt, E. N.; Sanders, C. K.; Lou, A. J.; Fox, D. T.; Koppisch, A. T.; et al. Evaluation of Ionic Liquids on Phototrophic Microbes and Their Use in Biofuel Extraction and Isolation. *J. Appl. Phycol.* **2013**, *25*, 973–981.
- (259) Luczak, J.; Jungnickel, C.; Łacka, I.; Stolte, S.; Hupka, J. Antimicrobial and Surface Activity of 1-Alkyl-3-Methylimidazolium Derivatives. *Green Chem.* **2010**, *12*, 593–601.
- (260) Santos, A. G.; Ribeiro, B. D.; Alviano, D. S.; Coelho, M. A. Z. Toxicity of Ionic Liquids toward Microorganisms Interesting to the Food Industry. *RSC Adv.* **2014**, *4*, 37157–37163.
- (261) Simmons, C. W.; Reddy, A. P.; Vanderghenst, J. S.; Simmons, B. A.; Singer, S. W. *Bacillus coagulans* Tolerance to 1-Ethyl-3-Methylimidazolium-Based Ionic Liquids in Aqueous and Solid-State Thermophilic Culture. *Biotechnol. Prog.* **2014**, *30*, 311–316.
- (262) Hajfarajollah, H.; Mokhtarani, B.; sharifi, A.; Mirzaei, M.; Afaghi, A. Toxicity of Various Kinds of Ionic Liquids towards the Cell Growth and End Product Formation of the Probiotic Strain, *Propionibacterium freudenreichii*. *RSC Adv.* **2014**, *4*, 13153–13160.
- (263) Zhang, C.; Malhotra, S. V.; Francis, A. J. Toxicity of Ionic Liquids to *Clostridium sp.* and Effects on Uranium Biosorption. *J. Hazard. Mater.* **2014**, *264*, 246–253.
- (264) Ouellet, M.; Datta, S.; Dibble, D. C.; Tamrakar, P. R.; Benke, P. I.; Li, C.; Singh, S.; Sale, K. L.; Adams, P. D.; Keasling, J. D.; et al. Impact of Ionic Liquid Pretreated Plant Biomass on *Saccharomyces cerevisiae* Growth and Biofuel Production. *Green Chem.* **2011**, *13*, 2743–2749.

- (265) Mehmood, N.; Husson, E.; Jacquard, C.; Wewetzer, S.; Buchs, J.; Sarazin, C.; Gosselin, I. Impact of Two Ionic Liquids, 1-Ethyl-3-Methylimidazolium Acetate and 1-Ethyl-3-Methylimidazolium Methylphosphonate, on *Saccharomyces cerevisiae*: Metabolic, Physiologic, and Morphological Investigations. *Biotechnol. Biofuels* **2015**, *8*, 17.
- (266) Shih, S. C.; Gach, P. C.; Sustarich, J.; Simmons, B. A.; Adams, P. D.; Singh, S.; Singh, A. K. A Droplet-to-Digital (D2D) Microfluidic Device for Single Cell Assays. *Lab Chip* **2015**, *15*, 225–236.
- (267) Khudyakov, J. I.; D'Haeseleer, P.; Borglin, S. E.; Deangelis, K. M.; Woo, H.; Lindquist, E. A.; Hazen, T. C.; Simmons, B. A.; Thelen, M. P. Global Transcriptome Response to Ionic Liquid by a Tropical Rain Forest Soil Bacterium, *Enterobacter lignolyticus*. *Proc. Natl. Acad. Sci. U. S. A.* **2012**, *109*, E2173–E2182.
- (268) Ruegg, T. L.; Kim, E. M.; Simmons, B. A.; Keasling, J. D.; Singer, S. W.; Lee, T. S.; Thelen, M. P. An Auto-Inducible Mechanism for Ionic Liquid Resistance in Microbial Biofuel Production. *Nat. Commun.* **2014**, *5*, 3490.
- (269) Frederix, M.; Hutter, K.; Leu, J.; Batth, T. S.; Turner, W. J.; Ruegg, T. L.; Blanch, H. W.; Simmons, B. A.; Adams, P. D.; Keasling, J. D.; et al. Development of a Native *Escherichia coli* Induction System for Ionic Liquid Tolerance. *PLoS One* **2014**, *9*, e101115.
- (270) Reddy, A. P.; Simmons, C. W.; Claypool, J.; Jabusch, L.; Burd, H.; Hadi, M. Z.; Simmons, B. A.; Singer, S. W.; VanderGheynst, J. S. Thermophilic Enrichment of Microbial Communities in the Presence of the Ionic Liquid 1-Ethyl-3-Methylimidazolium Acetate. *J. Appl. Microbiol.* **2012**, *113*, 1362–1370.
- (271) Frederix, M.; Mingardon, F.; Hu, M.; Sun, N.; Pray, T.; Singh, S.; Simmons, B. A.; Keasling, J. D.; Mukhopadhyay, A. Development of an *E. coli* Strain for One-Pot Biofuel Production from Ionic Liquid Pretreated Cellulose and Switchgrass. *Green Chem.* **2016**, *18*, 4189–4197.
- (272) Quijano, G.; Couvert, A.; Amrane, A.; Darracq, G.; Couriol, C.; Le Cloirec, P.; Paquin, L.; Carrié, D. Toxicity and Biodegradability of Ionic Liquids: New Perspectives towards Whole-Cell Biotechnological Applications. *Chem. Eng. J.* **2011**, *174*, 27–32.
- (273) Petkovic, M.; Ferguson, J.; Bohn, A.; Trindade, J.; Martins, I.; Carvalho, M. B.; Leitão, M. C.; Rodrigues, C.; Garcia, H.; Ferreira, R.; et al. Exploring Fungal Activity in the Presence of Ionic Liquids. *Green Chem.* **2009**, *11*, 889–894.
- (274) Singer, S. W.; Reddy, A. P.; Gladden, J. M.; Guo, H.; Hazen, T. C.; Simmons, B. A.; VanderGheynst, J. S. Enrichment, Isolation and Characterization of Fungi Tolerant to 1-Ethyl-3-Methylimidazolium Acetate. *J. Appl. Microbiol.* **2011**, *110*, 1023–1031.
- (275) Pernak, J.; Borucka, N.; Walkiewicz, F.; Markiewicz, B.; Fochtman, P.; Stolte, S.; Steudte, S.; Stepnowski, P. Synthesis, Toxicity, Biodegradability and Physicochemical Properties of 4-Benzyl-4-Methylmorpholinium-Based Ionic Liquids. *Green Chem.* **2011**, *13*, 2901–2910.
- (276) Pace, S.; Ceballos, S. J.; Harrold, D.; Stannard, W.; Simmons, B. A.; Singer, S. W.; Thelen, M. P.; VanderGheynst, J. S. Enrichment of Microbial Communities Tolerant to the Ionic Liquids Tetrabutylphosphonium Chloride and Tributylethylphosphonium Diethylphosphate. *Appl. Microbiol. Biotechnol.* **2016**, *100*, S639–S652.
- (277) Cole, M. R.; Li, M.; El-Zahab, B.; Janes, M. E.; Hayes, D.; Warner, I. M. Design, Synthesis, and Biological Evaluation of Beta-Lactam Antibiotic-Based Imidazolium- and Pyridinium-Type Ionic Liquids. *Chem. Biol. Drug Des.* **2011**, *78*, 33–41.
- (278) Bergamo, V. Z.; Donato, R. K.; Dalla Lana, D. F.; Donato, K. J.; Ortega, G. G.; Schrekker, H. S.; Fuentesfria, A. M. Imidazolium Salts as Antifungal Agents: Strong Antibiofilm Activity Against Multidrug-Resistant *Candida tropicalis* Isolates. *Lett. Appl. Microbiol.* **2015**, *60*, 66–71.
- (279) Carson, L.; Chau, P. K. W.; Earle, M. J.; Gilea, M. A.; Gilmore, B. F.; Gorman, S. P.; McCann, M. T.; Seddon, K. R. Antibiofilm Activities of 1-Alkyl-3-Methylimidazolium Chloride Ionic Liquids. *Green Chem.* **2009**, *11*, 492–497.
- (280) Busetti, A.; Crawford, D. E.; Earle, M. J.; Gilea, M. A.; Gilmore, B. F.; Gorman, S. P.; Laverty, G.; Lowry, A. F.; McLaughlin, M.; Seddon, K. R. Antimicrobial and Antibiofilm Activities of 1-Alkylquinolinium Bromide Ionic Liquids. *Green Chem.* **2010**, *12*, 420–425.
- (281) Venkata Nancharaiyah, Y.; Reddy, G. K.; Lalithamanasa, P.; Venugopalan, V. P. The Ionic Liquid 1-Alkyl-3-Methylimidazolium Demonstrates Comparable Antimicrobial and Antibiofilm Behavior to a Cationic Surfactant. *Biofouling* **2012**, *28*, 1141–1149.
- (282) He, B.; Ou, G.; Zhou, C.; Wang, M.; Chen, S. Antimicrobial Ionic Liquids with Fumarate Anion. *J. Chem.* **2013**, *2013*, 473153.
- (283) Walkiewicz, F.; Materna, K.; Kropacz, A.; Michalczyk, A.; Gwiazdowski, R.; Praczyk, T.; Pernak, J. Multifunctional Long-Alkyl-Chain Quaternary Ammonium Azolate Based Ionic Liquids. *New J. Chem.* **2010**, *34*, 2281–2289.
- (284) Hossain, M. I.; El-Harbawi, M.; Noaman, Y. A.; Bustam, M. A.; Alitheen, N. B.; Affandi, N. A.; Hefter, G.; Yin, C. Y. Synthesis and Anti-Microbial Activity of Hydroxylammonium Ionic Liquids. *Chemosphere* **2011**, *84*, 101–104.
- (285) O'Toole, G. A.; Wathier, M.; Zegans, M. E.; Shanks, R. M. Q.; Kowalski, R.; Grinstaff, M. W. Diphosphonium Ionic Liquids as Broad-Spectrum Antimicrobial Agents. *Cornea* **2012**, *31*, 810–816.
- (286) Brunel, F.; Lautard, C.; Garzino, F.; Giorgio, S.; Raimundo, J. M.; Bolla, J. M.; Camplo, M. Antibacterial Activities of Fluorescent Nano Assembled Triphenylamine Phosphonium Ionic Liquids. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 3770–3773.
- (287) Luo, Y.; Wang, Q.; Lu, Q.; Mu, Q.; Mao, D. An Ionic Liquid Facilitates the Proliferation of Antibiotic Resistance Genes Mediated by Class I Integrons. *Environ. Sci. Technol. Lett.* **2014**, *1*, 266–270.
- (288) Alves, P. C.; Hartmann, D. O.; Nunez, O.; Martins, I.; Gomes, T. L.; Garcia, H.; Galceran, M. T.; Hampson, R.; Becker, J. D.; Silva Pereira, C. Transcriptomic and Metabolomic Profiling of Ionic Liquid Stimuli Unveils Enhanced Secondary Metabolism in *Aspergillus nidulans*. *BMC Genomics* **2016**, *17*, 284.
- (289) Huang, R. T. W.; Peng, K. C.; Shih, H. N.; Lin, G. H.; Chang, T. F.; Hsu, S. J.; Hsu, T. S. T.; Lin, I. J. B. Antimicrobial Properties of Ethoxyether-Functionalized Imidazolium Salts. *Soft Matter* **2011**, *7*, 8392–8400.
- (290) Jing, C.; Mu, L.; Ren, T.; Li, B.; Chen, S.; Nan, W. Effect of 1-Octyl-3-Methylimidazolium Chloride on Cell Replication and Membrane Permeability of *Escherichia coli* DH5 α . *Bull. Environ. Contam. Toxicol.* **2014**, *93*, 60–63.
- (291) Petkovic, M.; Hartmann, D. O.; Adamová, G.; Seddon, K. R.; Rebelo, L. P. N.; Pereira, C. S. Unravelling the Mechanism of Toxicity of Alkyltributylphosphonium Chlorides in *Aspergillus nidulans* Conidia. *New J. Chem.* **2012**, *36*, 56–63.
- (292) Mester, P.; Wagner, M.; Rossmann, P. Antimicrobial Effects of Short Chained Imidazolium-Based Ionic Liquids-Influence of Anion Chaotropicity. *Ecotoxicol. Environ. Saf.* **2015**, *111*, 96–101.
- (293) Bica, K.; Cooke, L. R.; Nugent, P.; Rijksen, C.; Rogers, R. D. Toxic on Purpose: Ionic Liquid Fungicides as Combinatorial Crop Protecting Agents. *Green Chem.* **2011**, *13*, 2344–2346.
- (294) Foksowicz-Flaczyk, J.; Walentowska, J. Antifungal Activity of Ionic Liquid Applied to Linen Fabric. *Int. Biodeterior. Biodegrad.* **2013**, *84*, 412–415.
- (295) Koziróg, A.; Wysocka-Robak, A.; Przybysz, K. Antifungal Activity of Paper Modified with Ionic Liquids. *Fibres Text. East. Eur.* **2015**, *23*, 134–137.
- (296) Seter, M.; Thomson, M. J.; Stoimenovski, J.; MacFarlane, D. R.; Forsyth, M. Dual Active Ionic Liquids and Organic Salts for Inhibition of Microbially Influenced Corrosion. *Chem. Commun.* **2012**, *48*, 5983–5985.
- (297) Gindri, I. M.; Palmer, K. L.; Siddiqui, D. A.; Aghyarian, S.; Frizzo, C. P.; Martins, M. A. P.; Rodrigues, D. C. Evaluation of Mammalian and Bacterial Cell Activity on Titanium Surface Coated with Dicationic Imidazolium-Based Ionic Liquids. *RSC Adv.* **2016**, *6*, 36475–36483.
- (298) Ye, Q.; Gao, T.; Wan, F.; Yu, B.; Pei, X.; Zhou, F.; Xue, Q. Grafting Poly(Ionic Liquid) Brushes for Anti-Bacterial and Anti-Biofouling Applications. *J. Mater. Chem.* **2012**, *22*, 13123–13131.
- (299) Joubert, F.; Yeo, R. P.; Sharples, G. J.; Musa, O. M.; Hodgson, D. R.; Cameron, N. R. Preparation of an Antibacterial Poly(Ionic Liquid)

Graft Copolymer of Hydroxyethyl Cellulose. *Biomacromolecules* **2015**, *16*, 3970–3979.

(300) Majewski, P.; Pernak, A.; Grzymislawski, M.; Iwanik, K.; Pernak, J. Ionic Liquids in Embalming and Tissue Preservation. Can Traditional Formalin-Fixation Be Replaced Safely? *Acta Histochem.* **2003**, *105*, 135–142.

(301) Pernak, A.; Iwanik, K.; Majewski, P.; Grzymislawski, M.; Pernak, J. Ionic Liquids as an Alternative to Formalin in Histopathological Diagnosis. *Acta Histochem.* **2005**, *107*, 149–156.

(302) Zheng, Z.; Xu, Q.; Guo, J.; Qin, J.; Mao, H.; Wang, B.; Yan, F. Structure-Antibacterial Activity Relationships of Imidazolium-Type Ionic Liquid Monomers, Poly(ionic liquids) and Poly(ionic liquid) Membranes: Effect of Alkyl Chain Length and Cations. *ACS Appl. Mater. Interfaces* **2016**, *8*, 12684–12692.

(303) Rahman, M.; O'Donnell, J. M.; Brazel, C. S. Cytotoxicity of Plasticizers and Ionic Liquids Using *Drosophila melanogaster* S2 Cell Culture. *Chem. Eng. Technol.* **2011**, *34*, 429–438.

(304) Cvjetko, M.; Radošević, K.; Tomica, A.; Slivac, I.; Vorkapić-Furač, J.; Srček, V. G. Cytotoxic Effects of Imidazolium Ionic Liquids on Fish and Human Cell Lines. *Arch. Ind. Hyg. Toxicol.* **2012**, *63*, 15–20.

(305) Bubalo, M. C.; Radošević, K.; Srček, V. G.; Das, R. N.; Popelier, P.; Roy, K. Cytotoxicity towards CCO Cells of Imidazolium Ionic Liquids with Functionalized Side Chains: Preliminary QSTR Modeling Using Regression and Classification Based Approaches. *Ecotoxicol. Environ. Saf.* **2015**, *112*, 22–28.

(306) Radošević, K.; Železnjak, J.; Cvjetko Bubalo, M.; Radojčić Redovniković, I.; Slivac, I.; Gaurina Srček, V. Comparative In Vitro Study of Cholinium-Based Ionic Liquids and Deep Eutectic Solvents toward Fish Cell Line. *Ecotoxicol. Environ. Saf.* **2016**, *131*, 30–36.

(307) McLaughlin, M.; Earle, M. J.; Gilea, M. A.; Gilmore, B. F.; Gorman, S. P.; Seddon, K. R. Cytotoxicity of 1-Alkylquinolinium Bromide Ionic Liquids in Murine Fibroblast NIH 3T3 Cells. *Green Chem.* **2011**, *13*, 2794–2800.

(308) Egorova, K. S.; Seitkalieva, M. M.; Posvyatenko, A. V.; Ananikov, V. P. An Unexpected Increase of Toxicity of Amino Acid-Containing Ionic Liquids. *Toxicol. Res.* **2015**, *4*, 152–159.

(309) Li, X.; Ma, J.; Jing, C.; Wang, J. Expression Alterations of Cytochromes P4501A1, 2E1, and 3A, and Their Receptors AhR and PXR Caused by 1-Octyl-3-methylimidazolium Chloride in Mouse Mammary Carcinoma Cells. *Chemosphere* **2013**, *93*, 2488–2492.

(310) Weaver, K. D.; Kim, H. J.; Sun, J.; MacFarlane, D. R.; Elliott, G. D. Cyto-toxicity and Biocompatibility of a Family of Choline Phosphate Ionic Liquids Designed for Pharmaceutical Applications. *Green Chem.* **2010**, *12*, 507–513.

(311) Gindri, I. M.; Siddiqui, D. A.; Bhardwaj, P.; Rodriguez, L. C.; Palmer, K. L.; Frizzo, C. P.; Martins, M. A. P.; Rodrigues, D. C. Dicationic Imidazolium-Based Ionic Liquids: a New Strategy for Non-Toxic and Antimicrobial Materials. *RSC Adv.* **2014**, *4*, 62594–62602.

(312) Ranke, J.; Mölter, K.; Stock, F.; Bottin-Weber, U.; Poczobutt, J.; Hoffmann, J.; Ondruschka, B.; Filser, J.; Jastorff, B. Biological Effects of Imidazolium Ionic Liquids with Varying Chain Lengths in Acute *Vibrio fischeri* and WST-1 Cell Viability Assays. *Ecotoxicol. Environ. Saf.* **2004**, *58*, 396–404.

(313) Pisarova, L.; Steudte, S.; Dorr, N.; Pittenauer, E.; Allmaier, G.; Stepnowski, P.; Stolte, S. Ionic Liquid Long-Term Stability Assessment and Its Contribution to Toxicity and Biodegradation Study of Untreated and Altered Ionic Liquids. *Proc. Inst. Mech. Eng., Part J* **2012**, *226*, 903–922.

(314) Ranke, J.; Müller, A.; Bottin-Weber, U.; Stock, F.; Stolte, S.; Arning, J.; Stormann, R.; Jastorff, B. Lipophilicity Parameters for Ionic Liquid Cations and Their Correlation to In Vitro Cytotoxicity. *Ecotoxicol. Environ. Saf.* **2007**, *67*, 430–438.

(315) Stolte, S.; Arning, J.; Bottin-Weber, U.; Müller, A.; Pitner, W.-R.; Welz-Biermann, U.; Jastorff, B.; Ranke, J. Effects of Different Head Groups and Functionalised Side Chains on the Cytotoxicity of Ionic Liquids. *Green Chem.* **2007**, *9*, 760–767.

(316) Stasiewicz, M.; Mulkiewicz, E.; Tomczak-Wandzel, R.; Kumirska, J.; Siedlecka, E. M.; Golebiowski, M.; Gajdus, J.; Czerwicka, M.; Stepnowski, P. Assessing Toxicity and Biodegradation of Novel,

Environmentally Benign Ionic Liquids (1-Alkoxymethyl-3-hydroxypyridinium Chloride, Saccharinate and Acesulfamates) on Cellular and Molecular Level. *Ecotoxicol. Environ. Saf.* **2008**, *71*, 157–165.

(317) Steudte, S.; Bemowsky, S.; Mahrova, M.; Bottin-Weber, U.; Tojo-Suarez, E.; Stepnowski, P.; Stolte, S. Toxicity and Biodegradability of Dicationic Ionic Liquids. *RSC Adv.* **2014**, *4*, 5198–5205.

(318) Samori, C.; Malferrari, D.; Valbonesi, P.; Montecavalli, A.; Moretti, F.; Galletti, P.; Sartor, G.; Tagliavini, E.; Fabbri, E.; Pasteris, A. Introduction of Oxygenated Side Chain into Imidazolium Ionic Liquids: Evaluation of the Effects at Different Biological Organization Levels. *Ecotoxicol. Environ. Saf.* **2010**, *73*, 1456–1464.

(319) Li, X. Y.; Jing, C. Q.; Lei, W. L.; Li, J.; Wang, J. J. Apoptosis Caused by Imidazolium-Based Ionic Liquids in PC12 Cells. *Ecotoxicol. Environ. Saf.* **2012**, *83*, 102–107.

(320) Li, X. Y.; Jing, C. Q.; Zang, X. Y.; Yang, S.; Wang, J. J. Toxic Cytological Alteration and Mitochondrial Dysfunction in PC12 Cells Induced by 1-Octyl-3-methylimidazolium Chloride. *Toxicol. In Vitro* **2012**, *26*, 1087–1092.

(321) Schaffran, T.; Justus, E.; Elfert, M.; Chen, T.; Gabel, D. Toxicity of *N,N,N*-Trialkylammoniododecaborates as New Anions of Ionic Liquids in Cellular, Liposomal and Enzymatic Test Systems. *Green Chem.* **2009**, *11*, 1458–1464.

(322) Egorova, K. S.; Seitkalieva, M. M.; Posvyatenko, A. V.; Khrustalev, V. N.; Ananikov, V. P. Cytotoxic Activity of Salicylic Acid-Containing Drug Models with Ionic and Covalent Binding. *ACS Med. Chem. Lett.* **2015**, *6*, 1099–1104.

(323) Kumar, V.; Malhotra, S. V. Study on the Potential Anti-Cancer Activity of Phosphonium and Ammonium-Based Ionic Liquids. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4643–4646.

(324) Malhotra, S. V.; Kumar, V. A Profile of the In vitro Anti-Tumor Activity of Imidazolium-Based Ionic Liquids. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 581–585.

(325) Chen, H.-L.; Kao, H.-F.; Wang, J.-Y.; Wei, G.-T. Cytotoxicity of Imidazole Ionic Liquids in Human Lung Carcinoma A549 Cell Line. *J. Chin. Chem. Soc. (Weinheim, Ger.)* **2014**, *61*, 763–769.

(326) Malhotra, S. V.; Kumar, V.; Velez, C.; Zayas, B. Imidazolium-Derived Ionic Salts Induce Inhibition of Cancerous Cell Growth through Apoptosis. *MedChemComm* **2014**, *5*, 1404–1409.

(327) Carrera, G. V. S. M.; Frade, R. F. M.; Aires-de-Sousa, J.; Afonso, C. A. M.; Branco, L. C. Synthesis and Properties of New Functionalized Guanidinium Based Ionic liquids as Non-Toxic Versatile Organic Materials. *Tetrahedron* **2010**, *66*, 8785–8794.

(328) Frade, R. F. M.; Matias, A.; Branco, L. C.; Afonso, C. A. M.; Duarte, C. M. M. Effect of Ionic Liquids on Human Colon Carcinoma HT-29 and CaCo-2 Cell Lines. *Green Chem.* **2007**, *9*, 873–877.

(329) Garcia-Lorenzo, A.; Tojo, E.; Tojo, J.; Teixeira, M.; Rodríguez-Berrolca, F. J.; González, M. P.; Martínez-Zorzano, V. S. Cytotoxicity of Selected Imidazolium-Derived Ionic Liquids in the Human Caco-2 Cell Line. Sub-structural Toxicological Interpretation through a QSAR Study. *Green Chem.* **2008**, *10*, 508–516.

(330) Jaitely, V.; Karatas, A.; Florence, A. T. Water-Immiscible Room Temperature Ionic Liquids (RTILs) as Drug Reservoirs for Controlled Release. *Int. J. Pharm. (Amsterdam, Neth.)* **2008**, *354*, 168–173.

(331) Frade, R. F. M.; Rosatella, A. A.; Marques, C. S.; Branco, L. C.; Kulkarni, P. S.; Mateus, N. M. M.; Afonso, C. A. M.; Duarte, C. M. M. Toxicological Evaluation on Human Colon Carcinoma Cell Line (CaCo-2) of Ionic Liquids Based on Imidazolium, Guanidinium, Ammonium, Phosphonium, Pyridinium and Pyrrolidinium Cations. *Green Chem.* **2009**, *11*, 1660–1665.

(332) Pereiro, A. B.; Araújo, J. M. M.; Martinho, S.; Alves, F.; Nunes, S.; Matias, A.; Duarte, C. M. M.; Rebelo, L. P. N.; Marrucho, I. M. Fluorinated Ionic Liquids: Properties and Applications. *ACS Sustainable Chem. Eng.* **2013**, *1*, 427–439.

(333) Dobler, D.; Schmidts, T.; Klingenhof, I.; Runkel, F. Ionic Liquids as Ingredients in Topical Drug Delivery Systems. *Int. J. Pharm.* **2013**, *441*, 620–627.

(334) Messali, M.; Almtiria, M. N.; Abderrahman, B.; Salghib, R.; Aouad, M. R.; Alshahateet, S. F.; Alia, A. A. S. New Pyridazinium-Based Ionic Liquids: An Eco-Friendly Ultrasound-Assisted Synthesis,

Characterization and Biological Activity. *S. Afr. J. Chem.* **2015**, *68*, 219–225.

(335) Kaushik, N. K.; Attri, P.; Kaushik, N.; Choi, E. H. Synthesis and Antiproliferative Activity of Ammonium and Imidazolium Ionic Liquids against T98G Brain Cancer Cells. *Molecules* **2012**, *17*, 13727–13739.

(336) Gouveia, W.; Jorge, T. F.; Martins, S.; Meireles, M.; Carolino, M.; Cruz, C.; Almeida, T. V.; Araujo, M. E. Toxicity of Ionic Liquids Prepared from Biomaterials. *Chemosphere* **2014**, *104*, 51–56.

(337) Stepnowski, P.; Skladanowski, A. C.; Ludwiczak, A.; Laczynska, E. Evaluating the Cytotoxicity of Ionic Liquids Using Human Cell Line HeLa. *Hum. Exp. Toxicol.* **2004**, *23*, 513–517.

(338) Wang, X.; Ohlin, C. A.; Lu, Q.; Fei, Z.; Hu, J.; Dyson, P. J. Cytotoxicity of Ionic Liquids and Precursor Compounds towards Human Cell Line HeLa. *Green Chem.* **2007**, *9*, 1191–1197.

(339) Rusiecka, I.; Skladanowski, A. C. Induction of the Multixenobiotic/Multidrug Resistance System in HeLa Cells in Response to Imidazolium Ionic Liquids. *Acta Biochim. Polym.* **2011**, *58*, 187–192.

(340) Li, X.; Ma, J.; Wang, J. Cytotoxicity, Oxidative Stress, and Apoptosis in HepG2 Cells Induced by Ionic Liquid 1-Methyl-3-octylimidazolium Bromide. *Ecotoxicol. Environ. Saf.* **2015**, *120*, 342–348.

(341) Hossain, M. I.; Babaa, M.-R.; El-Harbawi, M.; Man, Z.; Hefter, G.; Yin, C.-Y. Synthesis, Characterization, Physical Properties, and Cytotoxicities of 1-(6-Hydroxyhexyl)-3-alkylimidazolium Chloride Ionic Liquids. *J. Chem. Eng. Data* **2011**, *56*, 4188–4193.

(342) Kumar, R. A.; Papaiconomou, N.; Lee, J. M.; Salminen, J.; Clark, D. S.; Prausnitz, J. M. In Vitro Cytotoxicities of Ionic Liquids: Effect of Cation Rings, Functional Groups, and Anions. *Environ. Toxicol.* **2009**, *24*, 388–395.

(343) Salminen, J.; Papaiconomou, N.; Kumar, R. A.; Lee, J.-M.; Kerr, J.; Newman, J.; Prausnitz, J. M. Physicochemical Properties and Toxicities of Hydrophobic Piperidinium and Pyrrolidinium Ionic Liquids. *Fluid Phase Equilib.* **2007**, *261*, 421–426.

(344) Wang, C. L.; Zhu, X. W.; Liu, S. S. Toxicity Studies of Ionic Liquids and Heavy Metal Compounds to MCF-7 and Photobacteria Q67. *Adv. Mater. Res.* **2013**, *610-613*, 721–724.

(345) Gal, N.; Malferrari, D.; Kolusheva, S.; Galletti, P.; Tagliavini, E.; Jelinek, R. Membrane Interactions of Ionic Liquids: Possible Determinants for Biological Activity and Toxicity. *Biochim. Biophys. Acta, Biomembr.* **2012**, *1818*, 2967–2974.

(346) Mosmann, T. Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays. *J. Immunol. Methods* **1983**, *65*, 55–63.

(347) Torrecilla, J. S.; Garcia, J.; Rojo, E.; Rodriguez, F. Estimation of Toxicity of Ionic Liquids in Leukemia Rat Cell Line and Acetylcholinesterase Enzyme by Principal Component Analysis, Neural Networks and Multiple Lineal Regressions. *J. Hazard. Mater.* **2009**, *164*, 182–194.

(348) Fatemi, M. H.; Izadiyan, P. Cytotoxicity Estimation of Ionic Liquids Based on Their Effective Structural Features. *Chemosphere* **2011**, *84*, 553–563.

(349) Paternò, A.; D'Anna, F.; Musumarra, G.; Noto, R.; Scirè, S. A Multivariate Insight into Ionic Liquids Toxicities. *RSC Adv.* **2014**, *4*, 23985–24000.

(350) Sosnowska, A.; Barycki, M.; Zaborowska, M.; Rybinska, A.; Puzyn, T. Towards Designing Environmentally Safe Ionic Liquids: the Influence of the Cation Structure. *Green Chem.* **2014**, *16*, 4749–4757.

(351) de Melo, E. B. A Structure-Activity Relationship Study of the Toxicity of Ionic Liquids Using an Adapted Ferreira-Kiralj Hydrophobicity Parameter. *Phys. Chem. Chem. Phys.* **2015**, *17*, 4516–4523.

(352) Zhang, Z.-b.; Fu, S.-b.; Duan, H.-f.; Lin, Y.-j.; Yang, Y. Brand-new Function of Well-designed Ionic Liquid: Inhibitor of Tumor Cell Growth. *Chem. Res. Chin. Univ.* **2010**, *26*, 757–760.

(353) Jing, B.; Lan, N.; Qiu, J.; Zhu, Y. Interaction of Ionic Liquids with a Lipid Bilayer: A Biophysical Study of Ionic Liquid Cytotoxicity. *J. Phys. Chem. B* **2016**, *120*, 2781–2789.

(354) Benedetto, A.; Ballone, P. Room Temperature Ionic Liquids Interacting with Bio-Molecules: An Overview of Experimental and Computational Studies. *Philos. Mag.* **2016**, *96*, 870–894.

(355) Anastas, P. T.; Warner, J. C. *Green Chemistry: Theory and Practice*; Oxford University Press: Oxford (England), New York, 1998.

(356) Sanderson, K. Chemistry: It's Not Easy Being Green. *Nature* **2011**, *469*, 18–20.

(357) Coleman, D.; Gathergood, N. Biodegradation Studies of Ionic Liquids. *Chem. Soc. Rev.* **2010**, *39*, 600–637.

(358) Stolte, S.; Steudte, S.; Igartua, A.; Stepnowski, P. The Biodegradation of Ionic Liquids - the View from a Chemical Structure Perspective. *Curr. Org. Chem.* **2011**, *15*, 1946–1973.

(359) OECD Guideline for the Testing of Chemicals. OECD, 2005.

(360) Boethling, R. S.; Sommer, E.; DiFiore, D. Designing Small Molecules for Biodegradability. *Chem. Rev.* **2007**, *107*, 2207–2227.

(361) Gendaszewska, D.; Liwarska-Bizukoja, E. Comparison of Imidazolium Ionic Liquids and Traditional Organic Solvents: Effect on Activated Sludge Processes. *Water Sci. Technol.* **2013**, *68*, 2654–2660.

(362) Markiewicz, M.; Piszora, M.; Caicedo, N.; Jungnickel, C.; Stolte, S. Toxicity of Ionic Liquid Cations and Anions towards Activated Sewage Sludge Organisms from Different Sources – Consequences for Biodegradation Testing and Wastewater Treatment Plant Operation. *Water Res.* **2013**, *47*, 2921–2928.

(363) Cho, C. W.; Pham, T. P.; Kim, S.; Song, M. H.; Chung, Y. J.; Yun, Y. S. Three Degradation Pathways of 1-Octyl-3-Methylimidazolium Cation by Activated Sludge from Wastewater Treatment Process. *Water Res.* **2016**, *90*, 294–300.

(364) Gathergood, N.; Garcia, M. T.; Scammells, P. J. Biodegradable Ionic Liquids: Part I. Concept, Preliminary Targets and Evaluation. *Green Chem.* **2004**, *6*, 166–175.

(365) Garcia, M. T.; Gathergood, N.; Scammells, P. J. Biodegradable Ionic Liquids: Part II. Effect of the Anion and Toxicology. *Green Chem.* **2005**, *7*, 9–14.

(366) Gathergood, N.; Scammells, P. J.; Garcia, M. T. Biodegradable Ionic Liquids: Part III. The First Readily Biodegradable Ionic Liquids. *Green Chem.* **2006**, *8*, 156–160.

(367) Docherty, K. M.; Aiello, S. W.; Buehler, B. K.; Jones, S. E.; Szymczyna, B. R.; Walker, K. A. Ionic Liquid Biodegradability Depends on Specific Wastewater Microbial Consortia. *Chemosphere* **2015**, *136*, 160–166.

(368) Liwarska-Bizukoja, E.; Maton, C.; Stevens, C. V. Biodegradation of Imidazolium Ionic Liquids by Activated Sludge Microorganisms. *Biodegradation* **2015**, *26*, 453–463.

(369) Stolte, S.; Abdulkarim, S.; Arning, J.; Blomeyer-Nienstedt, A.-K.; Bottin-Weber, U.; Matzke, M.; Ranke, J.; Jastorff, B.; Thöming, J. Primary Biodegradation of Ionic Liquid Cations, Identification of Degradation Products of 1-Methyl-3-Octylimidazolium Chloride and Electrochemical Wastewater Treatment of Poorly Biodegradable Compounds. *Green Chem.* **2008**, *10*, 214–224.

(370) Liwarska-Bizukoja, E.; Maton, C.; Stevens, C. V.; Gendaszewska, D. Biodegradability and Kinetics of the Removal of New Peralkylated Imidazolium Ionic Liquids. *J. Chem. Technol. Biotechnol.* **2014**, *89*, 763–768.

(371) Deng, Y.; Besse-Hoggan, P.; Sancelme, M.; Delort, A. M.; Husson, P.; Gomes, M. F. Influence of Oxygen Functionalities on the Environmental Impact of Imidazolium Based Ionic Liquids. *J. Hazard. Mater.* **2011**, *198*, 165–174.

(372) Petkovic, M.; Ferguson, J. L.; Gunaratne, H. Q. N.; Ferreira, R.; Leitão, M. C.; Seddon, K. R.; Rebelo, L. P. N.; Pereira, C. S. Novel Biocompatible Cholinium-Based Ionic Liquids - Toxicity and Biodegradability. *Green Chem.* **2010**, *12*, 643–649.

(373) Neumann, J.; Cho, C.-W.; Steudte, S.; Köser, J.; Uerdingen, M.; Thöming, J.; Stolte, S. Biodegradability of Fluoroorganic and Cyano-Based Ionic Liquid Anions under Aerobic and Anaerobic Conditions. *Green Chem.* **2012**, *14*, 410–418.

(374) Neumann, J.; Pawlik, M.; Bryniok, D.; Thoming, J.; Stolte, S. Biodegradation Potential of Cyano-Based Ionic Liquid Anions in a Culture of *Cupriavidus* spp. and Their In Vitro Enzymatic Hydrolysis by Nitrile Hydratase. *Environ. Sci. Pollut. Res.* **2014**, *21*, 9495–9505.

(375) Morrissey, S.; Pegot, B.; Coleman, D.; Garcia, M. T.; Ferguson, D.; Quilty, B.; Gathergood, N. Biodegradable, Non-Bactericidal

Oxygen-Functionalised Imidazolium Esters: A Step towards 'Greener' Ionic Liquids. *Green Chem.* **2009**, *11*, 475–483.

(376) Docherty, K. M.; Joyce, M. V.; Kulacki, K. J.; Kulpa, C. F. Microbial Biodegradation and Metabolite Toxicity of Three Pyridinium-Based Cation Ionic Liquids. *Green Chem.* **2010**, *12*, 701–712.

(377) Neumann, J.; Steudte, S.; Cho, C.-W.; Thöming, J.; Stolte, S. Biodegradability of 27 Pyrrolidinium, Morpholinium, Piperidinium, Imidazolium and Pyridinium Ionic Liquid Cations under Aerobic Conditions. *Green Chem.* **2014**, *16*, 2174–2184.

(378) Trivedi, T. J.; Rao, K. S.; Singh, T.; Mandal, S. K.; Sutradhar, N.; Panda, A. B.; Kumar, A. Task-Specific, Biodegradable Amino Acid Ionic Liquid Surfactants. *ChemSusChem* **2011**, *4*, 604–608.

(379) Romero, A.; Santos, A.; Tojo, J.; Rodríguez, A. Toxicity and Biodegradability of Imidazolium Ionic Liquids. *J. Hazard. Mater.* **2008**, *151*, 268–273.

(380) Gore, R. G.; Myles, L.; Spulak, M.; Beadham, I.; Garcia, T. M.; Connon, S. J.; Gathergood, N. A New Generation of Aprotic Yet Brønsted Acidic Imidazolium Salts: Effect of Ester/Amide Groups in the C-2, C-4 and C-5 on Antimicrobial Toxicity and Biodegradation. *Green Chem.* **2013**, *15*, 2747–2760.

(381) Stolte, S.; Schulz, T.; Cho, C.-W.; Arning, J.; Strassner, T. Synthesis, Toxicity, and Biodegradation of Tunable Aryl Alkyl Ionic Liquids (TAAILs). *ACS Sustainable Chem. Eng.* **2013**, *1*, 410–418.

(382) Thu, H. B. T.; Markiewicz, M.; Thöming, J.; Reich, R. M.; Korinth, V.; Cokoja, M.; Kühn, F. E.; Stolte, S. Catalytically Active Perrhenate Based Ionic Liquids: a Preliminary Ecotoxicity and Biodegradability Assessment. *New J. Chem.* **2015**, *39*, 5431–5436.

(383) Stolte, S.; Steudte, S.; Areitioaurtena, O.; Pagano, F.; Thoming, J.; Stepnowski, P.; Igartua, A. Ionic Liquids as Lubricants or Lubrication Additives: an Ecotoxicity and Biodegradability Assessment. *Chemosphere* **2012**, *89*, 1135–1141.

(384) Abrusci, C.; Palomar, J.; Pablos, J. L.; Rodriguez, F.; Catalina, F. Efficient Biodegradation of Common Ionic Liquids by *Sphingomonas paucimobilis* Bacterium. *Green Chem.* **2011**, *13*, 709–717.

(385) Kroon, M. C.; Buijs, W.; Peters, C. J.; Witkamp, G.-J. Decomposition of Ionic Liquids in Electrochemical Processing. *Green Chem.* **2006**, *8*, 241–245.

(386) Haerens, K.; Matthijs, E.; Binnemans, K.; Van der Bruggen, B. Electrochemical Decomposition of Choline Chloride Based Ionic Liquid Analogues. *Green Chem.* **2009**, *11*, 1357–1365.

(387) Czerwicka, M.; Stolte, S.; Müller, A.; Siedlecka, E. M.; Gołębiowski, M.; Kumirska, J.; Stepnowski, P. Identification of Ionic Liquid Breakdown Products in an Advanced Oxidation System. *J. Hazard. Mater.* **2009**, *171*, 478–483.

(388) Li, X.; Zhao, J.; Li, Q.; Wang, L.; Tsang, S. C. Ultrasonic Chemical Oxidative Degradations of 1,3-Dialkylimidazolium Ionic Liquids and Their Mechanistic Elucidations. *Dalton Trans.* **2007**, 1875–1880.

(389) Zhou, H.; Lv, P.; Shen, Y.; Wang, J.; Fan, J. Identification of Degradation Products of Ionic Liquids in an Ultrasound Assisted Zero-Valent Iron Activated Carbon Micro-Electrolysis System and Their Degradation Mechanism. *Water Res.* **2013**, *47*, 3514–3522.

(390) Zhou, H.; Shen, Y.; Lv, P.; Wang, J.; Li, P. Degradation Pathway and Kinetics of 1-Alkyl-3-Methylimidazolium Bromides Oxidation in an Ultrasonic Nanoscale Zero-Valent Iron/Hydrogen Peroxide System. *J. Hazard. Mater.* **2015**, *284*, 241–252.

(391) Pernak, J.; Branicka, M. Synthesis and Aqueous Ozonation of Some Pyridinium Salts with Alkoxyethyl and Alkylthiomethyl Hydrophobic Groups. *Ind. Eng. Chem. Res.* **2004**, *43*, 1966–1974.

(392) Munoz, M.; Domínguez, C. M.; de Pedro, Z. M.; Quintanilla, A.; Casas, J. A.; Ventura, S. P. M.; Coutinho, J. A. P. Role of the Chemical Structure of Ionic Liquids in Their Ecotoxicity and Reactivity towards Fenton Oxidation. *Sep. Purif. Technol.* **2015**, *150*, 252–256.

(393) Munoz, M.; Domínguez, C. M.; de Pedro, Z. M.; Quintanilla, A.; Casas, J. A.; Rodríguez, J. J. Ionic Liquids Breakdown by Fenton Oxidation. *Catal. Today* **2015**, *240*, 16–21.

(394) Chen, L.; Mullen, G. E.; Le Roch, M.; Cassity, C. G.; Gouault, N.; Fadamiro, H. Y.; Barletta, R. E.; O'Brien, R. A.; Sykora, R. E.;

Stenson, A. C.; et al. On the Formation of a Protic Ionic Liquid in Nature. *Angew. Chem., Int. Ed.* **2014**, *126*, 11762–11765.

(395) Frade, R. F.; Afonso, C. A. Impact of Ionic Liquids in Environment and Humans: an Overview. *Hum. Exp. Toxicol.* **2010**, *29*, 1038–1054.

(396) Biczak, R.; Pawłowska, B.; Balczewski, P.; Rychter, P. The Role of the Anion in the Toxicity of Imidazolium Ionic Liquids. *J. Hazard. Mater.* **2014**, *274*, 181–190.

(397) Cvjetko Bubalo, M.; Hanousek, K.; Radošević, K.; Gaurina Srček, V.; Jakovljević, T.; Radojčić Redovniković, I. Imidazolium Based Ionic Liquids: Effects of Different Anions and Alkyl Chains Lengths on the Barley Seedlings. *Ecotoxicol. Environ. Saf.* **2014**, *101*, 116–123.

(398) Stolte, S.; Matzke, M.; Arning, J.; Bösch, A.; Pitner, W.-R.; Welz-Biermann, U.; Jastorff, B.; Ranke, J. Effects of Different Head Groups and Functionalised Side Chains on the Aquatic Toxicity of Ionic Liquids. *Green Chem.* **2007**, *9*, 1170–1179.

(399) Kulacki, K. J.; Lamberti, G. A. Toxicity of Imidazolium Ionic Liquids to Freshwater Algae. *Green Chem.* **2008**, *10*, 104–110.

(400) Matzke, M.; Stolte, S.; Arning, J.; Uebers, U.; Filser, J. Imidazolium Based Ionic Liquids in Soils: Effects of the Side Chain Length on Wheat (*Triticum aestivum*) and Cress (*Lepidium sativum*) as Affected by Different Clays and Organic Matter. *Green Chem.* **2008**, *10*, 584–591.

(401) Latała, A.; Nędzi, M.; Stepnowski, P. Toxicity of Imidazolium and Pyridinium Based Ionic Liquids towards Algae. *Chlorella vulgaris*, *Oocystis submarina* (Green Algae) and *Cyclotella meneghiniana*, *Skeletonema marinoi* (Diatoms). *Green Chem.* **2009**, *11*, 580–588.

(402) Latała, A.; Nędzi, M.; Stepnowski, P. Toxicity of Imidazolium and Pyridinium Based Ionic Liquids towards Algae. *Bacillaria paxillifer* (a Microphytobenthic Diatom) and *Geitlerinema amphibium* (a Microphytobenthic Blue Green Alga). *Green Chem.* **2009**, *11*, 1371–1376.

(403) Kumar, M.; Trivedi, N.; Reddy, C. R.; Jha, B. Toxic Effects of Imidazolium Ionic Liquids on the Green Seaweed *Ulva lactuca*: Oxidative Stress and DNA Damage. *Chem. Res. Toxicol.* **2011**, *24*, 1882–1890.

(404) Chen, H.; Zou, Y.; Zhang, L.; Wen, Y.; Liu, W. Enantioselective Toxicities of Chiral Ionic Liquids 1-Alkyl-3-Methylimidazolium Lactate to Aquatic Algae. *Aquat. Toxicol.* **2014**, *154*, 114–120.

(405) Liu, T.; Zhu, L.; Xie, H.; Wang, J.; Sun, F.; Wang, F. Effects of the Ionic Liquid 1-Octyl-3-Methylimidazolium Hexafluorophosphate on the Growth of Wheat Seedlings. *Environ. Sci. Pollut. Res.* **2014**, *21*, 3936–3945.

(406) Pawłowska, B.; Biczak, R. Evaluation of the Effect of Tetraethylammonium Bromide and Chloride on the Growth and Development of Terrestrial Plants. *Chemosphere* **2016**, *149*, 24–33.

(407) Biczak, R.; Pawłowska, B.; Telesiński, A.; Ciesielski, W. The Effect of the Number of Alkyl Substituents on Imidazolium Ionic Liquids Phytotoxicity and Oxidative Stress in Spring Barley and Common Radish Seedlings. *Chemosphere* **2016**, *165*, 519–528.

(408) Biczak, R.; Telesiński, A.; Pawłowska, B. Oxidative Stress in Spring Barley and Common Radish Exposed to Quaternary Ammonium Salts with Hexafluorophosphate Anion. *Plant Physiol. Biochem.* **2016**, *107*, 248–256.

(409) Pernak, J.; Czerniak, K.; Niemczak, M.; Chrzanowski, Ł.; Ławniczak, Ł.; Fochtman, P.; Marcinkowska, K.; Praczyk, T. Herbicidal Ionic Liquids Based on Esterquats. *New J. Chem.* **2015**, *39*, 5715–5724.

(410) Pernak, J.; Niemczak, M.; Chrzanowski, Ł.; Ławniczak, Ł.; Fochtman, P.; Marcinkowska, K.; Praczyk, T. Betaine and Carnitine Derivatives as Herbicidal Ionic Liquids. *Chem. - Eur. J.* **2016**, *22*, 12012–12021.

(411) Wang, S. H.; Huang, P. P.; Li, X. Y.; Wang, C. Y.; Zhang, W. H.; Wang, J. J. Embryonic and Developmental Toxicity of the Ionic Liquid 1-Methyl-3-Octylimidazolium Bromide on Goldfish. *Environ. Toxicol.* **2010**, *25*, 243–250.

(412) Li, X. Y.; Zeng, S. H.; Dong, X. Y.; Ma, J. G.; Wang, J. J. Acute Toxicity and Responses of Antioxidant Systems to 1-Methyl-3-Octylimidazolium Bromide at Different Developmental Stages of Goldfish. *Ecotoxicology* **2012**, *21*, 253–259.

- (413) Li, X. Y.; Zeng, S. H.; Zhang, W. H.; Liu, L.; Ma, S.; Wang, J. J. Acute Toxicity and Superficial Damage to Goldfish from the Ionic Liquid 1-Methyl-3-Octylimidazolium Bromide. *Environ. Toxicol.* **2013**, *28*, 207–214.
- (414) Wang, C.; Wei, Z.; Feng, M.; Wang, L.; Wang, Z. Comparative Antioxidant Status in Freshwater Fish *Carassius auratus* Exposed to Eight Imidazolium Bromide Ionic Liquids: a Combined Experimental and Theoretical Study. *Ecotoxicol. Environ. Saf.* **2014**, *102*, 187–195.
- (415) Dong, M.; Liu, T.; Wang, J.; Zhu, L.; Zhang, J. Estimation of the Oxidative Stress and Molecular Damage Caused by 1-Butyl-3-Methylimidazolium Bromide Ionic Liquid in Zebrafish Livers. *J. Biochem. Mol. Toxicol.* **2016**, *30*, 232–238.
- (416) Hafez, N. F. M.; Mutalib, M. I. A.; Bustam, M. A. B.; El-Harbawi, M.; Leveque, J.-M. Ecotoxicity of Pyridinium Based ILs towards Guppy Fish and Four Bacterial Strains. *Procedia Eng.* **2016**, *148*, 830–838.
- (417) Nan, P.; Yan, S.; Wang, Y.; Du, Q.; Chang, Z. Gene Expression Profile Changes Induced by Acute Toxicity of [C₁₆mim]Cl in Loach (*Paramisgurnus dabryanus*). *Environ. Toxicol.* **2016**, ePub ahead of print; doi: 10.1002/tox.22244.
- (418) Ruokonen, S.-K.; Sanwald, C.; Sundvik, M.; Polnick, S.; Vyavaharkar, K.; Duša, F.; Holding, A. J.; King, A. W. T.; Kilpeläinen, L.; Lämmerhofer, M.; et al. Effect of Ionic Liquids on Zebrafish (*Danio rerio*) Viability, Behavior, and Histology; Correlation between Toxicity and Ionic Liquid Aggregation. *Environ. Sci. Technol.* **2016**, *50*, 7116–7125.
- (419) Foulet, A.; Ghanem, O. B.; El-Harbawi, M.; Lévêque, J.-M.; Mutalib, M. I. A.; Yin, C.-Y. Understanding the Physical Properties, Toxicities and Anti-Microbial Activities of Choline-Amino Acid-Based Salts: Low-Toxic Variants of Ionic Liquids. *J. Mol. Liq.* **2016**, *221*, 133–138.
- (420) Thamke, V. R.; Kodam, K. M. Toxicity Study of Ionic Liquid, 1-Butyl-3-methylimidazolium Bromide on Guppy Fish, *Poecilia reticulata* and Its Biodegradation by Soil Bacterium *Rhodococcus hoagii* VRT1. *J. Hazard. Mater.* **2016**, *320*, 408–416.
- (421) Bailey, M. M.; Townsend, M. B.; Jernigan, P. L.; Sturdivant, J.; Hough-Troutman, W. L.; Rasco, J. F.; Swatoski, R. P.; Rogers, R. D.; Hood, R. D. Developmental Toxicity Assessment of the Ionic Liquid 1-Butyl-3-Methylimidazolium Chloride in CD-1 Mice. *Green Chem.* **2008**, *10*, 1213–1217.
- (422) Sipes, I. G.; Knudsen, G. A.; Kuester, R. K. The Effects of Dose and Route on the Toxicokinetics and Disposition of 1-Butyl-3-Methylimidazolium Chloride in Male F-344 Rats and Female B6C3F1 Mice. *Drug Metab. Dispos.* **2008**, *36*, 284–293.
- (423) Jodynis-Liebert, J.; Nowicki, M.; Adamska, T.; Ewertowska, M.; Kujawska, M.; Petzke, E.; Konwerska, A.; Ostalska-Nowicka, D.; Pernak, J. Acute and Subacute (28-Day) Toxicity Studies of Ionic Liquid, Didecylmethyl Ammonium Acesulfamate, in Rats. *Drug Chem. Toxicol.* **2009**, *32*, 395–404.
- (424) Knudsen, G. A.; Cheng, Y.; Kuester, R. K.; Hooth, M. J.; Sipes, I. G. Effects of Dose and Route on the Disposition and Kinetics of 1-Butyl-1-Methylpyrrolidinium Chloride in Male F-344 Rats. *Drug Metab. Dispos.* **2009**, *37*, 2171–2177.
- (425) Bailey, M. M.; Jernigan, P. L.; Henson, M. B.; Sturdivant, J.; Rasco, J. F.; Lovich, A. N.; Lockhard, J. E.; Hough, W. L.; Di Bona, K. R.; Beard, J.; et al. A Comparison of the Effects of Prenatal Exposure of CD-1 Mice to Three Imidazolium-Based Ionic Liquids. *Birth Defects Res, Part B* **2010**, *89*, 233–238.
- (426) Jodynis-Liebert, J.; Nowicki, M.; Murias, M.; Adamska, T.; Ewertowska, M.; Kujawska, M.; Piotrowska, H.; Konwerska, A.; Ostalska-Nowicka, D.; Pernak, J. Cytotoxicity, Acute and Subchronic Toxicity of Ionic Liquid, Didecylmethylammonium Saccharinate, in Rats. *Regul. Toxicol. Pharmacol.* **2010**, *57*, 266–273.
- (427) Dumitrescu, G.; Ciocina, L. P.; Stana, L.; Cretescu, I.; Popescu, R.; Filimon, N. M.; Voia, O. S. Acute Effects of Tetrabutylammonium Chloride Ionic Liquid on the Histological Structure of Liver and Kidney in the Mouse. *Rom. Biotechnol. Lett.* **2014**, *19*, 8925–8934.
- (428) Costello, D. M.; Brown, L. M.; Lambert, G. A. Acute Toxic Effects of Ionic Liquids on Zebra Mussel (*Dreissena polymorpha*) Survival and Feeding. *Green Chem.* **2009**, *11*, 548–553.
- (429) Li, X. Y.; Dong, X. Y.; Bai, X.; Liu, L.; Wang, J. J. The Embryonic and Postembryonic Developmental Toxicity of Imidazolium-Based Ionic Liquids on *Physa acuta*. *Environ. Toxicol.* **2014**, *29*, 697–704.
- (430) Ma, J.; Dong, X.; Fang, Q.; Li, X.; Wang, J. Toxicity of Imidazolium-Based Ionic Liquids on *Physa acuta* and the Snail Antioxidant Stress Response. *J. Biochem. Mol. Toxicol.* **2014**, *28*, 69–75.
- (431) Luo, Y. R.; Wang, S. H.; Yun, M. X.; Li, X. Y.; Wang, J. J.; Sun, Z. J. The Toxic Effects of Ionic Liquids on the Activities of Acetylcholinesterase and Cellulase in Earthworms. *Chemosphere* **2009**, *77*, 313–318.
- (432) Luo, Y. R.; San-Hu, W.; Li, X. Y.; Yun, M. X.; Wang, J. J.; Sun, Z. J. Toxicity of Ionic Liquids on the Growth, Reproductive Ability, and ATPase Activity of Earthworm. *Ecotoxicol. Environ. Saf.* **2010**, *73*, 1046–1050.
- (433) Wu, X.; Tong, Z. H.; Li, L. L.; Yu, H. Q. Toxic Effects of Imidazolium-Based Ionic Liquids on *Caenorhabditis elegans*: the Role of Reactive Oxygen Species. *Chemosphere* **2013**, *93*, 2399–2404.
- (434) Charan, K. T. P.; Ranjan, P.; Manojkumar, K.; Pothanagandhi, N.; Jha, P. C.; Khedkar, V. M.; Sivaramakrishna, A.; Vijayakrishna, K. Evaluation of Imidazolium-Based Ionic Liquids towards Vermicidal Activity: In Vitro & In Silico Studies. *RSC Adv.* **2015**, *5*, 75415–75424.
- (435) Liu, X.; Zhang, S.; Wang, J.; Shao, Y.; Zhu, L. Biochemical Responses and DNA Damage in Earthworms (*Eisenia fetida*) Induced by Ionic Liquid [omim]PF₆. *Environ. Sci. Pollut. Res.* **2016**, *23*, 6836–6844.
- (436) Zhang, H. C.; Shi, C. Y.; Sun, L. Q.; Wang, F.; Chen, G. W. Toxic Effects of Ionic Liquid 1-Octyl-3-methylimidazolium Bromide on the Antioxidant Defense System of Freshwater Planarian, *Dugesia japonica*. *Toxicol. Ind. Health* **2016**, *32*, 1675–1683.
- (437) Zhang, H. C.; Shi, C. Y.; Yang, H. H.; Chen, G. W.; Liu, D. Z. Genotoxicity Evaluation of Ionic Liquid 1-Octyl-3-methylimidazolium Bromide in Freshwater Planarian *Dugesia japonica* Using RAPD Assay. *Ecotoxicol. Environ. Saf.* **2016**, *134P1*, 17–22.
- (438) Zhu, C. J.; Peng, Y.; Tong, Z. H.; Lu, L. Y.; Cui, Y. H.; Yu, H. Q. Hormetic Effect and Mechanism of Imidazolium-Based Ionic Liquids on the Nematode *Caenorhabditis elegans*. *Chemosphere* **2016**, *157*, 65–70.
- (439) Luo, Y. R.; Li, X. Y.; Chen, X. X.; Zhang, B. J.; Sun, Z. J.; Wang, J. J. The Developmental Toxicity of 1-Methyl-3-Octylimidazolium Bromide on *Daphnia magna*. *Environ. Toxicol.* **2008**, *23*, 736–744.
- (440) Bado-Nilles, A.; Diallo, A. O.; Marlar, G.; Pandar, P.; Chabot, L.; Geffard, A.; Len, C.; Porcher, J. M.; Sanchez, W. Coupling of OECD Standardized Test and Immunomarkers to Select the Most Environmentally Benign Ionic Liquids Option – Towards an Innovative “Safety by Design” Approach. *J. Hazard. Mater.* **2015**, *283*, 202–210.
- (441) Samori, C.; Pasteris, A.; Galletti, P.; Tagliavini, E. Acute Toxicity of Oxygenated and Nonoxygenated Imidazolium-Based Ionic Liquids to *Daphnia magna* and *Vibrio fischeri*. *Environ. Toxicol. Chem.* **2007**, *26*, 2379–2382.
- (442) Yu, M.; Wang, S. H.; Luo, Y. R.; Han, Y. W.; Li, X. Y.; Zhang, B. J.; Wang, J. J. Effects of the 1-Alkyl-3-Methylimidazolium Bromide Ionic Liquids on the Antioxidant Defense System of *Daphnia magna*. *Ecotoxicol. Environ. Saf.* **2009**, *72*, 1798–1804.
- (443) Montalbán, M. G.; Hidalgo, J. M.; Collado-González, M.; Díaz Baños, F. G.; Villora, G. Assessing Chemical Toxicity of Ionic Liquids on *Vibrio fischeri*: Correlation with Structure and Composition. *Chemosphere* **2016**, *155*, 405–414.
- (444) Deng, X. Y.; Hu, X. L.; Cheng, J.; Ma, Z. X.; Gao, K. Growth Inhibition and Oxidative Stress Induced by 1-Octyl-3-Methylimidazolium Bromide on the Marine Diatom *Skeletonema costatum*. *Ecotoxicol. Environ. Saf.* **2016**, *132*, 170–177.
- (445) Santos, J. I.; Gonçalves, A. M. M.; Pereira, J. L.; Figueiredo, B. F. H. T.; e Silva, F. A.; Coutinho, J. A. P.; Ventura, S. P. M.; Gonçalves, F. Environmental Safety of Cholinium-Based Ionic Liquids: Assessing Structure–Ecotoxicity Relationships. *Green Chem.* **2015**, *17*, 4657–4668.
- (446) Yoo, B.; Shah, J. K.; Zhu, Y.; Maginn, E. J. Amphiphilic Interactions of Ionic Liquids with Lipid Biomembranes: a Molecular Simulation Study. *Soft Matter* **2014**, *10*, 8641–8651.

- (447) Losada-Pérez, P.; Khorshid, M.; Renner, F. U. Interactions of Aqueous Imidazolium-Based Ionic Liquid Mixtures with Solid-Supported Phospholipid Vesicles. *PLoS One* **2016**, *11*, e0163518.
- (448) Yoo, B.; Jing, B.; Jones, S. E.; Lamberti, G. A.; Zhu, Y.; Shah, J. K.; Maginn, E. J. Molecular Mechanisms of Ionic Liquid Cytotoxicity Probed by an Integrated Experimental and Computational Approach. *Sci. Rep.* **2016**, *6*, 19889.
- (449) Yoo, B.; Zhu, Y.; Maginn, E. J. Molecular Mechanism of Ionic-Liquid-Induced Membrane Disruption: Morphological Changes to Bilayers, Multilayers, and Vesicles. *Langmuir* **2016**, *32*, 5403–5411.
- (450) Cromie, S. R.; Del Popolo, M. G.; Ballone, P. Interaction of Room Temperature Ionic Liquid Solutions with a Cholesterol Bilayer. *J. Phys. Chem. B* **2009**, *113*, 11642–11648.
- (451) Lee, H.; Kim, S. M.; Jeon, T. J. Effects of Imidazolium-Based Ionic Liquids on the Stability and Dynamics of Gramicidin A and Lipid Bilayers at Different Salt Concentrations. *J. Mol. Graphics Modell.* **2015**, *61*, 53–60.
- (452) Ryu, H.; Lee, H.; Iwata, S.; Choi, S.; Kim, M. K.; Kim, Y. R.; Maruta, S.; Kim, S. M.; Jeon, T. J. Investigation of Ion Channel Activities of Gramicidin A in the Presence of Ionic Liquids Using Model Cell Membranes. *Sci. Rep.* **2015**, *5*, 11935.
- (453) Modi, N.; Singh, P. R.; Mahendran, K. R.; Schulz, R.; Winterhalter, M.; Kleinekathöfer, U. Probing the Transport of Ionic Liquids in Aqueous Solution through Nanopores. *J. Phys. Chem. Lett.* **2011**, *2*, 2331–2336.
- (454) Kulkarni, M.; Mukherjee, A. Ionic Liquid Prolongs DNA Translocation through Graphene Nanopores. *RSC Adv.* **2016**, *6*, 46019–46029.
- (455) Galluzzi, M.; Zhang, S.; Mohamadi, S.; Vakurov, A.; Podestà, A.; Nelson, A. Interaction of Imidazolium-Based Room-Temperature Ionic Liquids with DOPC Phospholipid Monolayers: Electrochemical Study. *Langmuir* **2013**, *29*, 6573–6581.
- (456) Weaver, K. D.; Van Vorst, M. P.; Vijayaraghavan, R.; MacFarlane, D. R.; Elliott, G. D. Interaction of Choline Salts with Artificial Biological Membranes: DSC Studies Elucidating Cellular Interactions. *Biochim. Biophys. Acta, Biomembr.* **2013**, *1828*, 1856–1862.
- (457) Kontro, I.; Svedström, K.; Duša, F.; Ahvenainen, P.; Ruokonen, S. K.; Witos, J.; Wiedmer, S. K. Effects of phosphonium-based ionic liquids on phospholipid membranes studied by small-angle X-ray scattering. *Chem. Phys. Lipids* **2016**, *201*, 59–66.
- (458) Bernstein, J. *Polymorphism in Molecular Crystals*; Oxford University Press Inc.: New York, 2002.
- (459) Savjani, K. T.; Gajjar, A. K.; Savjani, J. K. Drug Solubility: Importance and Enhancement Techniques. *ISRN Pharm.* **2012**, *2012*, 195727.
- (460) Jain, S.; Patel, N.; Lin, S. Solubility and Dissolution Enhancement Strategies: Current Understanding and Recent Trends. *Drug Dev. Ind. Pharm.* **2015**, *41*, 875–887.
- (461) Singhal, D.; Curatolo, W. Drug Polymorphism and Dosage Form Design: a Practical Perspective. *Adv. Drug Delivery Rev.* **2004**, *56*, 335–347.
- (462) *Polymorphism in the Pharmaceutical Industry*; Hilfiker, R., Ed.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2006.
- (463) Patel, N. D.; Patel, K. V.; Panchal, L. A.; Shukla, A. K.; Shelat, P. K. An Emerging Technique For Poorly Soluble Drugs: Self Emulsifying Drug Delivery System. *Int. J. Pharm. Biol. Arch.* **2011**, *2*, 621–629.
- (464) Elder, D. P.; Holm, R.; Diego, H. L. Use of Pharmaceutical Salts and Cocrystals to Address the Issue of Poor Solubility. *Int. J. Pharm. (Amsterdam, Neth.)* **2013**, *453*, 88–100.
- (465) Serajuddin, A. T. Salt Formation to Improve Drug Solubility. *Adv. Drug Delivery Rev.* **2007**, *59*, 603–616.
- (466) He, Y.; Ho, C. Amorphous Solid Dispersions: Utilization and Challenges in Drug Discovery and Development. *J. Pharm. Sci.* **2015**, *104*, 3237–3258.
- (467) Feeney, O. M.; Crum, M. F.; McEvoy, C. L.; Trevaskis, N. L.; Williams, H. D.; Pouton, C. W.; Charman, W. N.; Bergstrom, C. A.; Porter, C. J. 50 Years of Oral Lipid-Based Formulations: Provenance, Progress and Future Perspectives. *Adv. Drug Delivery Rev.* **2016**, *101*, 167–194.
- (468) Chang, T.-L.; Zhan, H.; Liang, D.; Liang, J. F. Nanocrystal Technology for Drug Formulation and Delivery. *Front. Chem. Sci. Eng.* **2015**, *9*, 1–14.
- (469) Waterman, K. C.; Adami, R. C.; Alsante, K. M.; Antipas, A. S.; Arenson, D. R.; Carrier, R.; Hong, J.; Landis, M. S.; Lombardo, F.; Shah, J. C.; et al. Hydrolysis in Pharmaceutical Formulations. *Pharm. Dev. Technol.* **2002**, *7*, 113–146.
- (470) Hancock, B. C.; Dalton, C. R. The Effect of Temperature on Water Vapor Sorption by Some Amorphous Pharmaceutical Sugars. *Pharm. Dev. Technol.* **1999**, *4*, 125–131.
- (471) Walker, M. A. Novel Tactics for Designing Water-Soluble Molecules in Drug Discovery. *Expert Opin. Drug Discovery* **2014**, *9*, 1421–1433.
- (472) Jornada, D. H.; Dos Santos Fernandes, G. F.; Chiba, D. E.; de Melo, T. R.; Dos Santos, J. L.; Chung, M. C. The Prodrug Approach: A Successful Tool for Improving Drug Solubility. *Molecules* **2016**, *21*, 42.
- (473) Rautio, J.; Kumpulainen, H.; Heimbach, T.; Oliyai, R.; Oh, D.; Jarvinen, T.; Savolainen, J. Prodrugs: Design and Clinical Applications. *Nat. Rev. Drug Discovery* **2008**, *7*, 255–270.
- (474) Clas, S. D.; Sanchez, R. I.; Nofsinger, R. Chemistry-Enabled Drug Delivery (Prodrugs): Recent Progress and Challenges. *Drug Discovery Today* **2014**, *19*, 79–87.
- (475) Maag, H. Overcoming Poor Permeability – The Role of Prodrugs for Oral Drug Delivery. *Drug Discovery Today: Technol.* **2012**, *9*, e121–e130.
- (476) Mahato, R.; Tai, W.; Cheng, K. Prodrugs for Improving Tumor Targetability and Efficiency. *Adv. Drug Delivery Rev.* **2011**, *63*, 659–670.
- (477) Dahan, A.; Khamis, M.; Agbaria, R.; Karaman, R. Targeted Prodrugs in Oral Drug Delivery: the Modern Molecular Biopharmaceutical Approach. *Expert Opin. Drug Delivery* **2012**, *9*, 1001–1013.
- (478) Fang, J. Y.; Al-Suwayeh, S. A. Nanoparticles as Delivery Carriers for Anticancer Prodrugs. *Expert Opin. Drug Delivery* **2012**, *9*, 657–669.
- (479) Pavan, B.; Dalpiaz, A. Prodrugs and Endogenous Transporters: Are They Suitable Tools for Drug Targeting into the Central Nervous System? *Curr. Pharm. Des.* **2011**, *17*, 3560–3576.
- (480) Staben, L. R.; Koenig, S. G.; Lehar, S. M.; Vandlen, R.; Zhang, D.; Chuh, J.; Yu, S.-F.; Ng, C.; Guo, J.; Liu, Y.; et al. Targeted Drug Delivery through the Traceless Release of Tertiary and Heteroaryl Amines from Antibody–Drug Conjugates. *Nat. Chem.* **2016**, *8*, 1112–1119.
- (481) Iqbal, M.; Zafar, N.; Fessi, H.; Elaissari, A. Double Emulsion Solvent Evaporation Techniques Used for Drug Encapsulation. *Int. J. Pharm. (Amsterdam, Neth.)* **2015**, *496*, 173–190.
- (482) Ulbrich, K.; Holá, K.; Šubr, V.; Bakandritsos, A.; Tuček, J.; Zbořil, R. Targeted Drug Delivery with Polymers and Magnetic Nanoparticles: Covalent and Noncovalent Approaches, Release Control, and Clinical Studies. *Chem. Rev.* **2016**, *116*, 5338–5431.
- (483) Zelikin, A. N.; Ehrhardt, C.; Healy, A. M. Materials and Methods for Delivery of Biological Drugs. *Nat. Chem.* **2016**, *8*, 997–1007.
- (484) Torchilin, V. P. Multifunctional nanocarriers. *Adv. Drug Delivery Rev.* **2012**, *64*, 302–315.
- (485) Sagnella, S. M.; Gong, X.; Moghaddam, M. J.; Conn, C. E.; Kimpton, K.; Waddington, L. J.; Krodziewska, I.; Drummond, C. J. Nanostructured Nanoparticles of Self-Assembled Lipid Pro-Drugs as a Route to Improved Chemotherapeutic Agents. *Nanoscale* **2011**, *3*, 919–924.
- (486) Scalia, S.; Young, P. M.; Traini, D. Solid Lipid Microparticles as an Approach to Drug Delivery. *Expert Opin. Drug Delivery* **2015**, *12*, 583–599.
- (487) *Pharmaceutical Salts: Properties, Selection, and Use*; Stahl, P. H., Wermuth, C. G., Eds.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2011.
- (488) Seitkalieva, M. M.; Kachala, V. V.; Egorova, K. S.; Ananikov, V. P. Molecular Extraction of Peptides in Ionic Liquid Systems. *ACS Sustainable Chem. Eng.* **2015**, *3*, 357–364.
- (489) Rössmann, A. K.; Strassl, K.; Gaertner, P.; Zhao, B.; Greiner, L.; Bica, K. New Aspects for Biomass Processing with Ionic Liquids: Towards the Isolation of Pharmaceutically Active Betulin. *Green Chem.* **2012**, *14*, 940–944.

- (490) Wang, H.; Gurau, G.; Kelley, S. P.; Myerson, A. S.; Rogers, R. D. Hydrophobic vs. Hydrophilic Ionic Liquid Separations Strategies in Support of Continuous Pharmaceutical Manufacturing. *RSC Adv.* **2013**, *3*, 10019–10026.
- (491) Swatloski, R. P.; Spear, S. K.; Holbrey, J. D.; Rogers, R. D. Dissolution of Cellulose with Ionic Liquids. *J. Am. Chem. Soc.* **2002**, *124*, 4974–4975.
- (492) Cláudio, A. F. M.; Ferreira, A. M.; Freire, M. G.; Coutinho, J. A. P. Enhanced Extraction of Caffeine from Guarana Seeds Using Aqueous Solutions of Ionic Liquids. *Green Chem.* **2013**, *15*, 2002–2010.
- (493) Zhao, H. DNA Stability in Ionic Liquids and Deep Eutectic Solvents. *J. Chem. Technol. Biotechnol.* **2015**, *90*, 19–25.
- (494) Garcia, H.; Ferreira, R.; Petkovic, M.; Ferguson, J. L.; Leitão, M. C.; Gunaratne, H. Q. N.; Seddon, K. R.; Rebelo, L. P. N.; Silva Pereira, C. Dissolution of Cork Biopolymers in Biocompatible Ionic Liquids. *Green Chem.* **2010**, *12*, 367–369.
- (495) Cláudio, A. F.; Neves, M. C.; Shimizu, K.; Canongia Lopes, J. N.; Freire, M. G.; Coutinho, J. A. The Magic of Aqueous Solutions of Ionic Liquids: Ionic Liquids as a Powerful Class of Catanionic Hydrotropes. *Green Chem.* **2015**, *17*, 3948–3963.
- (496) Sanan, R.; Kaur, R.; Mahajan, R. K. Micellar Transitions in Catanionic Ionic Liquid–Ibuprofen Aqueous Mixtures; Effects of Composition and Dilution. *RSC Adv.* **2014**, *4*, 64877–64889.
- (497) Alawi, M. A.; Hamdan, I. I.; Sallam, A. A.; Heshmeh, N. A. Solubility Enhancement of Glibenclamide in Choline–Tryptophan Ionic Liquid: Preparation, Characterization and Mechanism of Solubilization. *J. Mol. Liq.* **2015**, *212*, 629–634.
- (498) Mahajan, S.; Sharma, R.; Mahajan, R. K. An Investigation of Drug Binding Ability of a Surface Active Ionic Liquid: Micellization, Electrochemical, and Spectroscopic Studies. *Langmuir* **2012**, *28*, 17238–17246.
- (499) Jin, W.; Yang, Q.; Zhang, Z.; Bao, Z.; Ren, Q.; Yang, Y.; Xing, H. Self-Assembly Induced Solubilization of Drug-Like Molecules in Nanostructured Ionic Liquids. *Chem. Commun.* **2015**, *51*, 13170–13173.
- (500) Washiro, S.; Hanatani, A. Preparation for External Use Comprising an Ionic Liquid. EP1795188, 2010.
- (501) Atanasoska, L.; Holman, T. J.; Weber, J.; Warner, R.; Schewe, S. Internal Medical Devices for Delivery of Therapeutic Agent in Conjunction With a Source of Electrical Power. US8152759, 2012.
- (502) Shekaari, H.; Zafarani-Moattar, M. T.; Mirheydari, S. N. Thermodynamic Study of Aspirin in the Presence of Ionic Liquid, 1-Hexyl-3-Methylimidazolium Bromide in Acetonitrile at T=(288.15 to 318.15) K. *J. Mol. Liq.* **2015**, *209*, 138–148.
- (503) Shekaari, H.; Moattar, M. T. Z.; Ghaffari, F. Solvation Properties of Acetaminophen in Aqueous Ionic Liquid, 1-Hexyl-3-Methylimidazolium Bromide, Solutions at Different Temperatures. *J. Mol. Liq.* **2015**, *202*, 86–94.
- (504) Mizuuchi, H.; Jaitely, V.; Murdan, S.; Florence, A. T. Room Temperature Ionic Liquids and Their Mixtures: Potential Pharmaceutical Solvents. *Eur. J. Pharm. Sci.* **2008**, *33*, 326–331.
- (505) dos Santos, A. D.; Morais, A. R. C.; Melo, C.; Bogel-Lukasik, R.; Bogel-Lukasik, E. Solubility of Pharmaceutical Compounds in Ionic Liquids. *Fluid Phase Equilib.* **2013**, *356*, 18–29.
- (506) Forte, A.; Melo, C. I.; Bogel-Lukasik, R.; Bogel-Lukasik, E. A Favorable Solubility of Isoniazid, an Antitubercular Antibiotic Drug, in Alternative Solvents. *Fluid Phase Equilib.* **2012**, *318*, 89–95.
- (507) Lourenço, C.; Melo, C. I.; Bogel-Lukasik, R.; Bogel-Lukasik, E. Solubility Advantage of Pyrazine-2-carboxamide: Application of Alternative Solvents on the Way to the Future Pharmaceutical Development. *J. Chem. Eng. Data* **2012**, *57*, 1525–1533.
- (508) Faria, R. A.; da Ponte, M. N.; Bogel-Lukasik, E. Solubility Studies on the System of Trihexyl(tetradecyl)phosphonium Bis-[(trifluoromethyl)sulfonyl]amide Ionic Liquid and Pharmaceutical and Bioactive Compounds. *Fluid Phase Equilib.* **2015**, *385*, 1–9.
- (509) Faria, R. A.; Bogel-Lukasik, E. Solubilities of Pharmaceutical and Bioactive Compounds in Trihexyl(tetradecyl)phosphonium Chloride Ionic Liquid. *Fluid Phase Equilib.* **2015**, *397*, 18–25.
- (510) Mehrdad, A.; Miri, A. H. Aqueous Solubility of Acetaminophen in the Presence of 1-Hexyl-3-methylimidazolium Bromide, Ionic Liquid as Co-Solvent. *Fluid Phase Equilib.* **2016**, *425*, 51–56.
- (511) Mehrdad, A.; Miri, A. H. Influence of 1-Butyl-3-methylimidazolium Bromide, Ionic Liquid as Co-solvent on Aqueous Solubility of Acetaminophen. *J. Mol. Liq.* **2016**, *221*, 1162–1167.
- (512) Ramos-Rodríguez, D.-A.; Rodríguez-Hidalgo, M.-d.-R.; Soto-Figueroa, C.; Vicente, L. Molecular and Mesoscopic Study of Ionic Liquids and Their Use as Solvents of Active Agents Released by Polymeric Vehicles. *Mol. Phys.* **2010**, *108*, 657–665.
- (513) Melo, C. I.; Bogel-Lukasik, R.; Nunes da Ponte, M.; Bogel-Lukasik, E. Ammonium Ionic Liquids as Green Solvents for Drugs. *Fluid Phase Equilib.* **2013**, *338*, 209–216.
- (514) McCrary, P. D.; Beasley, P. A.; Gurau, G.; Narita, A.; Barber, P. S.; Cojocar, O. A.; Rogers, R. D. Drug Specific, Tuning of an Ionic Liquid's Hydrophilic–Lipophilic Balance to Improve Water Solubility of Poorly Soluble Active Pharmaceutical Ingredients. *New J. Chem.* **2013**, *37*, 2196–2202.
- (515) Williams, H. D.; Sahbaz, Y.; Ford, L.; Nguyen, T. H.; Scammells, P. J.; Porter, C. J. Ionic Liquids Provide Unique Opportunities for Oral Drug Delivery: Structure Optimization and In Vivo Evidence of Utility. *Chem. Commun.* **2014**, *50*, 1688–1690.
- (516) Shi, S.; Yin, T.; Tao, X.; Shen, W. Light Induced Micelle to Vesicle Transition in an Aqueous Solution of a Surface Active Ionic Liquid. *RSC Adv.* **2015**, *5*, 75806–75809.
- (517) Tian, T.; Qin, J.; Gao, Y.; Yu, L. Experimental and DFT Studies on Aggregation Behavior of Dodecylsulfonate-Based Surface Active Ionic Liquids in Water and Ethylammonium Nitrate. *J. Mol. Liq.* **2016**, *218*, 457–464.
- (518) Nakamura, R.; Tokuda, M.; Suzuki, T.; Minami, H. Preparation of Poly(ionic liquid) Hollow Particles with Switchable Permeability. *Langmuir* **2016**, *32*, 2331–2337.
- (519) Kuchlyan, J.; Kundu, N. sarkar, N. Ionic Liquids in Micro-emulsions: Formulation and Characterization. *Curr. Opin. Colloid Interface Sci.* **2016**, *25*, 27–38.
- (520) Pal, A.; Yadav, A. Binding Interactions of Anesthetic Drug with Surface Active Ionic Liquid. *J. Mol. Liq.* **2016**, *222*, 471–479.
- (521) Kubota, K.; Shibata, A.; Yamaguchi, T. The Molecular Assembly of the Ionic Liquid/Aliphatic Carboxylic Acid/Aliphatic Amine as Effective and Safety Transdermal Permeation Enhancers. *Eur. J. Pharm. Sci.* **2016**, *86*, 75–83.
- (522) Araki, S.; Wakabayashi, R.; Moniruzzaman, M.; Kamiya, N.; Goto, M. Ionic Liquid-Mediated Transcutaneous Protein Delivery with Solid-in-Oil Nanodispersions. *MedChemComm* **2015**, *6*, 2124–2128.
- (523) Monti, D.; Egiziano, E.; Burgalassi, S.; Chetoni, P.; Chiappe, C.; Sanzone, A.; Tampucci, S. Ionic Liquids as Potential Enhancers for Transdermal Drug Delivery. *Int. J. Pharm.* **2017**, *516*, 45–51.
- (524) Zakrewsky, M.; Lovejoy, K. S.; Kern, T. L.; Miller, T. E.; Le, V.; Nagy, A.; Goumas, A. M.; Iyer, R. S.; Del Sesto, R. E.; Koppisch, A. T.; et al. Ionic Liquids as a Class of Materials for Transdermal Delivery and Pathogen Neutralization. *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111*, 13313–13318.
- (525) Pei, Y.; Huang, Y.; Li, L.; Wang, J. Phase Behaviour and Microstructure of the Micro-Emulsions Composed of Cholinium-Based Ionic Liquid, Triton X-100 and Water. *J. Chem. Thermodyn.* **2014**, *74*, 231–237.
- (526) Pei, Y.; Zhang, J.; Song, X.; Zhao, M.; Wang, J. Partition Behavior of Drug Molecules in Cholinium-Based Ionic Liquids. *Sep. Sci. Technol. (Philadelphia, PA, U. S.)* **2015**, *50*, 1641–1646.
- (527) Patra, D.; Barakat, C. Unique Role of Ionic Liquid [bmin][BF₄] during Curcumin-Surfactant Association and Micellization of Cationic, Anionic and Non-ionic Surfactant Solutions. *Spectrochim. Acta, Part A* **2011**, *79*, 1823–1828.
- (528) Ghatak, C.; Rao, V. G.; Mandal, S.; Ghosh, S.; Sarkar, N. An Understanding of the Modulation of Photophysical Properties of Curcumin inside a Micelle Formed by an Ionic Liquid: a New Possibility of Tunable Drug Delivery System. *J. Phys. Chem. B* **2012**, *116*, 3369–3379.

- (529) Moniruzzaman, M.; Tahara, Y.; Tamura, M.; Kamiya, N.; Goto, M. Ionic Liquid-Assisted Transdermal Delivery of Sparingly Soluble Drugs. *Chem. Commun.* **2010**, *46*, 1452–1454.
- (530) Moniruzzaman, M.; Kamiya, N.; Goto, M. Ionic Liquid Based Microemulsion with Pharmaceutically Accepted Components: Formulation and Potential Applications. *J. Colloid Interface Sci.* **2010**, *352*, 136–142.
- (531) Moniruzzaman, M.; Tamura, M.; Tahara, Y.; Kamiya, N.; Goto, M. Ionic Liquid-in-Oil Microemulsion as a Potential Carrier of Sparingly Soluble Drug: Characterization and Cytotoxicity Evaluation. *Int. J. Pharm. (Amsterdam, Neth.)* **2010**, *400*, 243–250.
- (532) Goindi, S.; Arora, P.; Kumar, N.; Puri, A. Development of Novel Ionic Liquid-Based Microemulsion Formulation for Dermal Delivery of 5-Fluorouracil. *AAPS PharmSciTech* **2014**, *15*, 810–821.
- (533) Goindi, S.; Kaur, R. An Ionic Liquid-in-Water Microemulsion as a Potential Carrier for Topical Delivery of Poorly Water Soluble Drug: Development, Ex-vivo and In-vivo Evaluation. *Int. J. Pharm. (Amsterdam, Neth.)* **2015**, *495*, 913–923.
- (534) Khan, A. B.; Ali, M.; Malik, N. A.; Ali, A.; Patel, R. Role of 1-Methyl-3-Octylimidazolium Chloride in the Micellization Behavior of Amphiphilic Drug Amitriptyline Hydrochloride. *Colloids Surf., B* **2013**, *112*, 460–465.
- (535) Patel, R.; Khan, A. B.; Dohare, N.; Maroof Ali, M.; Rajor, H. K. Mixed Micellization and Interfacial Properties of Ionic Liquid-Type Imidazolium Gemini Surfactant with Amphiphilic Drug Amitriptyline Hydrochloride and its Thermodynamics. *J. Surfactants Deterg.* **2015**, *18*, 719–728.
- (536) Azevedo, A. M.; Ribeiro, D. M.; Pinto, P. C.; Lucio, M.; Reis, S.; Saraiva, M. L. Imidazolium Ionic Liquids as Solvents of Pharmaceuticals: Influence on HSA Binding and Partition Coefficient of Nimesulide. *Int. J. Pharm. (Amsterdam, Neth.)* **2013**, *443*, 273–278.
- (537) Dandpat, S. S.; Sarkar, M. Investigating the Molecular and Aggregated States of a Drug Molecule Rutaecarpine Using Spectroscopy, Microscopy, Crystallography and Computational Studies. *Phys. Chem. Chem. Phys.* **2015**, *17*, 13992–14002.
- (538) Kundu, N.; Banik, D.; Roy, A.; Kuchlyan, J.; Sarkar, N. Modulation of the Aggregation Properties of Sodium Deoxycholate in Presence of Hydrophilic Imidazolium Based Ionic Liquid: Water Dynamics Study to Probe the Structural Alteration of the Aggregates. *Phys. Chem. Chem. Phys.* **2015**, *17*, 25216–25227.
- (539) Rao, V. G.; Mandal, S.; Ghosh, S.; Banerjee, C.; Sarkar, N. Ionic Liquid-in-Oil Microemulsions Composed of Double Chain Surface Active Ionic Liquid as a Surfactant: Temperature Dependent Solvent and Rotational Relaxation Dynamics of Coumarin-153 in [Py][TF₂N]/[C₄mim][AOT]/Benzene Microemulsions. *J. Phys. Chem. B* **2012**, *116*, 8210–8221.
- (540) Rao, V. G.; Banerjee, C.; Ghosh, S.; Mandal, S.; Kuchlyan, J.; Sarkar, N. A Step Toward the Development of High-Temperature Stable Ionic Liquid-in-Oil Microemulsions Containing Double-Chain Anionic Surface Active Ionic Liquid. *J. Phys. Chem. B* **2013**, *117*, 7472–7480.
- (541) Kabanov, A. V.; Vinogradov, S. V. Nanogels as Pharmaceutical Carriers: Finite Networks of Infinite Capabilities. *Angew. Chem., Int. Ed.* **2009**, *48*, 5418–5429.
- (542) Klyachko, N. L.; Manickam, D. S.; Brynskikh, A. M.; Uglanova, S. V.; Li, S.; Higginbotham, S. M.; Bronich, T. K.; Batrakova, E. V.; Kabanov, A. V. Cross-Linked Antioxidant Nanozymes for Improved Delivery to CNS. *Nanomedicine* **2012**, *8*, 119–129.
- (543) Yi, X.; Manickam, D. S.; Brynskikh, A.; Kabanov, A. V. Agile Delivery of Protein Therapeutics to CNS. *J. Controlled Release* **2014**, *190*, 637–663.
- (544) Cui, W.; Lu, X.; Cui, K.; Niu, L.; Wei, Y.; Lu, Q. Dual-Responsive Controlled Drug Delivery Based on Ionically Assembled Nanoparticles. *Langmuir* **2012**, *28*, 9413–9420.
- (545) Viau, L.; Tourne-Peteilh, C.; Devoisselle, J. M.; Vioux, A. Ionogels as Drug Delivery System: One-Step Sol-Gel Synthesis Using Imidazolium Ibuprofenate Ionic Liquid. *Chem. Commun.* **2010**, *46*, 228–230.
- (546) Le Bideau, J.; Viau, L.; Vioux, A. Ionogels, Ionic Liquid Based Hybrid Materials. *Chem. Soc. Rev.* **2011**, *40*, 907–925.
- (547) Winther-Jensen, O.; Vijayaraghavan, R.; Sun, J.; Winther-Jensen, B.; MacFarlane, D. R. Self Polymerising Ionic Liquid Gel. *Chem. Commun.* **2009**, 3041–3043.
- (548) Kumar, S. S.; Surianarayanan, M.; Vijayaraghavan, R.; Mandal, A. B.; MacFarlane, D. R. Curcumin Loaded Poly(2-Hydroxyethyl Methacrylate) Nanoparticles from Gelled Ionic Liquid – In Vitro Cytotoxicity and Anti-Cancer Activity in SKOV-3 Cells. *Eur. J. Pharm. Sci.* **2014**, *51*, 34–44.
- (549) Demirci, S.; Sahiner, N. PEI-Based Ionic Liquid Colloids for Versatile Use: Biomedical and Environmental Applications. *J. Mol. Liq.* **2014**, *194*, 85–92.
- (550) Mukesh, C.; Bhatt, J.; Prasad, K. A Polymerizable Bioionic Liquid Based Nanogel: A New Nanocarrier for an Anticancer Drug. *Macromol. Chem. Phys.* **2014**, *215*, 1498–1504.
- (551) Seo, J.-W.; Hwang, J.-Y.; Shin, U. S. Ionic Liquid-Doped and p-NIPAAm-Based Copolymer (p-NIBIm): Extraordinary Drug-Entrapping and -Releasing Behaviors at 38–42 °C. *RSC Adv.* **2014**, *4*, 26738–26747.
- (552) Hosseinzadeh, F.; Mahkam, M.; Galehassadi, M. Synthesis and Characterization of Ionic Liquid Functionalized Polymers for Drug Delivery of an Anti-Inflammatory Drug. *Des. Monomers Polym.* **2012**, *15*, 379–388.
- (553) Mahkam, M.; Latifpour, A.; Rafi, A. A.; Gheshlaghi, L. M.; Takfallah, A. Preparation of Montmorillonite-pH-Sensitive Positive Charges Nanocomposites as a Drug Delivery System. *Int. J. Polym. Mater.* **2015**, *64*, 32–37.
- (554) Mahkam, M.; Pakravan, A. Synthesis and Characterization of pH-Sensitive Positive-charge Silica Nanoparticles for Oral Anionic Drug Delivery. *J. Chin. Chem. Soc. (Weinheim, Ger.)* **2013**, *60*, 293–296.
- (555) Rasouli, S.; Davaran, S.; Rasouli, F.; Mahkam, M.; Salehi, R. Positively Charged Functionalized Silica Nanoparticles as Nontoxic Carriers for Triggered Anticancer Drug Release. *Des. Monomers Polym.* **2014**, *17*, 227–237.
- (556) Rasouli, S.; Davaran, S.; Rasouli, F.; Mahkam, M.; Salehi, R. Synthesis, Characterization and pH-Controllable Methotrexate Release from Biocompatible Polymer/Silica Nanocomposite for Anticancer Drug Delivery. *Drug Delivery* **2014**, *21*, 155–163.
- (557) Yang, X.; Chen, Q. Y.; Li, X.; Gao, J. Functional Ionic Liquids Induced the Formation of Mitochondria Targeted Fluorescent Core-Shell Ellipsoidal Nanoparticles with Anticancer Properties. *Colloids Surf., B* **2012**, *98*, 91–96.
- (558) Yang, X.; Chen, Q.-Y.; Kong, M.-Y.; Qu, L.-L.; Geng, Z.-R.; Wang, Z.-L. An Ionic Liquid-Modified Nano-Vehicle to Construct Nano-Models of Catalase to Target Mitochondria. *J. Mater. Chem.* **2012**, *22*, 20299–20304.
- (559) Meng, L.; Niu, L.; Li, L.; Lu, Q.; Fei, Z.; Dyson, P. J. Gold Nanoparticles Grown on Ionic Liquid-Functionalized Single-Walled Carbon Nanotubes: New Materials for Photothermal Therapy. *Chem. - Eur. J.* **2012**, *18*, 13314–13319.
- (560) Weber, C. C.; Kulkarni, S. A.; Kunov-Kruse, A. J.; Rogers, R. D.; Myerson, A. S. The Use of Cooling Crystallization in an Ionic Liquid System for the Purification of Pharmaceuticals. *Cryst. Growth Des.* **2015**, *15*, 4946–4951.
- (561) Weber, C. C.; Kunov-Kruse, A. J.; Rogers, R. D.; Myerson, A. S. Manipulation of Ionic Liquid Anion-Solute-Antisolvent Interactions for the Purification of Acetaminophen. *Chem. Commun.* **2015**, *51*, 4294–4297.
- (562) An, J.-H.; Kim, J.-M.; Chang, S.-M.; Kim, W.-S. Application of Ionic Liquid to Polymorphic Design of Pharmaceutical Ingredients. *Cryst. Growth Des.* **2010**, *10*, 3044–3050.
- (563) An, J.-H.; Jin, F.; Kim, H. S.; Ryu, H. C.; Kim, J. S.; Kim, H. M.; Kim, K. H.; Kiyonga, A. N.; Jung, K. Investigation of the Polymorphic Transformation of the Active Pharmaceutical Ingredient Clopidogrel Bisulfate Using the Ionic Liquid AEImBF₄. *Cryst. Growth Des.* **2016**, *16*, 1829–1836.

- (564) An, J.-H.; Kim, W.-S. Antisolvent Crystallization Using Ionic Liquids As Solvent and Antisolvent for Polymorphic Design of Active Pharmaceutical Ingredient. *Cryst. Growth Des.* **2013**, *13*, 31–39.
- (565) An, J. H.; Jin, F.; Kim, H. S.; Ryu, H. C.; Kim, J. S.; Kim, H. M.; Kiyonga, A. N.; Min, D. S.; Youn, W.; Kim, K. H.; et al. Application of Ionic Liquid to Polymorphic Transformation of Anti-Viral/HIV Drug Adefovir Dipivoxil. *Arch. Pharmacol. Res.* **2016**, *39*, 646–659.
- (566) Viçosa, A.; Letourneau, J.-J.; Espitalier, F.; Inês Ré, M. An Innovative Antisolvent Precipitation Process as a Promising Technique to Prepare Ultrafine Rifampicin Particles. *J. Cryst. Growth* **2012**, *342*, 80–87.
- (567) Petersen, S.; Kaule, S.; Stein, F.; Minrath, I.; Schmitz, K. P.; Kragl, U.; Sternberg, K. Novel Paclitaxel-Coated Angioplasty Balloon Catheter Based on Cetylpyridinium Salicylate: Preparation, Characterization and Simulated Use in an *In Vitro* Vessel Model. *Mater. Sci. Eng., C* **2013**, *33*, 4244–4250.
- (568) Bandomir, J.; Kaule, S.; Schmitz, K.-P.; Sternberg, K.; Petersen, S.; Kragl, U. Usage of Different Vessel Models in a Flow-Through Cell: *In Vitro* Study of a Novel Coated Balloon Catheter. *RSC Adv.* **2015**, *5*, 11604–11610.
- (569) Kaule, S.; Minrath, I.; Stein, F.; Kragl, U.; Schmidt, W.; Schmitz, K. P.; Sternberg, K.; Petersen, S. Correlating Coating Characteristics with the Performance of Drug-Coated Balloons – a Comparative *In Vitro* Investigation of Own Established Hydrogel- and Ionic Liquid-Based Coating Matrices. *PLoS One* **2015**, *10*, e0116080.
- (570) Guo, J.; Xu, Q.; Zheng, Z.; Zhou, S.; Mao, H.; Wang, B.; Yan, F. Intrinsically Antibacterial Poly(Ionic Liquid) Membranes: The Synergistic Effect of Anions. *ACS Macro Lett.* **2015**, *4*, 1094–1098.
- (571) Tang, W.; Liu, B.; Wang, S.; Liu, T.; Fu, C.; Ren, X.; Tan, L.; Duan, W.; Meng, X. Doxorubicin-Loaded Ionic Liquid–Polydopamine Nanoparticles for Combined Chemotherapy and Microwave Thermal Therapy of Cancer. *RSC Adv.* **2016**, *6*, 32434–32440.
- (572) Patel, R.; Parray, M. u. d.; Singh, U. K.; Islam, A.; Venkatesu, P.; Singh, S.; Bohidar, H. B. Effect of 1,4-Bis(3-dodecylimidazolium-1-yl) Butane Bromide on Channel Form of Gramicidin Vesicles. *Colloids Surf., A* **2016**, *508*, 150–158.
- (573) Guncheva, M.; Paunova, K.; Ossowicz, P.; Rozwadowski, Z.; Janus, E.; Idakieva, K.; Todinova, S.; Raynova, Y.; Uzunova, V.; Apostolova, S.; et al. Modification of *Rapana thomasiana* Hemocyanin with Choline Amino Acid Salts Significantly Enhances its Antiproliferative Activity against MCF-7 Human Breast Cancer Cells. *RSC Adv.* **2015**, *5*, 63345–63354.
- (574) Debeljuh, N.; Barrow, C. J.; Byrne, N. The Impact of Ionic Liquids on Amyloid Fibrilization of Abeta16-22: Tuning the Rate of Fibrilization Using a Reverse Hofmeister Strategy. *Phys. Chem. Chem. Phys.* **2011**, *13*, 16534–16536.
- (575) Debeljuh, N.; Barrow, C. J.; Henderson, L.; Byrne, N. Structure Inducing Ionic Liquids-Enhancement of Alpha Helicity in the Abeta(1-40) Peptide from Alzheimer's Disease. *Chem. Commun.* **2011**, *47*, 6371–6373.
- (576) Takekiyo, T.; Yamaguchi, E.; Abe, H.; Yoshimura, Y. Suppression Effect on the Formation of Insulin Amyloid by the Use of Ionic Liquids. *ACS Sustainable Chem. Eng.* **2016**, *4*, 422–428.
- (577) Jaitely, V.; Mizuuchi, H.; Florence, A. T. Current-Stimulated Release of Solutes Solubilized in Water-Immiscible Room Temperature Ionic Liquids (RTILs). *J. Drug Targeting* **2010**, *18*, 787–793.
- (578) Singh, O.; Kaur, R.; Aswal, V. K.; Mahajan, R. K. Composition and Concentration Gradient Induced Structural Transition from Micelles to Vesicles in the Mixed System of Ionic Liquid-Diclofenac Sodium. *Langmuir* **2016**, *32*, 6638–6647.
- (579) Zhang, L.; Liu, J.; Tian, T.; Gao, Y.; Ji, X.; Li, Z.; Luan, Y. Pharmaceutically Active Ionic Liquid Self-Assembled Vesicles for the Application as an Efficient Drug Delivery System. *ChemPhysChem* **2013**, *14*, 3454–3457.
- (580) Cojocar, O. A.; Bica, K.; Gurau, G.; Narita, A.; McCrary, P. D.; Shamshina, J. L.; Barber, P. S.; Rogers, R. D. Prodrug Ionic Liquids: Functionalizing Neutral Active Pharmaceutical Ingredients to Take Advantage of the Ionic Liquid Form. *MedChemComm* **2013**, *4*, 559–563.
- (581) Davis, J. H.; Forrester, K. J.; Merrigan, T. Novel Organic Ionic Liquids (OILs) Incorporating Cations Derived from the Antifungal Drug Miconazole. *Tetrahedron Lett.* **1998**, *39*, 8955–8958.
- (582) Rogers, R. D.; Daly, D. T.; Swatloski, R. P.; Hough, W. L.; Davis, J. H. J.; Smiglak, M.; Pernak, J.; Spear, S. K. Multi-Functional Ionic Liquid Compositions for Overcoming Polymorphism and Imparting Improved Properties for Active Pharmaceutical, Biological, Nutritional, and Energetic Ingredients. US8232265, CA2625004, US8802596, 2012.
- (583) Diogo, H. P.; Ramos, J. J. M. Slow Molecular Dynamics in Three Organic Salts of Active Pharmaceutical Ingredients. *Soft Mater.* **2014**, *12*, 125–137.
- (584) Frizzo, C. P.; Wust, K.; Tier, A. Z.; Beck, T. S.; Rodrigues, L. V.; Vaucher, R. A.; Bolzan, L. P.; Terra, S.; Soares, F.; Martins, M. A. P. Novel Ibuprofenate- and Docusate-Based Ionic Liquids: Emergence of Antimicrobial Activity. *RSC Adv.* **2016**, *6*, 100476–100486.
- (585) Araújo, J. M. M.; Florindo, C.; Pereiro, A. B.; Vieira, N. S. M.; Matias, A. A.; Duarte, C. M. M.; Rebelo, L. P. N.; Marrucho, I. M. Cholinium-Based Ionic Liquids with Pharmaceutically Active Anions. *RSC Adv.* **2014**, *4*, 28126–28132.
- (586) Balk, A.; Wiest, J.; Widmer, T.; Galli, B.; Holzgrabe, U.; Meinel, L. Transformation of Acidic Poorly Water Soluble Drugs into Ionic Liquids. *Eur. J. Pharm. Biopharm.* **2015**, *94*, 73–82.
- (587) Bica, K.; Rijkssen, C.; Nieuwenhuyzen, M.; Rogers, R. D. In Search of Pure Liquid Salt Forms of Aspirin: Ionic Liquid Approaches with Acetylsalicylic Acid and Salicylic Acid. *Phys. Chem. Chem. Phys.* **2010**, *12*, 2011–2017.
- (588) Bica, K.; Rogers, R. D. Confused Ionic Liquid Ions—a “Liquification” and Dosage Strategy for Pharmaceutically Active Salts. *Chem. Commun.* **2010**, *46*, 1215–1217.
- (589) Ferraz, R.; Branco, L. C.; Marrucho, I. M.; Araújo, J. M. M.; Rebelo, L. P. N.; da Ponte, M. N.; Prudêncio, C.; Noronha, J. P.; Petrovski, Ž. Development of Novel Ionic Liquids Based on Ampicillin. *MedChemComm* **2012**, *3*, 494–497.
- (590) Postleb, F.; Stefanik, D.; Seifert, H.; Giernoth, R. BIOionic Liquids: Imidazolium-based Ionic Liquids with Antimicrobial Activity. *Z. Naturforsch., B: J. Chem. Sci.* **2013**, *68*, 1123–1128.
- (591) Sahbaz, Y.; Williams, H. D.; Nguyen, T. H.; Saunders, J.; Ford, L.; Charman, S. A.; Scammells, P. J.; Porter, C. J. Transformation of Poorly Water-Soluble Drugs into Lipophilic Ionic Liquids Enhances Oral Drug Exposure from Lipid Based Formulations. *Mol. Pharmaceutics* **2015**, *12*, 1980–1991.
- (592) Song, Z.; Yu, Q.; Cai, M.; Huang, G.; Yao, M.; Li, D.; Liang, Y.; Fan, M.; Zhou, F. Green Ionic Liquid Lubricants Prepared from Anti-Inflammatory Drug. *Tribol. Lett.* **2015**, *60*, 38.
- (593) Tourne-Peteilh, C.; Devoisselle, J. M.; Vioux, A.; Judeinstein, P. In, M.; Viau, L. Surfactant Properties of Ionic Liquids Containing Short Alkyl Chain Imidazolium Cations and Ibuprofenate Anions. *Phys. Chem. Chem. Phys.* **2011**, *13*, 15523–15529.
- (594) Tourne-Peteilh, C.; Coasne, B.; In, M.; Brevet, D.; Devoisselle, J. M.; Vioux, A.; Viau, L. Surfactant Behavior of Ionic Liquids Involving a Drug: from Molecular Interactions to Self-Assembly. *Langmuir* **2014**, *30*, 1229–1238.
- (595) Sastry, N. V.; Singh, D. K. Surfactant and Gelation Properties of Acetylsalicylate Based Room Temperature Ionic Liquid in Aqueous Media. *Langmuir* **2016**, *32*, 10000–10016.
- (596) Shadid, M.; Gurau, G.; Shamshina, J. L.; Chuang, B. C.; Hailu, S.; Guan, E.; Chowdhury, S. K.; Wu, J. T.; Rizvi, S. A. A.; Griffin, R. J.; et al. Sulfasalazine in Ionic Liquid Form with Improved Solubility and Exposure. *MedChemComm* **2015**, *6*, 1837–1841.
- (597) Balk, A.; Widmer, T.; Wiest, J.; Bruhn, H.; Rybak, J. C.; Matthes, P.; Muller-Buschbaum, K.; Sakalis, A.; Luhmann, T.; Berghausen, J.; et al. Ionic Liquid versus Prodrug Strategy to Address Formulation Challenges. *Pharm. Res.* **2015**, *32*, 2154–2167.
- (598) Viciosa, M. T.; Santos, G.; Costa, A.; Danede, F.; Branco, L. C.; Jordao, N.; Correia, N. T.; Dionisio, M. Dipolar Motions and Ionic Conduction in an Ibuprofen Derived Ionic Liquid. *Phys. Chem. Chem. Phys.* **2015**, *17*, 24108–24120.

- (599) Zhao, H.; Holmes, S. S.; Baker, G. A.; Challa, S.; Bose, H. S.; Song, Z. Ionic Derivatives of Betulinic Acid as Novel HIV-1 Protease Inhibitors. *J. Enzyme Inhib. Med. Chem.* **2012**, *27*, 715–721.
- (600) Miwa, Y.; Hamamoto, H.; Ishida, T. Lidocaine Self-Sacrificially Improves the Skin Permeation of the Acidic and Poorly Water-Soluble Drug Etodolac via Its Transformation into an Ionic Liquid. *Eur. J. Pharm. Biopharm.* **2016**, *102*, 92–100.
- (601) Qamar, S.; Brown, P.; Ferguson, S.; Khan, R. A.; Ismail, B.; Khan, A. R.; Sayed, M.; Khan, A. M. The Interaction of a Model Active Pharmaceutical with Cationic Surfactant and the Subsequent Design of Drug Based Ionic Liquid Surfactants. *J. Colloid Interface Sci.* **2016**, *481*, 117–124.
- (602) Keramatnia, F.; Jouyban, A.; Valizadeh, H.; Delazar, A.; Shayanfar, A. Ketoconazole Ionic Liquids with Citric and Tartaric Acid: Synthesis, Characterization and Solubility Study. *Fluid Phase Equilib.* **2016**, *425*, 108–113.
- (603) Alves, F.; Oliveira, F. S.; Schroder, B.; Matos, C.; Marrucho, I. M. Synthesis, Characterization, and Liposome Partition of a Novel Tetracycline Derivative Using the Ionic Liquids Framework. *J. Pharm. Sci.* **2013**, *102*, 1504–1512.
- (604) Florindo, C.; Araujo, J. M.; Alves, F.; Matos, C.; Ferraz, R.; Prudencio, C.; Noronha, J. P.; Petrovski, Z.; Branco, L.; Rebelo, L. P.; et al. Evaluation of Solubility and Partition Properties of Ampicillin-Based Ionic Liquids. *Int. J. Pharm.* **2013**, *456*, 553–559.
- (605) Bica, K.; Rodriguez, H.; Gurau, G.; Cojocar, O. A.; Riisager, A.; Fehrmann, R.; Rogers, R. D. Pharmaceutically Active Ionic Liquids with Solids Handling, Enhanced Thermal Stability, and Fast Release. *Chem. Commun.* **2012**, *48*, 5422–5424.
- (606) Jouannin, C.; Tourné-Péteilh, C.; Darcos, V.; Sharkawi, T.; Devoisselle, J.-M.; Gaveau, P.; Dieudonné, P.; Vioux, A.; Viau, L. Drug Delivery Systems Based on Pharmaceutically Active Ionic Liquids and Biocompatible Poly(Lactic Acid). *J. Mater. Chem. B* **2014**, *2*, 3133–3141.
- (607) Choi, S. Y.; Rodríguez, H.; Gunaratne, H. Q. N.; Puga, A. V.; Gilpin, D.; McGrath, S.; Vyle, J. S.; Tunney, M. M.; Rogers, R. D.; McNally, T. Dual Functional Ionic Liquids as Antimicrobials and Plasticisers for Medical Grade PVCs. *RSC Adv.* **2014**, *4*, 8567–8581.
- (608) Stoimenovski, J.; MacFarlane, D. R. Enhanced Membrane Transport of Pharmaceutically Active Protic Ionic Liquids. *Chem. Commun.* **2011**, *47*, 11429–11431.
- (609) Stoimenovski, J.; Dean, P. M.; Izgorodina, E. I.; MacFarlane, D. R. Protic Pharmaceutical Ionic Liquids and Solids: Aspects of Protonics. *Faraday Discuss.* **2012**, *154*, 335–352.
- (610) Wang, H.; Gurau, G.; Shamshina, J.; Cojocar, O. A.; Janikowski, J.; MacFarlane, D. R.; Davis, J. H.; Rogers, R. D. Simultaneous Membrane Transport of Two Active Pharmaceutical Ingredients by Charge Assisted Hydrogen Bond Complex Formation. *Chem. Sci.* **2014**, *5*, 3449–3456.
- (611) Bielas, R.; Mielńczyk, A.; Siewniak, A.; Neugebauer, D. Trimethylammonium-Based Polymethacrylate Ionic Liquids with Tunable Hydrophilicity and Charge Distribution as Carriers of Salicylate Anions. *ACS Sustainable Chem. Eng.* **2016**, *4*, 4181–4191.
- (612) Cherukuvada, S.; Nangia, A. Polymorphism in an API Ionic Liquid: Ethambutol Dibenzoate Trimorphs. *CrystEngComm* **2012**, *14*, 7840–7843.
- (613) Cojocar, O. A.; Kelley, S. P.; Gurau, G.; Rogers, R. D. Procainium Acetate Versus Procainium Acetate Dihydrate: Irreversible Crystallization of a Room-Temperature Active Pharmaceutical Ingredient Ionic Liquid upon Hydration. *Cryst. Growth Des.* **2013**, *13*, 3290–3293.
- (614) Cybulski, J.; Wiśniewska, A.; Kulig-Adamiak, A.; Lewicka, L.; Cieniecka-Rosłonkiewicz, A.; Kita, K.; Fojutowski, A.; Nawrot, J.; Materna, K.; Pernak, J. Long-Alkyl-Chain Quaternary Ammonium Lactate Based Ionic Liquids. *Chem. - Eur. J.* **2008**, *14*, 9305–9311.
- (615) Cybulski, J.; Wiśniewska, A.; Kulig-Adamiak, A.; Dąbrowski, Z.; Praczyk, T.; Michalczyk, A.; Walkiewicz, F.; Materna, K.; Pernak, J. Mandelate and Prolinate Ionic Liquids: Synthesis, Characterization, Catalytic and Biological Activity. *Tetrahedron Lett.* **2011**, *52*, 1325–1328.
- (616) Kondratenko, Y.; Kochina, T.; Fundamensky, V.; Ignatyev, I.; Panikorovskii, T.; Nyanikova, G. Triethanolammonium Salicylate – Protic Alkanolammonium Ionic Liquid. *J. Mol. Liq.* **2016**, *221*, 1218–1224.
- (617) Markiewicz, B.; Sznajdrowska, A.; Chrzanowski, Ł.; Ławniczak, Ł.; Zgoła-Grzeškowiak, A.; Kubiak, K.; Nawrot, J.; Pernak, J. Ionic Liquids with a Theophyllinate Anion. *New J. Chem.* **2014**, *38*, 3146–3153.
- (618) Pinto, P. C. A. G.; Ribeiro, D. M. G. P.; Azevedo, A. M. O.; Dela Justina, V.; Cunha, E.; Bica, K.; Vasiloiu, M.; Reis, S.; Saraiva, M. L. M. F. S. Active Pharmaceutical Ingredients Based on Salicylate Ionic Liquids: Insights into the Evaluation of Pharmaceutical Profiles. *New J. Chem.* **2013**, *37*, 4095–4102.
- (619) Park, H. J.; Prausnitz, M. R. Lidocaine-Ibuprofen Ionic Liquid for Dermal Anesthesia. *AIChE J.* **2015**, *61*, 2732–2738.
- (620) Lovejoy, K. S.; Corley, C. A.; Cope, E. K.; Valentine, M. C.; Leid, J. G.; Purdy, G. M.; Wilkes, J. S.; Koppisch, A. T.; Del Sesto, R. E. Utilization of Metal Halide Species Ambiguity to Develop Amorphous, Stabilized Pharmaceutical Agents As Ionic Liquids. *Cryst. Growth Des.* **2012**, *12*, 5357–5364.
- (621) Magut, P. K.; Das, S.; Fernand, V. E.; Losso, J.; McDonough, K.; Naylor, B. M.; Aggarwal, S.; Warner, I. M. Tunable Cytotoxicity of Rhodamine 6G via Anion Variations. *J. Am. Chem. Soc.* **2013**, *135*, 15873–15879.
- (622) Vraneš, M.; Tot, A.; Jovanović-Šanta, S.; Karaman, M.; Dožić, S.; Tešanović, K.; Kojić, V.; Gadžurić, S. Toxicity Reduction of Imidazolium-Based Ionic Liquids by the Oxygenation of the Alkyl Substituent. *RSC Adv.* **2016**, *6*, 96289–96295.
- (623) Mester, P.; Jehle, A. K.; Leeb, C.; Kalb, R.; Grunert, T.; Rossmann, P. FTIR Metabolomic Fingerprint Reveals Different Modes of Action Exerted by Active Pharmaceutical Ingredient Based Ionic Liquids (API-ILs) on *Salmonella typhimurium*. *RSC Adv.* **2016**, *6*, 32220–32227.
- (624) Ferraz, R.; Teixeira, V.; Rodrigues, D.; Fernandes, R.; Prudêncio, C.; Noronha, J. P.; Petrovski, Z.; Branco, L. C. Antibacterial Activity of Ionic Liquids Based on Ampicillin against Resistant Bacteria. *RSC Adv.* **2014**, *4*, 4301–4307.
- (625) Ferraz, R.; Costa-Rodrigues, J.; Fernandes, M. H.; Santos, M. M.; Marrucho, I. M.; Rebelo, L. P.; Prudencio, C.; Noronha, J. P.; Petrovski, Z.; Branco, L. C. Antitumor Activity of Ionic Liquids Based on Ampicillin. *ChemMedChem* **2015**, *10*, 1480–1483.
- (626) Challa, S.; Zhao, H.; Gumbs, A.; Chetty, C. S.; Bose, H. S. New Ionic Derivatives of Betulinic Acid as Highly Potent Anti-Cancer Agents. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 1734–1738.
- (627) Borkowski, A.; Ławniczak, Ł.; Clapa, T.; Narożna, D.; Selwet, M.; Peziak, D.; Markiewicz, B.; Chrzanowski, Ł. Different Antibacterial Activity of Novel Theophylline-Based Ionic Liquids - Growth Kinetic and Cytotoxicity Studies. *Ecotoxicol. Environ. Saf.* **2016**, *130*, 54–64.
- (628) Silveira, L. T.; Liberatore, A. M. A.; Koh, I. H. J.; Bizeto, M. A.; Camilo, F. F. Combined Bactericidal Activity of Silver Nanoparticles and Hexadecylpyridinium Salicylate Ionic Liquid. *J. Nanopart. Res.* **2015**, *17*, 129.
- (629) Smith, D. J.; Shah, J. K.; Maginn, E. J. Molecular Dynamics Simulation Study of the Association of Lidocainium Docusate and its Derivatives in Aqueous Solution. *Mol. Pharmaceutics* **2015**, *12*, 1893–1901.
- (630) Yamaguchi, T.; Kawai, K.; Yamanaka, K.; Tatsumi, N. External Preparation Composition Comprising Fatty Acid-Based Ionic Liquid as Active Ingredient. US8623387, 2014.
- (631) Furukawa, S.-y.; Hattori, G.; Sakai, S.; Kamiya, N. Highly Efficient and Low Toxic Skin Penetrants Composed of Amino Acid Ionic Liquids. *RSC Adv.* **2016**, *6*, 87753–87755.
- (632) Reinhardt, A.; Horn, M.; Schmauck, J. P.; Brohl, A.; Giernoth, R.; Oelkrug, C.; Schubert, A.; Neundorff, I. Novel Imidazolium Salt-Peptide Conjugates and Their Antimicrobial Activity. *Bioconjugate Chem.* **2014**, *25*, 2166–2174.
- (633) Costa, S. P.; Justina, V. D.; Bica, K.; Vasiloiu, M.; Pinto, P. C.; Saraiva, M. L. Automated Evaluation of Pharmaceutically Active Ionic

Liquids' (Eco)toxicity through the Inhibition of Human Carboxylesterase and *Vibrio fischeri*. *J. Hazard. Mater.* **2014**, *265*, 133–141.

(634) Moreira, D. N.; Fresno, N.; Pérez-Fernández, R.; Frizzo, C. P.; Goya, P.; Marco, C.; Martins, M. A. P.; Elguero, J. Brønsted Acid–Base Pairs of Drugs as Dual Ionic Liquids: NMR Ionicity Studies. *Tetrahedron* **2015**, *71*, 676–685.

(635) Dean, P. M.; Turanjanin, J.; Yoshizawa-Fujita, M.; MacFarlane, D. R.; Scott, J. L. Exploring an Anti-Crystal Engineering Approach to the Preparation of Pharmaceutically Active Ionic Liquids. *Cryst. Growth Des.* **2009**, *9*, 1137–1145.

(636) Hamamoto, H.; Miwa, Y. Tape Preparation Comprising Etodolac in Ionic Liquid Form. US8697096, EP2233138, 2014.

(637) Miwa, Y.; Hamamoto, H.; Hikake, S.; Kuwabara, Y. A Phase I, Randomized, Open-Label, Cross-Over Study of the Pharmacokinetics, Dermal Tolerability, and Safety of MRX-7EAT Etodolac-Lidocaine Topical Patch in Healthy Volunteers. *J. Pain* **2013**, *14*, S72.

(638) Kuwabara, Y.; Hamamoto, H.; Hikake, S.; Miwa, Y. A Randomized, Multi-Center, Double-Blind, Placebo-Controlled Phase II/III Trial to Evaluate the Efficacy, Tolerability and Safety of MRX-7EAT Etodolac-Lidocaine Topical Patch in the Treatment of Pain. *J. Pain* **2013**, *14*, S73.

(639) ClinicalTrials.gov, <https://clinicaltrials.gov>, accessed Nov 2016.

(640) Ribeiro, R.; Pinto, P. C.; Azevedo, A. M.; Bica, K.; Ressmann, A. K.; Reis, S.; Saraiva, M. L. Automated Evaluation of Protein Binding Affinity of Anti-Inflammatory Choline Based Ionic Liquids. *Talanta* **2016**, *150*, 20–26.

(641) Tanzi, L.; Nardone, M.; Benassi, P.; Ramondo, F.; Caminiti, R.; Gontrani, L. Choline Salicylate Ionic Liquid by X-ray Scattering, Vibrational Spectroscopy and Molecular Dynamics. *J. Mol. Liq.* **2016**, *218*, 39–49.

(642) Carquigny, S.; Lakard, B.; Lakard, S.; Moutarlier, V.; Hihn, J.-Y.; Viau, L. Investigation of Pharmaceutically Active Ionic Liquids as Electrolyte for the Electrosynthesis of Polypyrrole and Active Component in Controlled Drug Delivery. *Electrochim. Acta* **2016**, *211*, 950–961.

(643) Saadeh, S. M.; Yasseen, Z.; Sharif, F. A.; Abu Shawish, H. M. New Room Temperature Ionic Liquids with Interesting Ecotoxicological and Antimicrobial Properties. *Ecotoxicol. Environ. Saf.* **2009**, *72*, 1805–1809.

(644) Armaković, S.; Armaković, S. J.; Vraneš, M.; Tot, A.; Gadžurić, S. DFT Study of 1-Butyl-3-Methylimidazolium Salicylate: a Third-Generation Ionic Liquid. *J. Mol. Model.* **2015**, *21*, 246.

(645) Shekaari, H.; Zafarani-Moattar, M. T.; Mirheydari, S. N. Effect of 1-Butyl-3-methylimidazolium Ibuprofenate as an Active Pharmaceutical Ingredient Ionic Liquid (API-IL) on the Thermodynamic Properties of Glycine and L-Alanine in Aqueous Solutions at Different Temperatures. *J. Solution Chem.* **2016**, *45*, 624–663.

(646) Vraneš, M.; Armaković, S.; Tot, A.; Papović, S.; Zec, N.; Armaković, S.; Banić, N.; Abramović, B.; Gadžurić, S. Structuring of Water in the New Generation Ionic Liquid – Comparative Experimental and Theoretical Study. *J. Chem. Thermodyn.* **2016**, *93*, 164–171.

(647) Shekaari, H.; Zafarani-Moattar, M. T.; Mirheydari, S. N. Thermodynamic Properties of 1-Butyl-3-methylimidazolium Salicylate as an Active Pharmaceutical Ingredient Ionic Liquid (API-IL) in Aqueous Solutions of Glycine and L-Alanine in at T=(288.15 to 318.15) K. *Thermochim. Acta* **2016**, *637*, 51–68.

(648) Banić, N.; Abramović, B.; Šibul, F.; Orčić, D.; Watson, M.; Vraneš, M.; Gadžurić, S. Advanced Oxidation Processes for the Removal of [bmim][Sal] Third Generation Ionic Liquids: Effect of Water Matrices and Intermediates Identification. *RSC Adv.* **2016**, *6*, 52826–52837.

(649) Shekaari, H.; Zafarani-Moattar, M. T.; Mirheydari, S. N. Conductometric Analysis of 1-Butyl-3-methylimidazolium Ibuprofenate as an Active Pharmaceutical Ingredient Ionic Liquid (API-IL) in the Aqueous Amino Acids Solutions. *J. Chem. Thermodyn.* **2016**, *103*, 165–175.

(650) Brevet, D.; Jouannin, C.; Tourné-Péteilh, C.; Devoisselle, J.-M.; Vioux, A.; Viau, L. Self-Encapsulation of a Drug-Containing Ionic Liquid

into Mesoporous Silica Monoliths or Nanoparticles by a Sol–Gel Process. *RSC Adv.* **2016**, *6*, 82916–82923.

(651) Gaitor, J. C.; Zayas, M. S.; Myrthil, D. J.; White, F.; Hendrich, J. M.; Sykora, R. E.; O'Brien, R. A.; Reilly, J. T.; Mirjafari, A. Crystal Structure of a Methimazole-Based Ionic Liquid. *Acta Crystallogr., Sect. E: Crystallogr. Commun.* **2015**, *71*, o1008–o1009.

(652) Vasanthakumar, A.; Bahadur, I.; Redhi, G.; Gengan, R. M. Synthesis and Characterization of 2',3'-Epoxy Propyl-N-methyl-2-oxopyrrolidinium Salicylate Ionic Liquid and Study of its Interaction with Water or Methanol. *RSC Adv.* **2016**, *6*, 61566–61575.

(653) Swiety-Pospiech, A.; Wojnarowska, Z.; Pionteck, J.; Pawlus, S.; Grzybowski, A.; Hensel-Bielowka, S.; Grzybowski, K.; Szulc, A.; Paluch, M. High Pressure Study of Molecular Dynamics of Protic Ionic Liquid Lidocaine Hydrochloride. *J. Chem. Phys.* **2012**, *136*, 224501.

(654) Wojnarowska, Z.; Grzybowski, K.; Hawelek, L.; Swiety-Pospiech, A.; Masiewicz, E.; Paluch, M.; Sawicki, W.; Chmielewska, A.; Bujak, P.; Markowski, J. Molecular Dynamics Studies on the Water Mixtures of Pharmaceutically Important Ionic Liquid Lidocaine HCl. *Mol. Pharmaceutics* **2012**, *9*, 1250–1261.

(655) Ishibashi, M.; Hamamoto, H. Lidocaine Tape Preparation. US8722065, EP2210599B, 2014.

(656) Wojnarowska, Z.; Roland, C. M.; Kolodziejczyk, K.; Swiety-Pospiech, A.; Grzybowski, K.; Paluch, M. Quantifying the Structural Dynamics of Pharmaceuticals in the Glassy State. *J. Phys. Chem. Lett.* **2012**, *3*, 1238–1241.

(657) Wojnarowska, Z.; Roland, C. M.; Swiety-Pospiech, A.; Grzybowski, K.; Paluch, M. Anomalous Electrical Conductivity Behavior at Elevated Pressure in the Protic Ionic Liquid Procainamide Hydrochloride. *Phys. Rev. Lett.* **2012**, *108*, 015701.

(658) Cherukuvada, S.; Nangia, A. Salts and Ionic Liquid of The Antituberculosis Drug S,S-Ethambutol. *Cryst. Growth Des.* **2013**, *13*, 1752–1760.

(659) Wojnarowska, Z.; Paluch, M.; Grzybowski, A.; Adrjanowicz, K.; Grzybowski, K.; Kaminski, K.; Włodarczyk, P.; Pionteck, J. Study of Molecular Dynamics of Pharmaceutically Important Protic Ionic Liquid-Verapamil Hydrochloride. I. Test of Thermodynamic Scaling. *J. Chem. Phys.* **2009**, *131*, 104505.

(660) Wojnarowska, Z.; Grzybowski, K.; Grzybowski, A.; Paluch, M.; Kaminski, K.; Włodarczyk, P.; Adrjanowicz, K.; Pionteck, J. Study of Molecular Dynamics of the Pharmaceutically Important Protic Ionic Liquid Verapamil Hydrochloride. II. Test of Entropic Models. *J. Chem. Phys.* **2010**, *132*, 094506.

(661) Martins, M. A.; Frizzo, C. P.; Moreira, D. N.; Zanatta, N.; Bonacorso, H. G. Ionic Liquids in Heterocyclic Synthesis. *Chem. Rev.* **2008**, *108*, 2015–2050.

(662) Trombini, C.; Lombardo, M.; Quintavalla, A.; Chiarucci, M. Multiphase Homogeneous Catalysis: Common Procedures and Recent Applications. *Synlett* **2010**, *12*, 1746–1765.

(663) Li, Y.; He, Y.-M.; Fan, Q.-H. Ionic Liquids in Transition Metal-Catalyzed Enantioselective Reactions. In *Ionic Liquids (ILs) in Organometallic Catalysis*; Dupont, J., Kollár, L., Eds.; Springer-Verlag: Berlin-Heidelberg, 2013; Vol. 51, pp 323–347.

(664) Monteiro, A. L.; Zinn, F. K.; de Souza, R. F.; Dupont, J. Asymmetric Hydrogenation of 2-Arylacrylic Acids Catalyzed by Immobilized Ru-BINAP Complex in 1-*n*-Butyl-3-methylimidazolium Tetrafluoroborate Molten Salt. *Tetrahedron: Asymmetry* **1997**, *8*, 177–179.

(665) Earle, M. J.; Seddon, K. R.; McCormac, P. B. The First High Yield Green Route to a Pharmaceutical in a Room Temperature Ionic Liquid. *Green Chem.* **2000**, *2*, 261–262.

(666) Hongwei, Y.; Jinchuan, W.; Chi Bun, C. Kinetic Resolution of Ibuprofen Catalyzed by *Candida rugosa* Lipase in Ionic Liquids. *Chirality* **2005**, *17*, 16–21.

(667) Contesini, F. J.; de Oliveira Carvalho, P. Esterification of (RS)-Ibuprofen by Native and Commercial Lipases in a Two-Phase System Containing Ionic Liquids. *Tetrahedron: Asymmetry* **2006**, *17*, 2069–2073.

(668) Deshmukh, A.; Gore, B.; Thulasiram, H. V.; Swamy, V. P. Recyclable Ionic Liquid Iodinating Reagent for Solvent Free,

Regioselective Iodination of Activated Aromatic and Heteroaromatic Amines. *RSC Adv.* **2015**, *5*, 88311–88315.

(669) Liu, B. K.; Wang, N.; Chen, Z. C.; Wu, Q.; Lin, X. F. Markedly Enhancing Lipase-Catalyzed Synthesis of Nucleoside Drugs' Ester by Using a Mixture System Containing Organic Solvents and Ionic Liquid. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 3769–3771.

(670) Kumar, V.; Malhotra, S. V. Synthesis of Nucleoside-Based Antiviral Drugs in Ionic Liquids. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5640–5642.

(671) Kumar, V.; Malhotra, S. V. Ionic Liquid Mediated Synthesis of 5-Halouracil Nucleosides: Key Precursors for Potential Antiviral Drugs. *Nucleosides, Nucleotides Nucleic Acids* **2009**, *28*, 821–834.

(672) Naik, P. U.; Harjani, J. R.; Nara, S. J.; Salunkhe, M. M. Ionic Liquid Enabled Sulfamoylation of Arenes: an Ambient, Expeditious and Regioselective Protocol for Aryl Sulfonamides. *Tetrahedron Lett.* **2004**, *45*, 1933–1936.

(673) Le, Z.-G.; Xu, J.-P.; Rao, H.-Y.; Ying, M. One-Pot Synthesis of 2-Aminobenzothiazoles Using a New Reagent of [bmim]₃Br₃ in [bmim]BF₄. *J. Heterocycl. Chem.* **2006**, *43*, 1123–1124.

(674) James, B.; Viji, S.; Mathew, S.; Nair, M. S.; Lakshmanan, D.; Ajay Kumar, R. Synthesis of Novel Highly Functionalized Biologically Active Polycyclic Caged Amides. *Tetrahedron Lett.* **2007**, *48*, 6204–6208.

(675) Orrling, K. M.; Wu, X.; Russo, F.; Larhed, M. Fast, Acid-Free, and Selective Lactamization of Lactones in Ionic Liquids. *J. Org. Chem.* **2008**, *73*, 8627–8630.

(676) Heravi, M. An Efficient Synthesis of Quinolines Derivatives Promoted by a Room Temperature Ionic Liquid at Ambient Conditions under Ultrasound Irradiation via the Tandem Addition/Annulation Reaction of *o*-Aminoaryl Ketones with α -Methylene Ketones. *Ultrason. Sonochem.* **2009**, *16*, 361–366.

(677) Lellouche, J.; Friedman, A.; Lellouche, J.-P.; Gedanken, A.; Banin, E. Improved antibacterial and antibiofilm activity of magnesium fluoride nanoparticles obtained by water-based ultrasound chemistry. *Nanomedicine* **2012**, *8*, 702–711.

(678) Dandia, A.; Jain, A. K. Ionic Liquid-Mediated Facile Synthesis of Novel Spiroheterobicyclic Rings as Potential Antifungal and Antibacterial Drugs. *J. Heterocycl. Chem.* **2013**, *50*, 104–113.

(679) Parmar, N. J.; Patel, R. A.; Parmar, B. D.; Talpada, N. P. An Efficient Domino Reaction in Ionic Liquid: Synthesis and Biological Evaluation of Some Pyrano- and Thiopyrano-Fused Heterocycles. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 1656–1661.

(680) Patel, R. V.; Chikhaliya, K. H.; Nile, S. H.; Park, S. W. Ionic Liquid Mediated Tandem Synthesis of Bioactive Quinoline Based Thiophene/Thiazole Linked Multi-Heterocomponent Ugi Adducts. *Curr. Org. Chem.* **2013**, *17*, 1125–1129.

(681) Satasia, S. P.; Kalaria, P. N.; Raval, D. K. Catalytic Regioselective Synthesis of Pyrazole Based Pyrido[2,3-*d*]pyrimidine-diones and Their Biological Evaluation. *Org. Biomol. Chem.* **2014**, *12*, 1751–1758.

(682) Anusha, S.; Cp, B.; Mohan, C. D.; Mathai, J.; Rangappa, S.; Mohan, S.; Chandra; Paricharak, S.; Mervin, L.; Fuchs, J. E.; et al. A Nano-MgO and Ionic Liquid-Catalyzed 'Green' Synthesis Protocol for the Development of Adamantyl-Imidazolo-Thiadiazoles as Anti-Tuberculosis Agents Targeting Sterol 14 α -Demethylase (CYP51). *PLoS One* **2015**, *10*, e0139798.

(683) Thanh, N. D.; Kim Van, H. T.; Thu, T. T. Synthesis and Characterization of Some Novel Thiosemicarbazones of Substituted Benzaldehydes and *N*-(Hepta-*O*-Acetyl- β -*D*-Lactosyl)-Thiosemicarbazide. *J. Carbohydr. Chem.* **2015**, *34*, 514–544.

(684) Ramalho, H. F.; Ferreira, K. M. C.; Machado, P. M. A.; Silva, T. B.; Rangel, E. T.; Prauchner, M. J.; Suarez, P. A. Z. Production of Additives with Antimicrobial Activity via Tandem Hydroformylation-amine Condensation of Soybean FAME Using an Ionic Liquid-Based Biphasic Catalytic System. *J. Braz. Chem. Soc.* **2016**, *27*, 321–333.

(685) Subhedar, D. D.; Shaikh, M. H.; Nawale, L.; Yeware, A.; Sarkar, D.; Khan, F. A. K.; Sangshetti, J. N.; Shingate, B. B. Novel Tetrazoloquinoline-Rhodanine Conjugates: Highly Efficient Synthesis and Biological Evaluation. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 2278–2283.

(686) Subhedar, D. D.; Shaikh, M. H.; Kalam Khan, F. A.; Sangshetti, J. N.; Khedkar, V. M.; Shingate, B. B. Facile Synthesis of New *N*-Sulfonamidyl-4-thiazolidinone Derivatives and Their Biological Evaluation. *New J. Chem.* **2016**, *40*, 3047–3058.

(687) Subramanian, G.; Babu Rajeev, C. P.; Mohan, C. D.; Sinha, A.; Chu, T. T.; Anusha, S.; Ximei, H.; Fuchs, J. E.; Bender, A.; Rangappa, K. S.; et al. Synthesis and In Vitro Evaluation of Hydrazinyl Phthalazines against Malaria Parasite, *Plasmodium falciparum*. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 3300–3306.

(688) Bhatt, J. D.; Chudasama, C. J.; Patel, K. D. Microwave Assisted Synthesis of Pyrimidines in Ionic Liquid and Their Potency as Non-Classical Malarial Antifolates. *Arch. Pharm.* **2016**, *349*, 791–800.

(689) Harjani, J. R.; Nara, S. J.; Salunkhe, M. M. Lewis Acidic Ionic Liquids for the Synthesis of Electrophilic Alkenes via the Knoevenagel Condensation. *Tetrahedron Lett.* **2002**, *43*, 1127–1130.

(690) Darvatkar, N. B.; Deorukhkar, A. R.; Bhilare, S. V.; Raut, D. G.; Salunkhe, M. M. Ionic Liquid-Mediated Synthesis of Coumarin-3-carboxylic Acids via Knoevenagel Condensation of Meldrum's Acid with ortho-Hydroxyaryl Aldehydes. *Synth. Commun.* **2008**, *38*, 3508–3513.

(691) Valizadeh, H.; Vaghefi, S. One-Pot Wittig and Knoevenagel Reactions in Ionic Liquid as Convenient Methods for the Synthesis of Coumarin Derivatives. *Synth. Commun.* **2009**, *39*, 1666–1678.

(692) Valizadeh, H.; Gholipour, H. Imidazolium-Based Phosphinite Ionic Liquid (IL-OPPh₂) as Reusable Catalyst and Solvent for the Knoevenagel Condensation Reaction. *Synth. Commun.* **2010**, *40*, 1477–1485.

(693) Valizadeh, H.; Mahmoodian, M.; Gholipour, H. ZrCl₄/[bmim]BF₄-catalyzed condensation of salicylaldehydes and malononitrile: Single-step synthesis of 3-cyanocoumarin derivatives. *J. Heterocycl. Chem.* **2011**, *48*, 799–802.

(694) Kurata, A.; Kitamura, Y.; Irie, S.; Takemoto, S.; Akai, Y.; Hirota, Y.; Fujita, T.; Iwai, K.; Furusawa, M.; Kishimoto, N. Enzymatic Synthesis of Caffeic Acid Phenethyl Ester Analogues in Ionic Liquid. *J. Biotechnol.* **2010**, *148*, 133–138.

(695) Heravi, M.; Ansari, P.; Saeedi, M.; Tavakoli-Hosseini, N.; Karimi, N. Green and Practical Synthesis of Benzopyran and 3-Substituted Coumarin Derivatives by Brønsted Acid Ionic Liquid [(CH₂)₄SO₃HMIM][HSO₄]. *Bull. Chem. Soc. Ethiop.* **2011**, *25*, 315–320.

(696) Rao, V. K.; Chhikara, B. S.; Tiwari, R.; Shirazi, A. N.; Parang, K.; Kumar, A. One-Pot Regioselective Synthesis of Tetrahydroindazolones and Evaluation of Their Antiproliferative and Src Kinase Inhibitory Activities. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 410–414.

(697) Chen, Z.; Bi, J.; Su, W. Synthesis and Antitumor Activity of Novel Coumarin Derivatives via a Three-component Reaction in Water. *Chin. J. Chem.* **2013**, *31*, 507–514.

(698) Rao, V. K.; Tiwari, R.; Chhikara, B. S.; Shirazi, A. N.; Parang, K.; Kumar, A. Copper Triflate-Mediated Synthesis of 1,3,5-Triarylpyrazoles in [bmim][PF₆] Ionic Liquid and Evaluation of Their Anticancer Activities. *RSC Adv.* **2013**, *3*, 15396–15403.

(699) Shallu; Sharma, M. L.; Singh, J. First Total Synthesis of a Guanidine Alkaloid Nitensidine D Using Immobilized Ionic Liquid, Microwaves and Formamidinesulfonic Acid. *J. Chem. Sci. (Bangalore, India)* **2014**, *126*, 1869–1874.

(700) Dicarboxylic Acid Derivatives with Anti-Tumor Effect and Preparation Method Thereof. CN102531964, 2014.

(701) Antitumor Dicarboxylic Acid Di-(Hydroxamic Acid) Ester and Preparation Method Thereof. CN102516124, 2015.

(702) Basiri, A.; Murugaiyah, V.; Osman, H.; Kumar, R. S.; Kia, Y.; Awang, K. B.; Ali, M. A. An Expedient, Ionic Liquid Mediated Multi-Component Synthesis of Novel Piperidone Grafted Cholinesterase Enzymes Inhibitors and Their Molecular Modeling Study. *Eur. J. Med. Chem.* **2013**, *67*, 221–229.

(703) Basiri, A.; Murugaiyah, V.; Osman, H.; Kumar, R. S.; Kia, Y.; Hooda, A.; Parsons, R. B. Cholinesterase Inhibitory Activity versus Aromatic Core Multiplicity: a Facile Green Synthesis and Molecular Docking Study of Novel Piperidone Embedded Thiazolopyrimidines. *Bioorg. Med. Chem.* **2014**, *22*, 906–916.

- (704) Abd Razik, B. M.; Osman, H.; Basiri, A.; Salhin, A.; Kia, Y.; Ezzat, M. O.; Murugaiyah, V. Ionic Liquid Mediated Synthesis and Molecular Docking Study of Novel Aromatic Embedded Schiff Bases as Potent Cholinesterase Inhibitors. *Bioorg. Chem.* **2014**, *57*, 162–168.
- (705) Ghaffari Khaligh, N. 1,1'-Butylenebis(3-methyl-3H-imidazol-1-ium) Hydrogen Sulfate as an Efficient Binuclear Bronsted Ionic Liquid for the Synthesis of Tacrine Analogues. *Monatsh. Chem.* **2015**, *146*, 321–326.
- (706) Rajerison, H.; Faye, D.; Roumesy, A.; Louaisil, N.; Boeda, F.; Faivre-Chauvet, A.; Gestin, J. F.; Legoupy, S. Ionic Liquid Supported Organotin Reagents to Prepare Molecular Imaging and Therapy Agents. *Org. Biomol. Chem.* **2016**, *14*, 2121–2126.
- (707) Yeung, K.-S.; Farkas, M. E.; Qiu, Z.; Yang, Z. Friedel-Crafts Acylation of Indoles in Acidic Imidazolium Chloroaluminate Ionic Liquid at Room Temperature. *Tetrahedron Lett.* **2002**, *43*, 5793–5795.
- (708) Zang, H.; Su, Q.; Mo, Y.; Cheng, B.-W.; Jun, S. Ionic Liquid [EMIM]OAc under Ultrasonic Irradiation towards the First Synthesis of Trisubstituted Imidazoles. *Ultrason. Sonochem.* **2010**, *17*, 749–751.
- (709) Zang, H.; Su, Q.; Mo, Y.; Cheng, B. Ionic Liquid under Ultrasonic Irradiation towards a Facile Synthesis of Pyrazolone Derivatives. *Ultrason. Sonochem.* **2011**, *18*, 68–72.
- (710) Zavozin, A. G.; Ignat'ev, N. V.; Schulte, M.; Zlotin, S. G. Synthesis of Thiazole Derivatives Bearing an Incorporated Z-5-Aminopent-3-enoic Acid Fragment. *Tetrahedron* **2013**, *69*, 6975–6980.
- (711) Siddiqui, I. R.; Srivastava, A.; Shamim, S.; Srivastava, A.; Shireen; Waseem, M. A. An Efficient One-Pot Regioselective Approach Towards the Synthesis of Thiopyrano[2,3-d]thiazole-2-thiones Catalyzed by Basic Ionic Liquid under Microwave Irradiation. *J. Heterocycl. Chem.* **2016**, *53*, 849–858.
- (712) Zhang, X.; Li, X.; Li, D.; Qu, G.; Wang, J.; Loiseau, P. M.; Fan, X. Ionic Liquid Mediated and Promoted Eco-Friendly Preparation of Thiazolidinone and Pyrimidine Nucleoside-Thiazolidinone Hybrids and Their Antiparasitic Activities. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 6280–6283.
- (713) Suresh; Sandhu, J. S. Ultrasound-Assisted Synthesis of 2,4-Thiazolidinedione and Rhodanine Derivatives Catalyzed by Task-Specific Ionic Liquid: [TMG][Lac]. *Org. Med. Chem. Lett.* **2013**, *3*, 2.
- (714) Method for Improving Reaction Rate of Enzymatic Ibuprofen Chiral Separation. CN101225427, 2011.
- (715) Wei, T.; Yang, K.; Bai, B.; Zang, J.; Yu, X.; Mao, D. Enzymatic Hydrolytic Resolution of Racemic Ibuprofen Ethyl Ester Using an Ionic Liquid as Cosolvent. *Molecules* **2016**, *21*, 905.
- (716) Gao, F.; Wang, L.; Liu, Y.; Wang, S.; Jiang, Y.; Hu, M.; Li, S.; Zhai, Q. Enzymatic Synthesis of (R)-Modafinil by Chloroperoxidase-Catalyzed Enantioselective Sulfoxidation of 2-(Diphenylmethylthio)Acetamide. *Biochem. Eng. J.* **2015**, *93*, 243–249.
- (717) Method for Synthesis Tioconazole. CN101519403, 2012.
- (718) Hydrocortisone Production Process Based on Ionic Liquid. CN101805386, 2012.
- (719) Synthesis Method for Ciclesonide. CN101857627, 2012.
- (720) Tao, Y.; Dong, R.; Pavlidis, I. V.; Chen, B.; Tan, T. Using Imidazolium-Based Ionic Liquids as Dual Solvent-Catalysts for Sustainable Synthesis of Vitamin Esters: Inspiration from Bio- and Organo-Catalysis. *Green Chem.* **2016**, *18*, 1240–1248.
- (721) Ternois, J.; Guillen, F.; Plaquevent, J.-C.; Coquerel, G. Asymmetric Synthesis of Modafinil and Its Derivatives by Enantioselective Oxidation of Thioethers: Comparison of Various Methods Including Synthesis in Ionic Liquids. *Tetrahedron: Asymmetry* **2007**, *18*, 2959–2964.
- (722) Chakraborty, B.; Sharma, C. D. A New Route to the Synthesis of Isoxazoline Derivatives from Dihydropyran via Cycloaddition Reaction in Ionic Liquid. *Tetrahedron Lett.* **2013**, *54*, 5532–5536.
- (723) Chen, Y.; Zu, Y.; Fu, Y.; Zhang, X.; Yu, P.; Sun, G.; Efferth, T. Efficient Lewis Acid Ionic Liquid-Catalyzed Synthesis of the Key Intermediate of Coenzyme Q10 under Microwave Irradiation. *Molecules* **2010**, *15*, 9486–9495.
- (724) Choi, H. J.; Uhm, K.-N.; Kim, H.-K. Production of Chiral Compound Using Recombinant *Escherichia coli* Cells Co-expressing Reductase and Glucose Dehydrogenase in an Ionic Liquid/Water Two Phase System. *J. Mol. Catal. B: Enzym.* **2011**, *70*, 114–118.
- (725) Kandar, S.; Suresh, A. K.; Noronha, S. B. (R)-PAC Biosynthesis in [BMIM][PF₆]/Aqueous Biphasic System Using *Saccharomyces cerevisiae* BY4741 Cells. *Appl. Biochem. Biotechnol.* **2015**, *175*, 1771–1788.
- (726) Kazemi, S.; Martín, Á. B.; Sordi, D.; Kroon, M. C.; Peters, C. J.; Arends, I. W. C. E. Vanadium-Catalyzed Epoxidation Reaction of Cinnamyl Alcohol in Ionic Liquids. *Green Process. Synth.* **2012**, *1*, 509–516.
- (727) Singh, M.; Singh, R. S.; Banerjee, U. C. Stereoselective Synthesis of (R)-1-Chloro-3-(3,4-difluorophenoxy)-2-propanol Using Lipases from *Pseudomonas aeruginosa* in Ionic Liquid-Containing System. *J. Mol. Catal. B: Enzym.* **2009**, *56*, 294–299.
- (728) Subhedar, D. D.; Shaikh, M. H.; Nawale, L.; Yeware, A.; Sarkar, D.; Shingate, B. B. [Et₃NH][HSO₄] Catalyzed Efficient Synthesis of 5-Arylidene-Rhodanine Conjugates and Their Antitubercular Activity. *Res. Chem. Intermed.* **2016**, *42*, 6607–6626.
- (729) Wu, D.-X.; Guan, Y.-X.; Wang, H.-Q.; Yao, S.-J. 11 α -Hydroxylation of 16 α ,17-Epoxyprogesterone by *Rhizopus nigricans* in a Biphasic Ionic Liquid Aqueous System. *Bioresour. Technol.* **2011**, *102*, 9368–9373.
- (730) Zhang, B.-B.; Cheng, J.; Lou, W.-Y.; Wang, P.; Zong, M.-H. Efficient Anti-Prelog Enantioselective Reduction of Acetyltrimethylsilane to (R)-1-Trimethylsilylethanol by Immobilized *Candida parapsilosis* CCTCC M203011 Cells in Ionic Liquid-Based Biphasic Systems. *Microb. Cell Fact.* **2012**, *11*, 108.
- (731) Kretzschmar, G.; Kraft, V.; Rossen, K.; Gräser, J. Process for the Preparation of Dronedarone Intermediates. EP2435413, 2014.
- (732) Lohse, O.; Penn, G.; Schilling, H. Process for the Preparation of 5-(Halocetyl)-8-(substituted oxy)-(1h)-quinolin-2-ones. US7605267, 2009.
- (733) Dang, D. T.; Ha, S. H.; Lee, S.-M.; Chang, W.-J.; Koo, Y.-M. Enhanced Activity and Stability of Ionic Liquid-Pretreated Lipase. *J. Mol. Catal. B: Enzym.* **2007**, *45*, 118–121.
- (734) Tietze, A. A.; Bordusa, F.; Giernoth, R.; Imhof, D.; Lenzer, T.; Maaß, A.; Mrestani-Klaus, C.; Neundorf, I.; Oum, K.; Reith, D.; et al. On the Nature of Interactions between Ionic Liquids and Small Amino-Acid-Based Biomolecules. *ChemPhysChem* **2013**, *14*, 4044–4064.
- (735) Attri, P.; Venkatesu, P. Thermodynamic Characterization of the Biocompatible Ionic Liquid Effects on Protein Model Compounds and Their Functional Groups. *Phys. Chem. Chem. Phys.* **2011**, *13*, 6566–6575.
- (736) Attri, P.; Venkatesu, P.; Kumar, A.; Byrne, N. A Protic Ionic Liquid Attenuates the Deleterious Actions of Urea on Alpha-Chymotrypsin. *Phys. Chem. Chem. Phys.* **2011**, *13*, 17023–17026.
- (737) Attri, P.; Venkatesu, P.; Kumar, A. Water and a Protic Ionic Liquid Acted as Refolding Additives for Chemically Denatured Enzymes. *Org. Biomol. Chem.* **2012**, *10*, 7475–7478.
- (738) Page, T. A.; Kraut, N. D.; Page, P. M.; Baker, G. A.; Bright, F. V. Dynamics of Loop 1 of Domain I in Human Serum Albumin When Dissolved in Ionic Liquids. *J. Phys. Chem. B* **2009**, *113*, 12825–12830.
- (739) Papoian, G. A.; Ulander, J.; Eastwood, M. P.; Luthey-Schulten, Z.; Wolynes, P. G. Water in Protein Structure Prediction. *Proc. Natl. Acad. Sci. U. S. A.* **2004**, *101*, 3352–3357.
- (740) Chaplin, M. Do We Underestimate the Importance of Water in Cell Biology? *Nat. Rev. Mol. Cell Biol.* **2006**, *7*, 861–866.
- (741) Ball, P. Water as an Active Constituent in Cell Biology. *Chem. Rev.* **2008**, *108*, 74–108.
- (742) Bellissent-Funel, M. C.; Hassanali, A.; Havenith, M.; Henchman, R.; Pohl, P.; Sterpone, F.; van der Spoel, D.; Xu, Y.; Garcia, A. E. Water Determines the Structure and Dynamics of Proteins. *Chem. Rev.* **2016**, *116*, 7673–7697.
- (743) Feng, G.; Jiang, X.; Qiao, R.; Kornyshev, A. A. Water in Ionic Liquids at Electrified Interfaces: the Anatomy of Electrosorption. *ACS Nano* **2014**, *8*, 11685–11694.
- (744) Motobayashi, K.; Osawa, M. Potential-Dependent Condensation of Water at the Interface between Ionic Liquid [BMIM][TfSA] and an Au Electrode. *Electrochem. Commun.* **2016**, *65*, 14–17.

- (745) Zhong, Y.; Yan, J.; Li, M.; Chen, L.; Mao, B. The Electric Double Layer in an Ionic Liquid Incorporated with Water Molecules: Atomic Force Microscopy Force Curve Study. *ChemElectroChem* **2016**, *3*, 2221–2226.
- (746) Cui, T.; Lahiri, A.; Carstens, T.; Borisenko, N.; Pulletikurthi, G.; Kuhl, C.; Endres, F. Influence of Water on the Electrified Ionic Liquid/Solid Interface: A Direct Observation of the Transition from a Multilayered Structure to a Double-Layer Structure. *J. Phys. Chem. C* **2016**, *120*, 9341–9349.
- (747) Cheng, H.-W.; Stock, P.; Moeremans, B.; Baimpos, T.; Banquy, X.; Renner, F. U.; Valtiner, M. Characterizing the Influence of Water on Charging and Layering at Electrified Ionic-Liquid/Solid Interfaces. *Adv. Mater. Interfaces* **2015**, *2*, 1500159.
- (748) Abe, H.; Nakama, K.; Hayashi, R.; Aono, M.; Takekiyo, T.; Yoshimura, Y.; Saihara, K.; Shimizu, A. Electrochemical Anomalies of Protic Ionic Liquid – Water Systems: A Case Study Using Ethylammonium Nitrate – Water System. *Chem. Phys.* **2016**, *475*, 119–125.
- (749) Latif, M. A.; Tejo, B. A.; Abedikargiban, R.; Abdul Rahman, M. B.; Micaelo, N. M. Modeling Stability and Flexibility of alpha-Chymotrypsin in Room Temperature Ionic Liquids. *J. Biomol. Struct. Dyn.* **2014**, *32*, 1263–1273.
- (750) Latif, M. A. M.; Micaelo, N. M.; Abdul Rahman, M. B. Influence of Anion–Water Interactions on the Behaviour of Lipases in Room Temperature Ionic Liquids. *RSC Adv.* **2014**, *4*, 48202–48211.
- (751) Jaganathan, M.; Ramakrishnan, C.; Velmurugan, D.; Dhathathreyan, A. Understanding Ethylammonium Nitrate Stabilized Cytochrome c – Molecular Dynamics and Experimental Approach. *J. Mol. Struct.* **2015**, *1081*, 334–341.
- (752) Jha, I.; Attri, P.; Venkatesu, P. Unexpected Effects of the Alteration of Structure and Stability of Myoglobin and Hemoglobin in Ammonium-Based Ionic Liquids. *Phys. Chem. Chem. Phys.* **2014**, *16*, 5514–5526.
- (753) Lesch, V.; Heuer, A.; Tatsis, V. A.; Holm, C.; Smiatek, J. Peptides in the Presence of Aqueous Ionic Liquids: Tunable Co-Solutes as Denaturants or Protectants? *Phys. Chem. Chem. Phys.* **2015**, *17*, 26049–26053.
- (754) Jaeger, V. W.; Pfaendtner, J. Structure, Dynamics, and Activity of Xylanase Solvated in Binary Mixtures of Ionic Liquid and Water. *ACS Chem. Biol.* **2013**, *8*, 1179–1186.
- (755) Micaelo, N. M.; Soares, C. M. Protein Structure and Dynamics in Ionic Liquids. Insights from Molecular Dynamics Simulation Studies. *J. Phys. Chem. B* **2008**, *112*, 2566–2572.
- (756) Ghosh, S.; Parui, S.; Jana, B.; Bhattacharyya, K. Ionic Liquid Induced Dehydration and Domain Closure in Lysozyme: FCS and MD Simulation. *J. Chem. Phys.* **2015**, *143*, 125103.
- (757) Rawat, K.; Bohidar, H. B. Universal Charge Quenching and Stability of Proteins in 1-Methyl-3-alkyl (Hexyl/Octyl) Imidazolium Chloride Ionic Liquid Solutions. *J. Phys. Chem. B* **2012**, *116*, 11065–11074.
- (758) Rawat, K.; Bohidar, H. B. Heparin-Like Native Protein Aggregate Dissociation by 1-Alkyl-3-methyl Imidazolium Chloride Ionic Liquids. *Int. J. Biol. Macromol.* **2015**, *73*, 23–30.
- (759) Johnson, L. B.; Park, S.; Gintner, L. P.; Snow, C. D. Characterization of Supercharged Cellulase Activity and Stability in Ionic Liquids. *J. Mol. Catal. B: Enzym.* **2016**, *132*, 84–90.
- (760) Lesch, V.; Heuer, A.; Holm, C.; Smiatek, J. Solvent Effects of 1-Ethyl-3-methylimidazolium Acetate: Solvation and Dynamic Behavior of Polar and Apolar Solutes. *Phys. Chem. Chem. Phys.* **2015**, *17*, 8480–8490.
- (761) Brogan, A. P.; Hallett, J. P. Solubilizing and Stabilizing Proteins in Anhydrous Ionic Liquids through Formation of Protein-Polymer Surfactant Nanoconstructs. *J. Am. Chem. Soc.* **2016**, *138*, 4494–4501.
- (762) Ou, G.; He, B.; Halling, P. Ionization Basis for Activation of Enzymes Soluble in Ionic Liquids. *Biochim. Biophys. Acta, Gen. Subj.* **2016**, *1860*, 1404–1408.
- (763) Klähn, M.; Lim, G. S.; Seduraman, A.; Wu, P. On the Different Roles of Anions and Cations in the Solvation of Enzymes in Ionic Liquids. *Phys. Chem. Chem. Phys.* **2011**, *13*, 1649–1662.
- (764) Klähn, M.; Lim, G. S.; Wu, P. How Ion Properties Determine the Stability of a Lipase Enzyme in Ionic Liquids: a Molecular Dynamics Study. *Phys. Chem. Chem. Phys.* **2011**, *13*, 18647–18660.
- (765) Shu, Y.; Liu, M.; Chen, S.; Chen, X.; Wang, J. New Insight into Molecular Interactions of Imidazolium Ionic Liquids with Bovine Serum Albumin. *J. Phys. Chem. B* **2011**, *115*, 12306–12314.
- (766) Yan, H.; Wu, J.; Dai, G.; Zhong, A.; Chen, H.; Yang, J.; Han, D. Interaction Mechanisms of Ionic Liquids [C_nmim]Br (n=4, 6, 8, 10) with Bovine Serum Albumin. *J. Lumin.* **2012**, *132*, 622–628.
- (767) Burney, P. R.; Pfaendtner, J. Structural and Dynamic Features of *Candida rugosa* Lipase 1 in Water, Octane, Toluene, and Ionic Liquids BMIM-PF₆ and BMIM-NO₃. *J. Phys. Chem. B* **2013**, *117*, 2662–2670.
- (768) Burney, P. R.; Nordwald, E. M.; Hickman, K.; Kaar, J. L.; Pfaendtner, J. Molecular Dynamics Investigation of the Ionic Liquid/Enzyme Interface: Application to Engineering Enzyme Surface Charge. *Proteins: Struct., Funct., Genet.* **2015**, *83*, 670–680.
- (769) Baker, J. L.; Furbish, J.; Lindberg, G. E. Influence of the Ionic Liquid [C₄mpy][Tf₂N] on the Structure of the Miniprotein Trp-Cage. *J. Mol. Graphics Modell.* **2015**, *62*, 202–212.
- (770) Chevrot, G.; Fileti, E. E.; Chaban, V. V. Enhanced Stability of the Model Mini-Protein in Amino Acid Ionic Liquids and Their Aqueous Solutions. *J. Comput. Chem.* **2015**, *36*, 2044–2051.
- (771) Yu, Y.; Wang, J.; Shao, Q.; Shi, J.; Zhu, W. The Effects of Organic Solvents on the Folding Pathway and Associated Thermodynamics of Proteins: A Microscopic View. *Sci. Rep.* **2016**, *6*, 19500.
- (772) Sandoval, M.; Cortés, Á.; Civera, C.; Treviño, J.; Ferreras, E.; Vaultier, M.; Berenguer, J.; Lozano, P.; Hernández, M. J. Efficient and Selective Enzymatic Synthesis of N-Acetyl-lactosamine in Ionic Liquid: a Rational Explanation. *RSC Adv.* **2012**, *2*, 6306–6314.
- (773) Brakowski, R.; Pontius, K.; Franzreb, M. Investigation of the Transglycosylation Potential of β-Galactosidase from *Aspergillus oryzae* in the Presence of the Ionic Liquid [Bmim][PF₆]. *J. Mol. Catal. B: Enzym.* **2016**, *130*, 48–57.
- (774) Chen, F.; Liu, S. S.; Yu, M.; Qu, R.; Wang, M. C. Blocking the Entrance of AMP Pocket Results in Hormetic Stimulation of Imidazolium-Based Ionic Liquids to Firefly Luciferase. *Chemosphere* **2015**, *132*, 108–113.
- (775) Nordwald, E. M.; Armstrong, G. S.; Kaar, J. L. NMR-Guided Rational Engineering of an Ionic-Liquid-Tolerant Lipase. *ACS Catal.* **2014**, *4*, 4057–4064.
- (776) Li, W.; Wang, L.; Zhou, R.; Mu, Y. Ionic Liquid Induced Inactivation of Cellobiohydrolase I from *Trichoderma reesei*. *Green Chem.* **2015**, *17*, 1618–1625.
- (777) Ha, S. H.; Van Anh, T.; Koo, Y. M. Optimization of Lipase-Catalyzed Synthesis of Caffeic Acid Phenethyl Ester in Ionic Liquids by Response Surface Methodology. *Bioprocess Biosyst. Eng.* **2013**, *36*, 799–807.
- (778) Pfruender, H.; Amidjojo, M.; Kragl, U.; Weuster-Botz, D. Efficient Whole-Cell Biotransformation in a Biphasic Ionic Liquid/Water System. *Angew. Chem., Int. Ed.* **2004**, *43*, 4529–4531.
- (779) Weuster-Botz, D. Process Intensification of Whole-Cell Biocatalysis with Ionic Liquids. *Chem. Rec.* **2007**, *7*, 334–340.
- (780) Wood, N.; Ferguson, J. L.; Gunaratne, H. Q. N.; Seddon, K. R.; Goodacre, R.; Stephens, G. M. Screening Ionic Liquids for Use in Biotransformations with Whole Microbial Cells. *Green Chem.* **2011**, *13*, 1843–1851.
- (781) Fileti, E. E.; Chaban, V. V. Imidazolium Ionic Liquid Helps to Disperse Fullerenes in Water. *J. Phys. Chem. Lett.* **2014**, *5*, 1795–1800.
- (782) Saha, A.; Payra, S.; Verma, S. K.; Mandal, M.; Thareja, S.; Banerjee, S. *In Silico* Binding Affinity to Cyclooxygenase-II and Green Synthesis of Benzylpyrazolyl Coumarin Derivatives. *RSC Adv.* **2015**, *5*, 100978–100983.
- (783) Soni, S. K.; Sarkar, S.; Selvakannan, P. R.; Sarkar, D.; Bhargava, S. K. Intrinsic Therapeutic and Biocatalytic Roles of Ionic Liquid Mediated Self-Assembled Platinum-Phytase Nanospheres. *RSC Adv.* **2015**, *5*, 62871–62881.
- (784) Dorj, B.; Won, J. E.; Purevdorj, O.; Patel, K. D.; Kim, J. H.; Lee, E. J.; Kim, H. W. A Novel Therapeutic Design of Microporous

Structured Biopolymer Scaffolds for Drug Loading and Delivery. *Acta Biomater.* **2014**, *10*, 1238–1250.

(785) Kim, S. H.; Seo, J. W.; Shin, U. S. Application of Room-Temperature Ionic Liquids in Preparation of Highly Porous Polymer Membranes and Microspheres. *Bull. Korean Chem. Soc.* **2015**, *36*, 643–649.

(786) Shin, U.-S.; Kim, J.-G. 3D Micromorphology Producing within Poly(lactic acid) Skeleton Using Room-Temperature Ionic Liquids: From Particulate, Fibrous or Porous Scaffolds to Beads. *Bull. Korean Chem. Soc.* **2012**, *33*, 2295–2298.

(787) Kim, S. Y.; Hwang, J. Y.; Seo, J. W.; Shin, U. S. Production of CNT-Taxol-Embedded PCL Microspheres Using an Ammonium-Based Room Temperature Ionic Liquid: As a Sustained Drug Delivery System. *J. Colloid Interface Sci.* **2015**, *442*, 147–153.

(788) Kim, S. Y.; Hwang, J. Y.; Shin, U. S. Preparation of Nano/Macroporous Polycaprolactone Microspheres for an Injectable Cell Delivery System Using Room Temperature Ionic Liquid and Camphene. *J. Colloid Interface Sci.* **2016**, *465*, 18–25.

(789) Yuan, Y.-Y.; Du, J.-Z.; Song, W.-J.; Wang, F.; Yang, X.-Z.; Xiong, M.-H.; Wang, J. Biocompatible and Functionalizable Polyphosphate Nanogel with a Branched Structure. *J. Mater. Chem.* **2012**, *22*, 9322–9329.

(790) Swatloski, R. P.; Rogers, R. D.; Holbrey, J. D. Dissolution and Processing of Cellulose Using Ionic Liquids. US6824599, CA2462460, EP1458805, EP2325246, 2004.

(791) Daly, D. T.; Spear, S. K.; Turner, M. B.; Hough, W. L.; Rogers, R. D. Cellulosic Biocomposites as Molecular Scaffolds for Nano-Architectures. US8883193, 2014.

(792) Rogers, R. D.; Daly, D. T.; Turner, M. B.; Spear, S. K.; Holbrey, J. D. Ionic Liquid Reconstituted Cellulose Composites as Solid Support Matrices. EP1907470, 2013.

(793) Kim, M. H.; An, S.; Won, K.; Kim, H. J.; Lee, S. H. Entrapment of Enzymes into Cellulose–Biopolymer Composite Hydrogel Beads Using Biocompatible Ionic Liquid. *J. Mol. Catal. B: Enzym.* **2012**, *75*, 68–72.

(794) Guo, Y.; Wang, X.; Shu, X.; Shen, Z.; Sun, R. C. Self-Assembly and Paclitaxel Loading Capacity of Cellulose-Graft-Poly(Lactide) Nanomicelles. *J. Agric. Food Chem.* **2012**, *60*, 3900–3908.

(795) Li, L.; Meng, L.; Zhang, X.; Fu, C.; Lu, Q. The Ionic Liquid-Associated Synthesis of a Cellulose/SWCNT Complex and Its Remarkable Biocompatibility. *J. Mater. Chem.* **2009**, *19*, 3612–3617.

(796) Tsiopstias, C.; Panayiotou, C. Preparation of Cellulose-Nanohydroxyapatite Composite Scaffolds from Ionic Liquid Solutions. *Carbohydr. Polym.* **2008**, *74*, 99–105.

(797) George, A.; Brandt, A.; Tran, K.; Zahari, S. M. S. N. S.; Klein-Marcuschamer, D.; Sun, N.; Sathitsuksanoh, N.; Shi, J.; Stavila, V.; Parthasarathi, R.; et al. Design of Low-Cost Ionic Liquids for Lignocellulosic Biomass Pretreatment. *Green Chem.* **2015**, *17*, 1728–1734.

(798) Mai, N. L.; Koo, Y. M. Computer-Aided Design of Ionic Liquids for High Cellulose Dissolution. *ACS Sustainable Chem. Eng.* **2016**, *4*, 541–547.

(799) Walther, P.; Ota, A.; Müller, A.; Hermanutz, F.; Gähr, F.; Buchmeiser, M. R. Chitin Foils and Coatings Prepared from Ionic Liquids. *Macromol. Mater. Eng.* **2016**, *301*, 1337–1344.

(800) Tran, C. D.; Mututuvuri, T. M. Cellulose, Chitosan, and Keratin Composite Materials. Controlled Drug Release. *Langmuir* **2015**, *31*, 1516–1526.

(801) Hua, D.; Jiang, J.; Kuang, L.; Jiang, J.; Zheng, W.; Liang, H. Smart Chitosan-Based Stimuli-Responsive Nanocarriers for the Controlled Delivery of Hydrophobic Pharmaceuticals. *Macromolecules* **2011**, *44*, 1298–1302.

(802) Liu, Y.; Huang, Y.; Boamah, P.-O.; Cao, L.; Zhang, Q.; Lu, Z.; Li, H. Homogeneous Synthesis of Linoleic Acid-Grafted Chitosan Oligosaccharide in Ionic Liquid and Its Self-Assembly Performance in Aqueous Solution. *J. Appl. Polym. Sci.* **2015**, *132*, 41727.

(803) Elshaarawy, R. F.; Refaee, A. A.; El-Sawi, E. A. Pharmacological Performance of Novel Poly-(Ionic Liquid)-Grafted Chitosan-N-salicylidene Schiff Bases and Their Complexes. *Carbohydr. Polym.* **2016**, *146*, 376–387.

(804) Liu, H.; Sale, K. L.; Holmes, B. M.; Simmons, B. A.; Singh, S. Understanding the Interactions of Cellulose with Ionic Liquids: a Molecular Dynamics Study. *J. Phys. Chem. B* **2010**, *114*, 4293–4301.

(805) Pang, J.; Luan, Y.; Li, F.; Cai, X.; Li, Z. Ionic Liquid-Assisted Synthesis of Silica Particles and Their Application in Drug Release. *Mater. Lett.* **2010**, *64*, 2509–2512.

(806) Zhou, G.; Luo, Z.; Fu, X. Preparation of Starch Nanoparticles in a Water-in-Ionic Liquid Microemulsion System and Their Drug Loading and Releasing Properties. *J. Agric. Food Chem.* **2014**, *62*, 8214–8220.

(807) Wang, X.; Chen, H.; Luo, Z.; Fu, X. Preparation of Starch Nanoparticles in Water in Oil Microemulsion System and Their Drug Delivery Properties. *Carbohydr. Polym.* **2016**, *138*, 192–200.

(808) Wang, X.; Cheng, J.; Ji, G.; Peng, X.; Luo, Z. Starch Nanoparticles Prepared in a Two Ionic Liquid Based Microemulsion System and Their Drug Loading and Release Properties. *RSC Adv.* **2016**, *6*, 4751–4757.

(809) Ji, G.; Luo, Z.; Xiao, Z.; Peng, X. Synthesis of Starch Nanoparticles in a Novel Microemulsion with Two ILs Substituting Two Phases. *J. Mater. Sci.* **2016**, *51*, 7085–7092.

(810) Casal-Dujat, L.; Rodrigues, M.; Yagüe, A.; Calpena, A. C.; Amabilin, D. B.; González-Linares, J.; Borràs, M.; Pérez-García, L. Gemini Iimidazolium Amphiphiles for the Synthesis, Stabilization, and Drug Delivery from Gold Nanoparticles. *Langmuir* **2012**, *28*, 2368–2381.

(811) Serrà, A.; Gimeno, N.; Gómez, E.; Mora, M.; Sagristá, M. L.; Vallés, E. Magnetic Mesoporous Nanocarriers for Drug Delivery with Improved Therapeutic Efficacy. *Adv. Funct. Mater.* **2016**, *26*, 6601–6611.

(812) Dorjnamjin, D.; Ariunaa, M.; Shim, Y. K. Synthesis of Silver Nanoparticles Using Hydroxyl Functionalized Ionic Liquids and Their Antimicrobial Activity. *Int. J. Mol. Sci.* **2008**, *9*, 807–820.

(813) Srivastava, C. M.; Purwar, R. Fabrication of Robust *Antheraea assama* Fibroin Nanofibrous Mat Using Ionic Liquid for Skin Tissue Engineering. *Mater. Sci. Eng., C* **2016**, *68*, 276–290.

(814) Dai, L.; Shen, Y.; Li, D.; Xiao, S.; He, J. Cellulose-Graft-Poly(L-Lactide) as a Degradable Drug-Delivery System: Synthesis, Degradation and Drug Release. *Cell. Chem. Technol.* **2014**, *48*, 237–245.

(815) Qin, W.; Li, S. F. An Ionic Liquid Coating for Determination of Sildenafil and UK-103,320 in Human Serum by Capillary Zone Electrophoresis-Ion Trap Mass Spectrometry. *Electrophoresis* **2002**, *23*, 4110–4116.

(816) Chen, J.; Wang, Y.; Zeng, Q.; Ding, X.; Huang, Y. Partition of Proteins with Extraction in Aqueous Two-Phase System by Hydroxyl Ammonium-Based Ionic Liquid. *Anal. Methods* **2014**, *6*, 4067–4076.

(817) Taha, M.; Almeida, M. R.; Silva, F. A.; Domingues, P.; Ventura, S. P.; Coutinho, J. A.; Freire, M. G. Novel Biocompatible and Self-Buffering Ionic Liquids for Biopharmaceutical Applications. *Chem. - Eur. J.* **2015**, *21*, 4781–4788.

(818) Shi, X.; Qiao, L.; Xu, G. Recent Development of Ionic Liquid Stationary Phases for Liquid Chromatography. *J. Chromatogr. A* **2015**, *1420*, 1–15.

(819) Tietze, A. A.; Heimer, P.; Stark, A.; Imhof, D. Ionic Liquid Applications in Peptide Chemistry: Synthesis, Purification and Analytical Characterization Processes. *Molecules* **2012**, *17*, 4158–4185.

(820) Foston, M.; Samuel, R.; He, J.; Ragauskas, A. J. A Review of Whole Cell Wall NMR by the Direct-Dissolution of Biomass. *Green Chem.* **2016**, *18*, 608–621.

(821) Tholey, A.; Heinzle, E. Ionic (Liquid) Matrices for Matrix-Assisted Laser Desorption/Ionization Mass Spectrometry-Applications and Perspectives. *Anal. Bioanal. Chem.* **2006**, *386*, 24–37.

(822) Li, Y. L.; Gross, M. L. Ionic-Liquid Matrices for Quantitative Analysis by MALDI-TOF Mass Spectrometry. *J. Am. Soc. Mass Spectrom.* **2004**, *15*, 1833–1837.

(823) Opallo, M.; Lesniewski, A. A Review on Electrodes Modified with Ionic Liquids. *J. Electroanal. Chem.* **2011**, *656*, 2–16.

(824) Behera, K.; Pandey, S.; Kadyan, A. Ionic Liquid-Based Optical and Electrochemical Carbon Dioxide Sensors. *Sensors* **2015**, *15*, 30487–30503.

- (825) Abo-Hamad, A.; AlSaadi, M. A.; Hayyan, M.; Juneidi, I.; Hashim, M. A. Ionic Liquid-Carbon Nanomaterial Hybrids for Electrochemical Sensor Applications: a Review. *Electrochim. Acta* **2016**, *193*, 321–343.
- (826) Fernández-Navarro, J. J.; García-Álvarez-Coque, M. C.; Ruiz-Ángel, M. J. The Role of the Dual Nature of Ionic Liquids in the Reversed-Phase Liquid Chromatographic Separation of Basic Drugs. *J. Chromatogr. A* **2011**, *1218*, 398–407.
- (827) Fernández-Navarro, J. J.; Torres-Lapasíó, J. R.; Ruiz-Ángel, M. J.; García-Álvarez-Coque, M. C. 1-Hexyl-3-methyl Imidazolium Tetrafluoroborate: an Efficient Column Enhancer for the Separation of Basic Drugs by Reversed-Phase Liquid Chromatography. *J. Chromatogr. A* **2012**, *1258*, 168–174.
- (828) Mavroudi, M. C.; Kapnissi-Christodoulou, C. P. Combined Use of L-Alanine Tert Butyl Ester Lactate and Trimethyl-beta-cyclodextrin for the Enantiomeric Separations of 2-Arylpropionic Acids Nonsteroidal Anti-Inflammatory Drugs. *Electrophoresis* **2015**, *36*, 2442–2450.
- (829) Tran, C. D.; Mejac, I. Chiral Ionic Liquids for the Separation of Pharmaceutical Products by Capillary Electrophoresis. *J. Chromatogr. A* **2008**, *1204*, 204–209.
- (830) Zhang, Q.; Du, Y.; Du, S.; Zhang, J.; Feng, Z.; Zhang, Y.; Li, X. Tetramethylammonium-Lactobionate: A Novel Ionic Liquid Chiral Selector Based on Saccharides in Capillary Electrophoresis. *Electrophoresis* **2015**, *36*, 1216–1223.
- (831) Yu, J.; Zuo, L.; Liu, H.; Zhang, L.; Guo, X. Synthesis and Application of a Chiral Ionic Liquid Functionalized Beta-Cyclodextrin as a Chiral Selector in Capillary Electrophoresis. *Biomed. Chromatogr.* **2013**, *27*, 1027–1033.
- (832) Tran, C. D.; Oliveira, D. Fluorescence Determination of Enantiomeric Composition of Pharmaceuticals via Use of Ionic Liquid that Serves as Both Solvent and Chiral Selector. *Anal. Biochem.* **2006**, *356*, 51–58.
- (833) Breadmore, M. C. Ionic Liquid-Based Liquid Phase Microextraction with Direct Injection for Capillary Electrophoresis. *J. Chromatogr. A* **2011**, *1218*, 1347–1352.
- (834) Bianchi, F.; Dugheri, S.; Musci, M.; Bonacchi, A.; Salvadori, E.; Arcangeli, G.; Cupelli, V.; Lanciotti, M.; Masieri, L.; Serni, S.; et al. Fully Automated Solid-Phase Microextraction-Fast Gas Chromatography-Mass Spectrometry Method Using a New Ionic Liquid Column for High-Throughput Analysis of Sarcosine and N-Ethylglycine in Human Urine and Urinary Sediments. *Anal. Chim. Acta* **2011**, *707*, 197–203.
- (835) Padró, J. M.; Marsón, M. E.; Mastrantonio, G. E.; Altchek, J.; García-Bourmissen, F.; Reta, M. Development of an Ionic Liquid-Based Dispersive Liquid-Liquid Microextraction Method for the Determination of Nifurtimox and Benznidazole in Human Plasma. *Talanta* **2013**, *107*, 95–102.
- (836) Abdelhamid, H. N.; Khan, M. S.; Wu, H. F. Design, Characterization and Applications of New Ionic Liquid Matrices for Multifunctional Analysis of Biomolecules: a Novel Strategy for Pathogenic Bacteria Biosensing. *Anal. Chim. Acta* **2014**, *823*, 51–60.
- (837) Arkan, E.; Saber, R.; Karimi, Z.; Mostafaie, A.; Shamsipur, M. Multiwall Carbon Nanotube-Ionic Liquid Electrode Modified with Gold Nanoparticles as a Base for Preparation of a Novel Impedimetric Immunosensor for Low Level Detection of Human Serum Albumin in Biological Fluids. *J. Pharm. Biomed. Anal.* **2014**, *92*, 74–81.
- (838) Salimi, A.; Kavosi, B.; Fathi, F.; Hallaj, R. Highly Sensitive Immunosensing of Prostate-Specific Antigen Based on Ionic Liquid-Carbon Nanotubes Modified Electrode: Application as Cancer Biomarker for Prostate Biopsies. *Biosens. Bioelectron.* **2013**, *42*, 439–446.
- (839) Zhong, Z.; Li, M.; Qing, Y.; Dai, N.; Guan, W.; Liang, W.; Wang, D. Signal-On Electrochemical Immunoassay for APE1 Using Ionic Liquid Doped Au Nanoparticle/Graphene as a Nanocarrier and Alkaline Phosphatase as Enhancer. *Analyst* **2014**, *139*, 6563–6568.
- (840) Daneshvar, L.; Rounaghi, G.; E'Shaghi, Z.; Chamsaz, M.; Tarahomi, S. Electrochemical Determination of Carbamazepin in the Presence of Paracetamol Using a Carbon Ionic Liquid Paste Electrode Modified with a Three-Dimensional Graphene/MWCNT Hybrid Composite Film. *J. Mol. Liq.* **2016**, *215*, 316–322.
- (841) Liu, N.; Chen, X.; Ma, Z. Ionic Liquid Functionalized Graphene/Au Nanocomposites and Its Application for Electrochemical Immunosensor. *Biosens. Bioelectron.* **2013**, *48*, 33–38.
- (842) Liu, N.; Ma, Z. Au-Ionic Liquid Functionalized Reduced Graphene Oxide Immunosensing Platform for Simultaneous Electrochemical Detection of Multiple Analytes. *Biosens. Bioelectron.* **2014**, *51*, 184–190.
- (843) Yu, X.; Wang, Y.; Chen, X.; Wu, K.; Chen, D.; Ma, M.; Huang, Z.; Wu, W.; Li, C. White-Light-Exciting, Layer-by-Layer-Assembled ZnCdHgSe Quantum Dots/Polymerized Ionic Liquid Hybrid Film for Highly Sensitive Photoelectrochemical Immunosensing of Neuron Specific Enolase. *Anal. Chem.* **2015**, *87*, 4237–4244.
- (844) Manning, G. S. Counterion Binding in Polyelectrolyte Theory. *Acc. Chem. Res.* **1979**, *12*, 443–449.
- (845) Clark, K. D.; Nacham, O.; Yu, H.; Li, T.; Yamsek, M. M.; Ronning, D. R.; Anderson, J. L. Extraction of DNA by Magnetic Ionic Liquids: Tunable Solvents for Rapid and Selective DNA Analysis. *Anal. Chem.* **2015**, *87*, 1552–1559.
- (846) Cheng, D. H.; Chen, X. W.; Wang, J. H.; Fang, Z. L. An Abnormal Resonance Light Scattering Arising from Ionic-Liquid/DNA/Ethidium Interactions. *Chem. - Eur. J.* **2007**, *13*, 4833–4839.
- (847) Freyer, J. L.; Brucks, S. D.; Gobieski, G. S.; Russell, S. T.; Yozwiak, C. E.; Sun, M.; Chen, Z.; Jiang, Y.; Bandar, J. S.; Stockwell, B. R.; et al. Clickable Poly(ionic liquids): A Materials Platform for Transfection. *Angew. Chem., Int. Ed.* **2016**, *55*, 12382–12386.
- (848) Cardoso, L.; Micaelo, N. M. DNA Molecular Solvation in Neat Ionic Liquids. *ChemPhysChem* **2011**, *12*, 275–277.
- (849) Chandran, A.; Ghoshdastidar, D.; Senapati, S. Groove Binding Mechanism of Ionic Liquids: a Key Factor in Long-Term Stability of DNA in Hydrated Ionic Liquids? *J. Am. Chem. Soc.* **2012**, *134*, 20330–20339.
- (850) Jumbri, K.; Abdul Rahman, M. B.; Abdulmalek, E.; Ahmad, H.; Micaelo, N. M. An Insight into Structure and Stability of DNA in Ionic Liquids from Molecular Dynamics Simulation and Experimental Studies. *Phys. Chem. Chem. Phys.* **2014**, *16*, 14036–14046.
- (851) Jumbri, K.; Ahmad, H.; Abdulmalek, E.; Abdul Rahman, M. B. Binding Energy and Biophysical Properties of Ionic Liquid-DNA Complex: Understanding the Role of Hydrophobic Interactions. *J. Mol. Liq.* **2016**, *223*, 1197–1203.
- (852) Tateishi-Karimata, H.; Nakano, M.; Sugimoto, N. Comparable Stability of Hoogsteen and Watson-Crick Base Pairs in Ionic Liquid Choline Dihydrogen Phosphate. *Sci. Rep.* **2014**, *4*, 3593.
- (853) Mazid, R. R.; Cooper, A.; Zhang, Y.; Vijayaraghavan, R.; MacFarlane, D. R.; Cortez-Jugo, C.; Cheng, W. Enhanced Enzymatic Degradation Resistance of Plasmid DNA in Ionic Liquids. *RSC Adv.* **2015**, *5*, 43839–43844.
- (854) Clark, K. D.; Sorensen, M.; Nacham, O.; Anderson, J. L. Preservation of DNA in Nuclease-Rich Samples Using Magnetic Ionic Liquids. *RSC Adv.* **2016**, *6*, 39846–39851.
- (855) Satpathi, S.; Sengupta, A.; Hridya, V. M.; Gavvala, K.; Koninti, R. K.; Roy, B.; Hazra, P. A Green Solvent Induced DNA Package. *Sci. Rep.* **2015**, *5*, 9137.
- (856) Liu, H.; Dong, Y.; Wu, J.; Chen, C.; Liu, D.; Zhang, Q.; Du, S. Evaluation of Interaction between Imidazolium-Based Chloride Ionic Liquids and Calf Thymus DNA. *Sci. Total Environ.* **2016**, *S66-S67*, 1–7.
- (857) Kim, K. S.; Tarakeshwar, P.; Lee, J. Y. Molecular Clusters of π -Systems: Theoretical Studies of Structures, Spectra, and Origin of Interaction Energies. *Chem. Rev.* **2000**, *100*, 4145–4186.
- (858) Li, S.; Cooper, V. R.; Thonhauser, T.; Lundqvist, B. I.; Langreth, D. C. Stacking Interactions and DNA Intercalation. *J. Phys. Chem. B* **2009**, *113*, 11166–11172.
- (859) Cooper, V. R.; Thonhauser, T.; Puzder, A.; Schroder, E.; Lundqvist, B. I.; Langreth, D. C. Stacking Interactions and the Twist of DNA. *J. Am. Chem. Soc.* **2008**, *130*, 1304–1308.
- (860) Park, K. Controlled Drug Delivery Systems: Past Forward and Future Back. *J. Controlled Release* **2014**, *190*, 3–8.
- (861) Tibbitt, M. W.; Dahlman, J. E.; Langer, R. Emerging Frontiers in Drug Delivery. *J. Am. Chem. Soc.* **2016**, *138*, 704–717.

(862) Nebert, D. W. Pharmacogenetics and Pharmacogenomics: Why is This Relevant to the Clinical Geneticist? *Clin. Genet.* **1999**, *56*, 247–258.

(863) Johnson, J. A. Pharmacogenetics: Potential for Individualized Drug Therapy through Genetics. *Trends Genet.* **2003**, *19*, 660–666.

(864) Kandpal, R. P.; Saviola, B.; Felton, J. The Era of 'Omics Unlimited. *BioTechniques* **2009**, *46*, 351–355.

(865) We define Ioliomics as a research discipline dealing with the studies of ions in liquids (or liquid phases) and stipulated with fundamental differences of ionic interactions (see [Figure 1](#) and other discussions in this Review).